

# Peripheral Neuron-Organoid Interaction Induces Colonic Epithelial Differentiation via Non-Synaptic Substance P Secretion

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**Background and Objectives:** The colonic epithelial layer is a complex structure consisting of multiple cell types that regulate various aspects of colonic physiology, yet the mechanisms underlying epithelial cell differentiation during development remain unclear. Organoids have emerged as a promising model for investigating organogenesis, but achieving organ-like cell configurations within colonic organoids is challenging. Here, we investigated the biological significance of peripheral neurons in the formation of colonic organoids.

**Methods and Results:** Colonic organoids were co-cultured with human embryonic stem cell (hESC)-derived peripheral neurons, resulting in the morphological maturation of columnar epithelial cells, as well as the presence of enterochromaffin cells. Substance P released from immature peripheral neurons played a critical role in the development of colonic epithelial cells. These findings highlight the vital role of inter-organ interactions in organoid development and provide insights into colonic epithelial cell differentiation mechanisms.

**Conclusions:** Our results suggest that the peripheral nervous system may have a significant role in the development of colonic epithelial cells, which could have important implications for future studies of organogenesis and disease modeling.

**Keywords:** Organogenesis, Colonic organoid, Gut development, Peripheral neuron, Inter-organ crosstalk, Pluripotent stem cell

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## Introduction

The colonic epithelial layer is a complex and dynamic structure that consists of multiple cell types, including goblet cells, enterocytes, Paneth cells, enteroendocrine cells, and intestinal stem cells (1). Each of these cells plays a crucial role in regulating various aspects of colonic physiology, such as mucosal formation, colonic absorption, colonic motility, immunity, and regeneration (2). Although these cells are derived from colonic stem cells (3), the mechanisms underlying their diversification during development are not yet fully understood.

Organoids, which are three-dimensional structures that recapitulate the process of organ formation *in vitro*, have emerged as a promising research model for investigating

organogenesis and understanding the mechanisms underlying cell diversification (4). However, achieving organ-like cell configurations within colonic organoids remains a significant challenge (5). Since various types of cells arise as the end product of the developmental process, simulating the developmental process during the organoid establishment is recognized as a necessary condition. Such simulations require appropriate intrinsic and extrinsic signaling stimuli, and recent research has shown that interactions between cells, tissues, or organs also play an important role. One of the most significant examples of inter-organ interaction is the connection between nerves and target organs (6-8). Peripheral nerves not only accept and deliver environmental information and commands but also contribute to the maintenance of structural and functional homeostasis and the development of the target organ to which they connect (9-11). In the case of the large intestine, peripheral nerve connections are observed from the early stages of organ formation (12), however, the precise role of the nervous system in colonic development remains unclear.

The aim of this study was to evaluate the biological significance of peripheral neuron contributions in the formation of colonic organoids. We co-cultured colonic organoids with human embryonic stem cell (hESC)-derived peripheral neurons following induction of colonic specification through temporarily restricted stimulation of bone morphogenetic protein (BMP). Our investigation revealed that substance P (SP) spontaneously released from immature peripheral neurons plays a critical role in the development of colonic epithelial cells, specifically colon enterochromaffin cells. Our findings highlight the essential role of inter-organ interactions in organoid development and provide insights into the mechanisms underlying colonic epithelial cell differentiation.

## Materials and Methods

### Human embryonic stem cells (hESCs)

The hESC line WA09 (H9) was obtained from WiCell. The cells were maintained in Essential 8 medium (Thermo Fisher, USA) on iMatrix-511 coating material (Matrixome, Japan) under a 5% CO<sub>2</sub> condition in a humidified incubator. To prevent any unintended effects of genetic mutations, hESCs between passages 30 to 40 were collected using either Versene solution (Thermo Fisher, USA) or Accutase (Sigma Aldrich, USA) for further experiments. All experiments involving hESCs were conducted under institutional IRB regulation KHSIRB-20-489 approved by Kyung Hee University (13).

