



Roles of Reactive Oxygen Species in Rheumatoid Arthritis Pathogenesis

Su-Jin Yoo^{2,*}, Eunbyeol Go^{1,*}, Ye-Eun Kim¹, Sunyoung Lee², Jaeyul Kwon¹

Departments of ¹Medical Education and ²Internal Medicine, Chungnam National University School of Medicine, Daejeon, Korea

Rheumatoid arthritis (RA) is an autoimmune disease that starts with decreased tolerance to modified self-antigens and eventually leads to synovitis and destruction of bone and cartilage. Age is a risk factor for developing RA. Major changes in the immune system come with age due to chronic oxidative stress on the deoxyribonucleic acid (DNA) damage pathway, somatic mutation, modifications of auto-antigens, T cell tolerance and activation of fibroblast-like synoviocytes (FLS). Reactive oxygen species (ROS) generated by nicotinamide adenine dinucleotide phosphate oxidase 2 (NADPH oxidase 2) suppress T cell receptor signaling. Sirtuin 1 (SIRT1) is a critical immune suppressor of T cell activation and a key regulator of oxidative stress. When oxidative stress reduces activity of SIRT1, the breakdown of tolerance to modified self-antigens is expected. Generation of ROS can be perpetuated by enhanced DNA damage and dysfunctional mitochondria in a feedback loop during the development of RA. Through major T cell loss and selective proliferation of peripheral T cells, pro-inflammatory T cell pools with abnormal features are established in the T cell compartment. Hypoxic and inflammatory condition in synovium perpetuates ROS generation, which leads to the activation of FLS. In both T cell and synovium compartment, oxidative stress reshapes the immune system into the development of pre-clinical RA. **(J Rheum Dis 2016;23:340-347**)

Key Words. Rheumatoid arthritis, Reactive oxygen species, NADPH oxidase, Sirtuin 1, Oxidative stress

INTRODUCTION

Reactive oxygen species (ROS) include superoxide, hydrogen peroxide and hydroxyl radicals produced by the sequential reduction of oxygen. It is commonly thought that ROS are pro-inflammatory agents because inflammatory diseases have been linked to chronically elevated ROS production ("oxidative stress"). However, it is not clear what pathological processes are initiated or regulated by ROS in the immune system. Recently ROS have been studied as specific and critical regulators of immune system signaling [1,2]. ROS are generated from various sources including mitochondria and nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase, Nox). The discovery of a family of NADPH oxidases related to the phagocyte oxidase (Nox/Duox family) provides new opportunities to investigate the distinct roles of ROS generation by genetically manipulating these sources [3].

Rheumatoid arthritis (RA) is a long-lasting autoimmune disease that primarily affects joints due to the inflammation of the synovium and consequently causes damage to the cartilage and bones [4]. The etiology of RA is unclear. However, inflammatory cytokines are known to play important roles in the pathogenesis of RA to promote autoimmunity, chronic inflammation and tissue destruction. Oxidative stress has also been shown to be closely correlated with the pathogenesis of RA [5]. This review aims to provide an exploration of the possible roles of ROS generation in the immune systems involved in the initiation and development of RA.

The effects of ROS at different stages of RA develop-

Received : October 21, 2016, Revised : November 14, 2016, Accepted : November 15, 2016

pISSN: 2093-940X, eISSN: 2233-4718

Corresponding to: Jaeyul Kwon, Department of Medical Education, Chungnam National University School of Medicine, 266 Munhwa-ro, Jung-gu, Daejeon 35015, Korea. E-mail: kwonja@cnu.ac.kr

^{*}The first two authors contributed equally to this work.

Copyright © 2016 by The Korean College of Rheumatology. All rights reserved.

This is a Free Access article, which permits unrestricted non-commerical use, distribution, and reproduction in any medium, provided the original work is properly cited.

ment vary with the level and location of ROS production and the cell type or tissues involved [4]. ROS generally function as damaging or modifying agents of cellular components or as signaling molecules in an immune response. ROS are generated from several sources by stimulation of inflammatory cytokines such as tumor necrosis factor (TNF) or angiogenic factor such as vascular endothelial growth factor (VEGF) [6,7]. Mitochondria and NADPH oxidase are well studied sources of ROS [8]. Nox2 and Duox1 have been suggested to be critical [9,10] in T cell receptor (TCR) signaling of T cells and Nox2-dependent ROS generation was identified even as suppressor of arthritis [11]. Hypoxia-driven ROS generation is also very important in terms of RA pathogenesis because hypoxia develops even in the pre-clinical stage of synovitis and worsens the inflammation which in turn further promotes hypoxic conditions and creates a vicious cycle that may contribute to the establishment and progression of RA [12].

