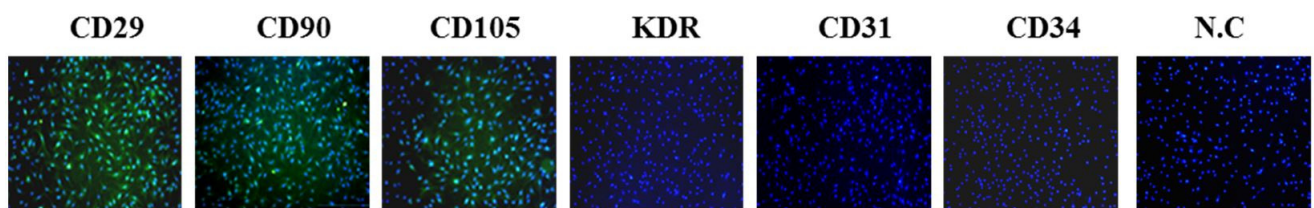


## Supporting Information



**Fig. S1.** Immunofluorescence staining of ASC. ASC were stained with CD29, CD90 and CD105 for mesenchymal stem cell identification, with KDR, CD31 and CD34 for endothelial lineage cell identification.

**Table S1.** Photobiomodulation irradiation

	<i>In vitro</i> study
Frequency of irradiation	10 min during 3 days
Light irradiated site	24 well cell culture plate (12.5×8.5 cm)
Distance from the LED light	10 cm
Power density	10 mW/cm <sup>2</sup>
Light dosage (fluence)	6 J/cm <sup>2</sup>
Irradiance (wavelength)	660 nm

**Table S2.** List of antibodies for immunofluorescence staining

Antibody	Host	Company	Catalogue number
anti-human CD29	mouse	Millipore	MAB2253Z
anti-human Flk-1	mouse	Santa Cruz	Sc-6251
anti-human CD34	mouse	Millipore	MAB4211
anti-human CD31	mouse	Dako	M0823
anti-mouse CD31	rabbit	BD biosciences	550274
anti-human CD90	mouse	BD biosciences	555595
anti-human CD105	mouse	Caltac Laboratories	MHCD10500
Alpha-Smooth Muscle Actin Monoclonal	mouse	Invitrogen	14-9760-82
HIF-1 alpha	rabbit	Novus	NB100-134
anti-human FGF	rabbit	abcam	ab8880
anti-human VEGF	rabbit	abcam	ab52917
anti-human HGF	rabbit	Santa Cruz	Sc-13087
Alexa Fluor 488 anti-mouse IgG	goat	Invitrogen	A11001
Alexa Fluor 594 anti-rabbit IgG	goat	Invitrogen	A11012

**Table S3.** Histological scoring system

Scores	Re-epithelialization	Dermal regeneration	Granulation tissue formation	Angiogenesis
1	Minimal epidermal regeneration (<50%)	No skin appendage formation	Thin granulation around wound edges only	Little angiogenesis (<10 vessels/HPF)
2	Moderate epidermal regeneration (50%)	A few skin appendage formation (<5 appendages/wound area)	Moderate granulation in the wound bed	Moderate angiogenesis (10~20 vessels/HPF)
3	Complete epidermal regeneration (100%)	Considerable skin appendage formation (>5 appendages/wound area)	Thick granulation in 100% of the wound bed	Marked newly formed and well-structured capillary vessels (>20 vessels/HPF)

High-power field (HPF); original magnification  $\times 400$ .

## Supporting Materials and methods

### Preparation of chitosan-coated culture plates

Chitosan was coated on tissue culture plates using methods described in a previous study, with some modifications (8). Briefly, 0.5 ml 1% (w/w) chitosan solution (C-3646, Sigma, St. Louis, MO) dissolved in 0.67% (w/v) acetic acid was added into each well of 24-well tissue culture plates (TCPS; Greiner bio-one, Frickenhausen, Germany) and dried in an oven at 60°C for 24 h to form a thin film, after which it was neutralized by 0.5 N NaOH aqueous solution (Sigma) for 2 h. Next, the plates were washed thoroughly with distilled water before being exposed to ultraviolet light overnight.