### Generation of colonic organoids

To generate colonic organoids, we modified previously reported protocols (14-16). Briefly, 2.4 million hESCs were plated in a 24-well plate and allowed to reach 70% confluency as day 0. From day 0 to day 3, definitive endoderm (DE) was induced using RPMI 1640 medium (Welgene, Korea) containing 2% fetal bovine serum (FBS) (Welgene, Korea), 3  $\mu$ M CHIR99021 (Sigma Aldrich, USA), and 100 ng/ml Activin A (Peprotech, USA). From day 4 to day 7, hindgut cells were differentiated using DMEM/F-12 medium (Welgene, Korea) containing 2% FBS, 1% B27 (Thermo Fisher, USA), 3  $\mu$ M CHIR99021, and 500 ng/ml fibroblast growth factor 4 (FGF4) (Peprotech, USA), and then the resulting cell aggregates were embedded in Geltrex (Thermo Fisher, USA) after detachment using dispase (Thermo Fisher, USA). From day 8, DMEM/F-12 medium containing 1% B-27, 3  $\mu$ M CHIR99021, 100 nM LDN193189 (STEMGENT, USA), and 100 ng/ml epidermal growth factor (EGF) (Peprotech, USA) was applied continuously, and from day 14 to day 20, 20 ng/ml BMP4 (Peprotech, USA) was temporarily added to the same medium composition to generate colonic organoids.

### Differentiation of neural crest stem cell (NCSC) and peripheral neuron

To differentiate NCSCs from PHOX2B::eGFP reporter hESCs, colonies were dissociated into single cells using Accutase (Sigma Aldrich) and plated on Geltrex (Thermo Fisher, USA), allowing them to reach 70%~80% confluence following previous protocols (17). Differentiation into NCSCs was achieved by gradually changing Essential 8 medium to Neurobasal medium (Thermo Fisher, USA) supplemented with B27 and N-2 (Thermo Fisher, USA), containing 500 nM LDN193189 and 10  $\mu$ M SB431542 for the first three days, followed by the addition of 3  $\mu$ M CHIR99021 for an additional eight days. FACS sorting was used to isolate NCSCs expressing surface antigen p75 nerve growth factor receptor (NGFR) and human natural killer 1 (HNK-1). Enteric NCSCs were differentiated using a previously reported protocol as a control (18). After reaching 70%~80% confluence on Geltrex, cells were treated with 500 nM LDN193189 and 10  $\mu$ M SB431542 in gradually changing Neurobasal medium from day 0 to day 3. From day 4 to day 6, cells were treated with 3  $\mu$ M CHIR99021, SB431542, 10  $\mu$ M DAPT (Cayman Chemical, USA), and 0.2  $\mu$ M PD173074 (Sigma Aldrich, USA). From day 7 to day 12, cells were treated with 100 ng/ml sonic hedgehog (SHH) (Peprotech, USA) and 1  $\mu$ M purlmorphamine (PMP) (Sigma Aldrich). From day 10 to 14, cells were treated with 10  $\mu$ g/ml BMP4. On day 14,

PHOX2B::eGFP-expressing cells were sorted by FACS. Peripheral neurons were further differentiated by adding 10 nM nerve growth factor (NGF) (Peprotech, USA), 20 nM brain derived neurotrophic factor (BDNF) (Peprotech, USA), 20 nM glial cell derived neurotrophic factor (GDNF) (Peprotech, USA), 100  $\mu$ M dibutyryl cyclic adenosine monophosphate (dbcAMP) (Sigma Aldrich, USA), and 100  $\mu$ M sodium L-ascorbate (Sigma Aldrich, USA) to the culture medium, and cells were cultured for an additional seven weeks (13).