As aging progresses, biochemical imbalance between the formation and clearance of ROS generates a state referred to as "oxidative stress", leading to the damage of various cell components including proteins, lipids and deoxyribonucleic acid (DNA) [12]. Based on the concept that autoimmune diseases are a consequence of immune aging, age-related changes such as chronic oxidative and inflammatory stress are relevant to the initiation of RA [13-17]. Oxidative stress has already been shown to be involved in autoimmune responses. Surprisingly the p47phox subunit of Nox2 was first discovered as a protective factor in arthritis models, which suggested that Nox2-originated oxidative bursts suppressed autoimmune T cells [11,18,19]. ROS generation was proposed to regulate the expression of inflammatory cytokines and chemokines and to affect tissue damage in RA [20]. Excessive production of ROS may be critical for joint destruction and osteoclast activation [21,22]. ROS generation derived from the hypoxia-activated Nox2 is an initiating factor in angiogenesis for joint inflammation [23].

MAIN SUBJECTS

Effects of ROS in the pathogenesis of RA 1) Somatic mutation

Elevated ROS generation at the site of chronic inflammation causes somatic mutations [24]. Somatic mutations in the p53 gene have been observed in the RA synovium and cultured fibroblast-like synoviocytes (FLS) [25]. Many mutations produced by oxidative stress are present in the mitochondrial genome. A high frequency of mitochondrial somatic mutations was reported in synovial tissue of patients with RA and was strongly associated with low level of oxygen in the synovium as well as with high synovial lipid peroxidation [26].

2) Defect in DNA damage repair pathways

Impairments of DNA damage repair pathways increase the risk of RA in older people [27]. Naïve T cells in old people have more chance to accumulate genomic DNA damage than those in young people because these cells in older people have a relatively long life span in the periphery and are exposed to oxidative stress [28]. DNA damage such as DNA double-strand breaks needs to be detected and repaired by DNA damage repair pathways in order to maintain genomic stability. Increases in DNA-dependent protein kinases and deficiencies in ataxia telangiectasia mutated (ATM) and p53 in RAT cells have been shown to impair these repair pathways and lead to markedly increased DNA damage and apoptosis in naïve CD4⁺ T cells [27,29]. The significant loss of naïve T cells imposes lymphopenia-induced proliferation, leading to premature immunosenescence and possibly an autoimmune-biased T cell repertoire [17,27]. Dysfunctional T cells in patients with RA display the characteristics of inflammation-activated cells and sustain chronic inflammatory immune responses in the synovium [30].

3) Oxidative modification of auto-antigens

Oxidative stress-induced modifications in protein, lipid and DNA may have important roles in the pathogenesis of RA [31]. A strong correlation between levels of ROS and disease activity score with markers of oxidative damage was observed in patients with RA. Measurement of oxidatively modified proteins, lipids or DNA could serve as a biomarker for monitoring disease activity of RA [32]. Type II collagen oxidized by ROS (ROS-CII) were strongly detected in the serum and synovial fluid of patients with RA. 92.9% of sera from disease-modifying antirheumatic drug (DMARD)-naïve patients with early RA showed autoreactivity to ROS-CII [33]. Neo-epitopes can be generated by oxidative modification of proteins and be involved in autoimmune responses [34]. The immune system via pattern recognition receptors (PRRs) such as scavenger receptors, receptor of advanced glycation end products (RAGE) and toll-like receptor 4 (TLR4) can

sense neo-epitopes as pathogen- or danger-associated molecular patterns (PAMPs/DAMPs) [35].

Advanced glycation end products (AGEs) are accumulated by increased oxidative stress in aging and RA [36]. Advanced oxidation protein products (AOPPs) are accumulated in RA patients and are involved in various chronic inflammatory conditions through Nox-dependent ROS production [37]. As one of the receptors for AGEs and AOPPs, RAGE has been suggested as a risk factor for cardiovascular disease in RA patients [38]. Bone-targeting endogenous secretory RAGEs were shown to rescue RA in the murine collagen-induced arthritis (CIA) model [39].

4) Signaling role in T cell tolerance

Several types of ROS have been reported to be involved in T cell activation and differentiation in autoimmune responses. ROS generated from Nox2, Duox1 and mitochondria in T cells were reported to be relevant to these functions [9,10,40]. ROS from macrophages and other immune cells are also involved [41]. ROS have been shown to regulate autoimmune responses. Impairment of Nox2-dependent ROS generation in neutrophil cytosolic factor 1 (Ncf1)-mutated mice results in enhanced disease severity in several different animal models of arthritis [11,18,19]. Macrophage-restricted expression of functional Ncf1 restored arthritis resistance in a CIA model but not in a T cell-independent anti-collagen antibody-induced arthritis model. Restoration of Ncf1 in Ncf1-deficient mice suppressed T cell activation [41].

5) Regulation of T cell differentiation

Several types of ROS generation were reported to modulate differentiation of naïve CD4⁺ T cells. In both mouse and human models oxidative stress led the differentiation of the naïve CD4⁺ T cells towards Th2 phenotype [42,43]. In the absence of ROS T cells differentiated to Th1 type [9,41]. In addition, activation of naïve T cells from Nox2-deficient mice exhibited a skewed Th17 phenotype [19].

The immediate-early response gene X-1 (IEX-1, also known as IER3) is involved in preventing the production of ROS in mitochondria. Consequently the elevated generation of mitochondrial ROS from null mutation of IER3 facilitates the differentiation of Th17 cells and immunization with collagen lead to more severe arthritis in IER3 null mice than in wild-type mice. This finding indicates that mitochondrial alterations provide substantial con-

tributions to the dominant T cells [44].

Naïve CD4⁺ T cells from patients with RA have excess NADPH production. This leads to excessively reduced glutathione and reduced ROS generation [45]. ROS loss and ATM insufficiency in naïve CD4⁺ T cells from patients with RA skew T cell differentiation into interferon (IFN)- γ and interleukin (IL)-17 producing effector T cells. These biases are reversed by increasing intracellular ROS by treatment with menadione that generates intracellular ROS via redox cycle [45]. These observations indicate importance of ROS-based signal transduction in shaping T cell differentiation in RA.

Accelerated immune aging

Aging is characterized by increasing inflammatory and oxidative stress. The main feature of the aging process is a chronic progressive increase in the proinflammatory status described originally as inflamm-aging [13]. Based on the close relationship between oxidative stress, inflammation and aging, the oxidation-inflammatory theory of aging (oxi-inflamm-aging) was proposed [14]. RA, closely associated with aging, displays the characteristics described in the oxi-inflamm-aging [16,17]. During chronic oxidative and inflammatory stress oxidative modification of cellular components leads to the status described in the inflamm-aging and influences the homeostasis and health of the body. The relationship between the redox state and the function of immune cells influences the speed of aging and lifespan of the cells. At old age the body maintains a pro-inflammatory status and innate immune responses are actively induced more than adaptive immune response [15]. These pathways bring about a constant low level activation of granulocytes, macrophages and dendritic cells. The oxidative burst associated with an innate immune response upregulates ROS formation and reduced cellular antioxidant capacity. Overproduced oxidants react with membrane lipids and proteins and impair their function and create a circular loop of DAMP signaling activation. Moreover DAMPs can activate immune cells and their signaling pathway mediators such as nuclear factor kappa-light-chain-enhancer of activated B cells (NF- K B) and NADPH oxidase to further increase ROS production.

Extracellular DAMPs such as S100 calcium-binding protein A8/A9 (S100A8/A9) are well known to act as critical alarmins. They modulate the inflammatory response and interact with the PRR, TLR4 and RAGE to promote cell activation and recruitment [46]. S100A8/A9 was identified as a potential biomarker for monitoring disease activity of RA and has been tested successfully in localizing sites of sterile injury in pre-clinical imaging studies. Surprisingly the S100A8/A9 protein works as a partner for the cytosolic factors of NADPH oxidase activation in neutrophils [47]. Neutrophils and monocytes are recruited to sites of inflammation during infection or sterile injury.

Mitochondrial defects

Mitochondrial defects are a crucial component of the aging process and several age-related diseases. Mitochondrial disturbances lead to the deterioration of protein quality control and can especially contribute to the decline in autophagic degradation with aging [48]. Elevated production of ROS due to mitochondrial disturbances increases with aging and enhances signaling of DAMPs [49,50]. Expressions of certain genes related to the function of mitochondria were altered in patients with RA. A functional annotation study of RA and osteoarthritis (OA) by integrative genome-wide gene expression profiling analysis indicates that both RA and OA can be classified as mitochondrial disorders [51]. A five-fold increase in mitochondrial ROS production in whole blood and monocytes of patients with RA relative to that of healthy subjects suggests that oxidative stress is a pathogenic hallmark in RA [32].

SIRT1

Sirtuin (SIRT), a class III protein deacetylase, has been considered to be a longevity factor for its ability to combat oxidative stress and promote cellular survival. NF- κ B signaling is activated during aging [52] and is a potent inducer of the expression of several NADPH oxidase components including gp91phox and p22phox [53]. SIRT1 has also been suggested as a potent inhibitor of NF- κ B signaling by suppressing oxidative stress and inflammatory responses [54]. In response to oxidative stress SIRT1 induced antioxidant expression via forkhead box O (FoxO) pathways. SIRT1 can deacetylate FoxO factors (FoxO1, FoxO3a, and FoxO4) to stimulate the expression of antioxidants such as catalase, manganese superoxide dismutase (MnSOD) and thioredoxin and also potentiate SIRT1 expression via an auto-feedback loop [55]. Phosphatase and tensin homolog (PTEN) activated by SIRT1-dependent deacetylation activates FoxO transcription factors, which stimulate the expression of several antioxidants and SIRT1 as well as many autophagy

proteins. SIRT1 participates in the DNA damage repair process in an ATM-dependent way. The stress resistance was generally increased by these responses, which results in an extended life span [56].

Increased oxidative stress has been associated with the aging process and the expression and activity of SIRT1 was downregulated by chronic oxidative stress in inflammatory conditions [57]. For instance ROS can inhibit SIRT1 activity by evoking oxidative modifications on its cysteine residues. SIRT1 as a potent inducer of autophagy deacetylated Atg5, Atg7 and Atg8 proteins to stimulate autophagosome formation [58]. Furthermore FoxO1 and FoxO3 can act as downstream effectors of SIRT1 to promote autophagy [59], a process which declines with aging and is disturbed in several age-related diseases. Decreased activity of SIRT1 in aging leads to impairments in autophagy and subsequently enhances oxidative stress. A low-grade inflammatory phenotype was sustained in aging tissues because of impairments of autophagy. Consequently the deficiency in autophagy could enhance ROS and inflammatory responses in tissues and induce a state called inflamm-aging [60].

SIRT1 expression and activity was found to be decreased in RA patients and anti-citrullinated protein antibody (ACPA)-positive patients with RA showed lower SIRT1 activity relative to ACPA-negative patients with RA. The rate of apoptosis of peripheral blood mononuclear cells (PBMCs) in patients with RA was increased and negatively correlated with SIRT1 expression levels. SIRT1 is required to maintain T-cell tolerance [61]. And the lack of SIRT1 resulted in hyperacetylation of c-Jun and the breakdown of T cell tolerance [62]. Therefore, the decreased activity of SIRT1 in aged people and RA patients may result in the activation of autoimmune T cells.

Treatment with resveratrol reduced synovial hyperplasia, cartilage destruction, leukocyte infiltration, macrophage and T cell activation, and collagen-specific immunoglobulin levels in both CIA and lipopolysaccharides-induced acute inflammatory arthritis models [63,64]. Resveratrol-induced SIRT1 activation leads to the inhibition of RelA acetylation and a reduction in NF- κ B-induced expression of inflammatory factors such as TNF- α , IL-1 β , IL-6, matrix metalloproteinases (MMPs) such as MMP1 and MMP3, and cyclooxygenase 2, all of which have been implicated in the pathogenesis of RA. Similarly resveratrol-treated bone-derived cells showed reduced receptor activator of nuclear factor kappa-B ligand (RANKL)-induced NF- κ B acetylation and activa-

tion, as well as reduced osteoblastic activity associated with RA [65]. Resveratrol is also able to avoid excessive ROS induced lipid peroxidation and DNA damage.

Reshaping of peripheral naïve T cells

Although synovial inflammation-induced cartilage damage and destruction of bone is the dominant manifestation of clinical RA, systemic immune abnormalities that are not joint specific are already apparent many years before onset of the RA [4,66]. With advancing age, problems in the homeostasis of the T cell compartments and signaling thresholds for T cell activation lead to the loss of naïve T cells, the accumulation of inflammatory T cell populations and loss of tolerance to modified self-antigens [17,27].

The breakdown of tolerance to modified self-antigens-induced activation of self-reactive T cells could be driven by activation of DAMP signaling in addition to TCR signaling. DAMPs such as heat shock proteins and high mobility group box 1 (HMGB1) released from injured tissue can activate TLR4 and TLR2, respectively [46]. Similarly, increased production of ROS and post-translationally modified molecules such as oxidized lipoproteins activate the TLR8 and TLR2 pathway, respectively. TLR2, TLR4 and TLR8 activation will proceed to initiate an inflammatory response whose key mediators are IL-1, IL-6 and TNF- α . The increased expression of cytokines IL-1 β , IL-6, and TNF- α play key roles in the initiation of arthritis and pathogenesis of destructive arthritis in experimental animal models [67].

As thymic activity decreases around the age of 40 to 50 years, prolonged residence of naïve T cells in the periphery progressively lead to the accumulation of oxidative DNA damage [28]. A defect in the maintenance of genomic integrity with age causes excessive loss of peripheral T cells that needs to be compensated by homeostatic proliferation to maintain compartment size and leads to the eventual emergence of senescence biomarkers. During this enforced T-cell proliferation in periphery infrequent self-reactive T cells could be clonally expanded and lead to overaged and autoreactive T cells [27].

Naïve CD4⁺ T cells from patients with RA are metabolically reprogrammed, favoring NADPH production over adenosine triphosphate (ATP) generation [45]. Excessive NADPH supplies the cell with excessively reduced glutathione and depletes ROS. Such reductive stress fastens the cell cycle of T cells because they skip the G2/M cell cycle checkpoint due to insufficient ATM activation. ROS loss and ATM insufficiency promote T cell differentiation into Th1 and Th17 effector cells. p53 mRNA levels were significantly lower in PBMCs from patients with RA than from healthy controls. And PTEN expression down-regulated by p53 deficiency induced the activation of signal transducer and activator of transcription 3 (STAT3) [68].

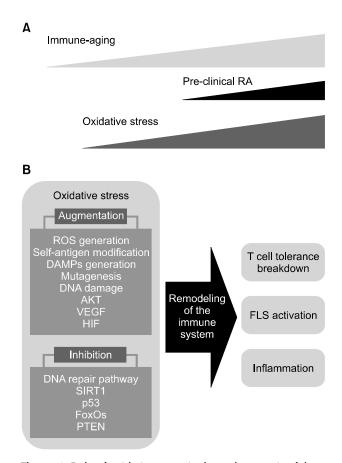


Figure 1. Role of oxidative stress in the pathogenesis of rheumatoid arthritis (RA). (A) With advancing age, oxidative stress plays critical roles in the generation of pre-clinical RA in the process of the immune-aging. (B) Important pathways regulated by the oxidative stress in the immune-aging process reshape the immune systems to lead breakdown of T cell tolerance, activation of fibroblast-like synoviocytes (FLS) and the generation of the inflammatory networks for pathogenesis of RA. Oxidative stress could contribute to development of RA in several ways. Enhanced generation of reactive oxygen species (ROS), oxidative modification of self-antigens, generation of danger-associated molecular patterns (DAMPs), mutagenesis of genomic deoxyribonucleic acid (DNA) and mitochondrial DNA, dysfunctional mitochondria, reduced autophagy, DNA damage, defects in DNA damage pathways, reduced activity of sirtuin 1 (SIRT1), p53, forkhead box O (FoxOs), and antioxidants, enhanced activity of AKT, activation of vascular endothelial growth factor (VEGF) and hypoxia-inducible factor (HIF) pathway and inhibition of phosphatase and tensin homolog (PTEN).

Loss of p53 exacerbated autoimmune arthritis and dysregulated the population of Th17 and regulatory T (T_{reg}) cells. The oxidative stress-dependent inhibition of PTEN may have similar effects on the T cell differentiation [69].

CONCLUSION

There are various immune reaction steps and cell types in which different type of ROS may have specific roles in the pathogenesis of RA. Here we focus on the possible roles of ROS in the development of pre-clinical RA (Figure 1). Chronic oxidative stress in old age could generate mutations in both genomic and mitochondrial DNA, leading to enhanced ROS generation in a feedback loop and an eventually remodeling of immune systems. ROS production may contribute the breakdown of T cell tolerance through several pathways. Nox2-generated ROS was identified as a negative regulator for Th17 differentiation and T cell activation. ROS from mitochondria on the other hand works in an opposite way. SIRT1 also has been shown to be a critical immune suppressor of both T cell and macrophage activation. SIRT1 activity which is down-regulated by oxidative stress may augment ROS generation through several signaling pathways involving NF- κ B, hypoxia-inducible factor (HIF), FoxOs or PTEN. With advancing age, FLS are activated in synovium and take tumor-like properties in which ROS generation is strongly involved. Several ROS-based signaling pathways appear to play critical roles in reshaping the compartment of T cells and synovium in the pre-clinical phase of RA.

ACKNOWLEDGMENTS

This work was supported by grants for Evaluation of effectiveness of alternative Herbal Medicine Resources (K16402) from Korea Institute of Oriental Medicine (KIOM); Korea Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry and Fisheries (IPET) through The study and dissemination of the standard evaluation system of immunity-boosting food, funded by Ministry of Agriculture, Food and Rural Affairs (MAFRA) (315064-3); research fund of Chungnam National University; Chungnam National University Hospital Research Fund, 2015.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

REFERENCES

- 1. Williams MS, Kwon J. T cell receptor stimulation, reactive oxygen species, and cell signaling. Free Radic Biol Med 2004;37:1144-51.
- Holmström KM, Finkel T. Cellular mechanisms and physiological consequences of redox-dependent signalling. Nat Rev Mol Cell Biol 2014;15:411-21.
- Lambeth JD, Neish AS. Nox enzymes and new thinking on reactive oxygen: a double-edged sword revisited. Annu Rev Pathol 2014;9:119-45.
- 4. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. N Engl J Med 2011;365:2205-19.
- Filippin LI, Vercelino R, Marroni NP, Xavier RM. Redox signalling and the inflammatory response in rheumatoid arthritis. Clin Exp Immunol 2008;152:415-22.
- Kim YS, Morgan MJ, Choksi S, Liu ZG. TNF-induced activation of the Nox1 NADPH oxidase and its role in the induction of necrotic cell death. Mol Cell 2007;26:675-87.
- 7. García-Quintans N, Prieto I, Sánchez-Ramos C, Luque A, Arza E, Olmos Y, et al. Regulation of endothelial dynamics by PGC-1 α relies on ROS control of VEGF-A signaling. Free Radic Biol Med 2016;93:41-51.
- Bae YS, Oh H, Rhee SG, Yoo YD. Regulation of reactive oxygen species generation in cell signaling. Mol Cells 2011; 32:491-509.
- Jackson SH, Devadas S, Kwon J, Pinto LA, Williams MS. T cells express a phagocyte-type NADPH oxidase that is activated after T cell receptor stimulation. Nat Immunol 2004;5:818-27.
- Kwon J, Shatynski KE, Chen H, Morand S, de Deken X, Miot F, et al. The nonphagocytic NADPH oxidase Duox1 mediates a positive feedback loop during T cell receptor signaling. Sci Signal 2010;3:ra59.
- 11. Olofsson P, Holmberg J, Tordsson J, Lu S, Akerström B, Holmdahl R. Positional identification of Ncf1 as a gene that regulates arthritis severity in rats. Nat Genet 2003;33: 25-32.
- Jeon CH, Ahn JK, Chai JY, Kim HJ, Bae EK, Park SH, et al. Hypoxia appears at pre-arthritic stage and shows co-localization with early synovial inflammation in collagen induced arthritis. Clin Exp Rheumatol 2008;26:646-8.
- Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. Ann N Y Acad Sci 2000; 908:244-54.
- Cannizzo ES, Clement CC, Sahu R, Follo C, Santambrogio L. Oxidative stress, inflamm-aging and immunosenescence. J Proteomics 2011;74:2313-23.
- Xia S, Zhang X, Zheng S, Khanabdali R, Kalionis B, Wu J, et al. An update on inflamm-aging: mechanisms, prevention, and treatment. J Immunol Res 2016;2016:8426874.
- Boren E, Gershwin ME. Inflamm-aging: autoimmunity, and the immune-risk phenotype. Autoimmun Rev 2004;3:401-6.

- Goronzy JJ, Shao L, Weyand CM. Immune aging and rheumatoid arthritis. Rheum Dis Clin North Am 2010;36: 297-310.
- Hultqvist M, Olofsson P, Holmberg J, Bäckström BT, Tordsson J, Holmdahl R. Enhanced autoimmunity, arthritis, and encephalomyelitis in mice with a reduced oxidative burst due to a mutation in the Ncfl gene. Proc Natl Acad Sci U S A 2004;101:12646-51.
- Lee K, Won HY, Bae MA, Hong JH, Hwang ES. Spontaneous and aging-dependent development of arthritis in NADPH oxidase 2 deficiency through altered differentiation of CD11b+ and Th/Treg cells. Proc Natl Acad Sci U S A 2011;108:9548-53.
- Shah D, Wanchu A, Bhatnagar A. Interaction between oxidative stress and chemokines: possible pathogenic role in systemic lupus erythematosus and rheumatoid arthritis. Immunobiology 2011;216:1010-7.
- 21. Henrotin YE, Bruckner P, Pujol JP. The role of reactive oxygen species in homeostasis and degradation of cartilage. Osteoarthritis Cartilage 2003;11:747-55.
- 22. Garrett IR, Boyce BF, Oreffo RO, Bonewald L, Poser J, Mundy GR. Oxygen-derived free radicals stimulate osteoclastic bone resorption in rodent bone in vitro and in vivo. J Clin Invest 1990;85:632-9.
- 23. Biniecka M, Connolly M, Gao W, Ng CT, Balogh E, Gogarty M, et al. Redox-mediated angiogenesis in the hypoxic joint of inflammatory arthritis. Arthritis Rheumatol 2014;66: 3300-10.
- 24. Tak PP, Zvaifler NJ, Green DR, Firestein GS. Rheumatoid arthritis and p53: how oxidative stress might alter the course of inflammatory diseases. Immunol Today 2000; 21:78-82.
- 25. Yamanishi Y, Boyle DL, Green DR, Keystone EC, Connor A, Zollman S, et al. p53 tumor suppressor gene mutations in fibroblast-like synoviocytes from erosion synovium and non-erosion synovium in rheumatoid arthritis. Arthritis Res Ther 2005;7:R12-8.
- Biniecka M, Fox E, Gao W, Ng CT, Veale DJ, Fearon U, et al. Hypoxia induces mitochondrial mutagenesis and dysfunction in inflammatory arthritis. Arthritis Rheum 2011;63: 2172-82.
- 27. Weyand CM, Fujii H, Shao L, Goronzy JJ. Rejuvenating the immune system in rheumatoid arthritis. Nat Rev Rheumatol 2009;5:583-8.
- Rane S, Das R, Ranganathan V, Prabhu S, Das A, Mattoo H, et al. Peripheral residence of naïve CD4 T cells induces MHC class II-dependent alterations in phenotype and function. BMC Biol 2014;12:106.
- 29. Shao L, Fujii H, Colmegna I, Oishi H, Goronzy JJ, Weyand CM. Deficiency of the DNA repair enzyme ATM in rheumatoid arthritis. J Exp Med 2009;206:1435-49.
- 30. Ponchel F, Morgan AW, Bingham SJ, Quinn M, Buch M, Verburg RJ, et al. Dysregulated lymphocyte proliferation and differentiation in patients with rheumatoid arthritis. Blood 2002;100:4550-6.
- 31. Ryan BJ, Nissim A, Winyard PG. Oxidative post-translational modifications and their involvement in the pathogenesis of autoimmune diseases. Redox Biol 2014;2:715-24.
- 32. Quiñonez-Flores CM, González-Chávez SA, Del Río Nájera D, Pacheco-Tena C. Oxidative stress relevance in the patho-

genesis of the rheumatoid arthritis: a systematic review. Biomed Res Int 2016;2016:6097417.

- 33. Strollo R, Ponchel F, Malmström V, Rizzo P, Bombardieri M, Wenham CY, et al. Autoantibodies to posttranslationally modified type II collagen as potential biomarkers for rheumatoid arthritis. Arthritis Rheum 2013;65:1702-12.
- 34. Eggleton P, Nissim A, Ryan BJ, Whiteman M, Winyard PG. Detection and isolation of human serum autoantibodies that recognize oxidatively modified autoantigens. Free Radic Biol Med 2013;57:79-91.
- 35. Matzinger P. The danger model: a renewed sense of self. Science 2002;296:301-5.
- 36. de Groot L, Hinkema H, Westra J, Smit AJ, Kallenberg CG, Bijl M, et al. Advanced glycation endproducts are increased in rheumatoid arthritis patients with controlled disease. Arthritis Res Ther 2011;13:R205.
- 37. Zheng S, Zhong ZM, Qin S, Chen GX, Wu Q, Zeng JH, et al. Advanced oxidation protein products induce inflammatory response in fibroblast-like synoviocytes through NADPH oxidase-dependent activation of NF- κ B. Cell Physiol Biochem 2013;32:972-85.
- 38. Steenvoorden MM, van der Helm-van Mil AH, Stoeken G, Bank RA, Devries RR, Huizinga TW, et al. The RAGE G82S polymorphism is not associated with rheumatoid arthritis independently of HLA-DRB1*0401. Rheumatology (Oxford) 2006;45:488-90.
- 39. Takahashi T, Katsuta S, Tamura Y, Nagase N, Suzuki K, Nomura M, et al. Bone-targeting endogenous secretory receptor for advanced glycation end products rescues rheumatoid arthritis. Mol Med 2013;19:183-94.
- 40. Sena LA, Li S, Jairaman A, Prakriya M, Ezponda T, Hildeman DA, et al. Mitochondria are required for antigen-specific T cell activation through reactive oxygen species signaling. Immunity 2013;38:225-36.
- 41. Gelderman KA, Hultqvist M, Pizzolla A, Zhao M, Nandakumar KS, Mattsson R, et al. Macrophages suppress T cell responses and arthritis development in mice by producing reactive oxygen species. J Clin Invest 2007;117:3020-8.
- 42. King MR, Ismail AS, Davis LS, Karp DR. Oxidative stress promotes polarization of human T cell differentiation toward a T helper 2 phenotype. J Immunol 2006;176:2765-72.
- 43. Peterson JD, Herzenberg LA, Vasquez K, Waltenbaugh C. Glutathione levels in antigen-presenting cells modulate Th1 versus Th2 response patterns. Proc Natl Acad Sci U S A 1998;95:3071-6.
- 44. Zhi L, Ustyugova IV, Chen X, Zhang Q, Wu MX. Enhanced Th17 differentiation and aggravated arthritis in IEX-1-deficient mice by mitochondrial reactive oxygen species-mediated signaling. J Immunol 2012;189:1639-47.
- 45. Yang Z, Shen Y, Oishi H, Matteson EL, Tian L, Goronzy JJ, et al. Restoring oxidant signaling suppresses proarthritogenic T cell effector functions in rheumatoid arthritis. Sci Transl Med 2016;8:331ra38.
- 46. Nefla M, Holzinger D, Berenbaum F, Jacques C. The danger from within: alarmins in arthritis. Nat Rev Rheumatol 2016;12:669-83.
- Kerkhoff C, Nacken W, Benedyk M, Dagher MC, Sopalla C, Doussiere J. The arachidonic acid-binding protein S100A8/ A9 promotes NADPH oxidase activation by interaction with p67phox and Rac-2. FASEB J 2005;19:467-9.
- 48. Green DR, Galluzzi L, Kroemer G. Mitochondria and the au-

tophagy-inflammation-cell death axis in organismal aging. Science 2011;333:1109-12.

- Tschopp J. Mitochondria: Sovereign of inflammation? Eur J Immunol 2011;41:1196-202.
- 50. Salminen A, Ojala J, Kaarniranta K, Kauppinen A. Mitochondrial dysfunction and oxidative stress activate inflammasomes: impact on the aging process and age-related diseases. Cell Mol Life Sci 2012;69:2999-3013.
- 51. Li ZC, Xiao J, Peng JL, Chen JW, Ma T, Cheng GQ, et al. Functional annotation of rheumatoid arthritis and osteoarthritis associated genes by integrative genome-wide gene expression profiling analysis. PLoS One 2014;9:e85784.
- Helenius M, Kyrylenko S, Vehviläinen P, Salminen A. Characterization of aging-associated up-regulation of constitutive nuclear factor-kappa B binding activity. Antioxid Redox Signal 2001;3:147-56.
- Anrather J, Racchumi G, Iadecola C. NF-kappaB regulates phagocytic NADPH oxidase by inducing the expression of gp91phox. J Biol Chem 2006;281:5657-67.
- Rajendran R, Garva R, Krstic-Demonacos M, Demonacos C. Sirtuins: molecular traffic lights in the crossroad of oxidative stress, chromatin remodeling, and transcription. J Biomed Biotechnol 2011;2011:368276.
- 55. Brunet A, Sweeney LB, Sturgill JF, Chua KF, Greer PL, Lin Y, et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. Science 2004;303:2011-5.
- 56. Salminen A, Kaarniranta K. SIRT1: regulation of longevity via autophagy Cell Signal 2009;21:1356-60.
- 57. Rajendrasozhan S, Yang SR, Kinnula VL, Rahman I. SIRT1, an antiinflammatory and antiaging protein, is decreased in lungs of patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2008;177:861-70.
- 58. Lee IH, Cao L, Mostoslavsky R, Lombard DB, Liu J, Bruns NE, et al. A role for the NAD-dependent deacetylase Sirt1 in the regulation of autophagy. Proc Natl Acad Sci U S A 2008;105:3374-9.
- 59. Sengupta A, Molkentin JD, Paik JH, DePinho RA, Yutzey KE. FoxO transcription factors promote cardiomyocyte survival upon induction of oxidative stress. J Biol Chem

2011;286:7468-78.

- Salminen A, Kaarniranta K, Kauppinen A. Inflammaging: disturbed interplay between autophagy and inflammasomes. Aging (Albany NY) 2012;4:166-75.
- 61. Niederer F, Ospelt C, Brentano F, Hottiger MO, Gay RE, Gay S, et al. SIRT1 overexpression in the rheumatoid arthritis synovium contributes to proinflammatory cytokine production and apoptosis resistance. Ann Rheum Dis 2011; 70:1866-73.
- Zhang J, Lee SM, Shannon S, Gao B, Chen W, Chen A, et al. The type III histone deacetylase Sirt1 is essential for maintenance of T cell tolerance in mice. J Clin Invest 2009;119: 3048-58.
- 63. Elmali N, Baysal O, Harma A, Esenkaya I, Mizrak B. Effects of resveratrol in inflammatory arthritis. Inflammation 2007;30:1-6.
- Xuzhu G, Komai-Koma M, Leung BP, Howe HS, McSharry C, McInnes IB, et al. Resveratrol modulates murine collagen-induced arthritis by inhibiting Th17 and B-cell function. Ann Rheum Dis 2012;71:129-35.
- 65. Shakibaei M, Buhrmann C, Mobasheri A. Resveratrol-mediated SIRT-1 interactions with p300 modulate receptor activator of NF-kappaB ligand (RANKL) activation of NFkappaB signaling and inhibit osteoclastogenesis in bone-derived cells. J Biol Chem 2011;286:11492-505.
- 66. Kokkonen H, Söderström I, Rocklöv J, Hallmans G, Lejon K, Rantapää Dahlqvist S. Up-regulation of cytokines and chemokines predates the onset of rheumatoid arthritis. Arthritis Rheum 2010;62:383-91.
- 67. van den Berg WB. Lessons from animal models of arthritis over the past decade. Arthritis Res Ther 2009;11:250.
- Lee SH, Park JS, Byun JK, Jhun J, Jung K, Seo HB, et al. PTEN ameliorates autoimmune arthritis through down-regulating STAT3 activation with reciprocal balance of Th17 and Tregs. Sci Rep 2016;6:34617.
- 69. Kwon J, Lee SR, Yang KS, Ahn Y, Kim YJ, Stadtman ER, et al. Reversible oxidation and inactivation of the tumor suppressor PTEN in cells stimulated with peptide growth factors. Proc Natl Acad Sci U S A 2004;101:16419-24.