### Co-culture of colonic organoids with NCSCs

FACS-sorted NGFR and HNK1 double-positive NCSCs (250,000 cells/well) were cultured on Matrigel-coated plates in Neurobasal medium supplemented with B27 and N-2, 20 ng/ml bFGF, and 20 ng/ml EGF for 24 hours. After washing the colonic organoids embedded in Matrigel with cold PBS, they were separated from the matrix using Cell Recovery Solution (Corning, USA). Approximately 50 organoids per well were allowed to naturally settle onto the NCSCs. The co-culture medium for colonic organoids and NCSCs was a 1 : 1 mixture of Neurobasal medium containing B27 and N-2 supplements and DMEM/F-12 medium. Half of the medium was changed every other day with fresh medium. To establish co-culture without direct contact between colonic organoids and NCSCs, NGFR and HNK1 double-positive NCSCs and colonic organoids were cultured separately on Matrigel-coated plates for 24 hours in B27 and N-2-supplemented Neurobasal medium containing 20 ng/ml EGF and 20 ng/ml bFGF or DMEM/F-12 medium. A transwell system was used, where the colonic organoids were cultured in the lower chamber and the NCSCs in the upper chamber, with the two chambers separated by a 0.4  $\mu$ m pore membrane. The culture medium was a 1 : 1 mixture of Neurobasal medium and DMEM/F-12, and half of the medium was changed every other day with a fresh one.

### Flow cytometry and fluorescence-activated cell sorting (FACS)

For flow cytometry analysis, the cells were mechanically dissociated after exposure to Accutase and treated with 40  $\mu$ g/ml DNase I (Roche Applied Science, Germany) for 20 minutes at 37°C. Next, the cells were incubated with appropriate antibodies for 1 hour at 4°C, washed twice with Phosphate Buffered Saline (DPBS), and treated with 7-AAD (BD Pharmingen, USA) to eliminate dead cells, following the manufacturer's instructions. The cells were analyzed using an FC500 (Beckman Coulter, USA) or isolated using FACS.

### Immunofluorescence staining

Cells and cryosectioned organoid tissues were fixed with 4% paraformaldehyde and permeabilized with PBS containing 0.5% Triton X-100. After blocking with 5% normal goat serum (Jackson Laboratory, USA) in PBST, the cells were incubated overnight at 4°C with primary antibodies. The primary antibodies were diluted in PBS with 0.1% normal serum. The cells were then incubated with secondary antibodies at a 1 : 500 dilution in PBS for 1 hour at room temperature. Finally, the slides were mounted using a mounting medium with DAPI (Vector labs, USA) and imaged using fluorescence microscopy (Olympus Corporation, Japan). To eliminate any background fluorescence, negative controls without primary antibodies were included in the experiment. Antibodies used in the research are listed in Table 1.

### Quantitative real-time polymerase reaction (qRT-PCR)

Total RNA was extracted using Trizol (Invitrogen), RNA concentration was measured spectrophotometrically using a Nanodrop (DeNvix Inc., USA). Subsequently, 2  $\mu$ g of total RNA was reverse transcribed to cDNA using High-Capacity cDNA Reverse Transcription Kit (Thermo Fisher, USA). The quantitative PCR was performed on the StepOne Plus Real-Time PCR system (Applied Biosystems, USA) using AccuPower GreenSrat qPCR PreMix (Bioneer, Korea). Relative mRNA levels were normalized to those of 18S for each gene. The primer sequences are listed in Table 1.

### Western blotting

Proteins were separated by SDS-PAGE and transferred onto nitrocellulose (NC) membranes (Bio-Rad, USA) for immunoblotting. Immunoblotting was performed with indicated primary antibodies (Table 1) and relevant secondary antibodies (Thermo Fisher, USA). Immunoblot bands were visualized with chemiluminescence (Pierce, USA) and developed manually using X-ray film (Kodak, USA).

### Statistical analysis and software

All data were analyzed using GraphPad Prism 9.0 software (GraphPad, USA). The data were presented as mean  $\pm$  standard deviation (SD) of at least three independent biological experiments. Morphological images representative of at least three biologically independent experiments with similar results were captured. The statistical significance was evaluated using paired t-test, unpaired t-tests, one-way ANOVA, or two-way ANOVA as appropriate. A p-value less than 0.05 was considered statistically significant. The bands' densitometric analysis for immunoblotting was conducted using FIJI software (Image J, USA). FlowJo 10.0

**Table 1.** List of primers and antibodies

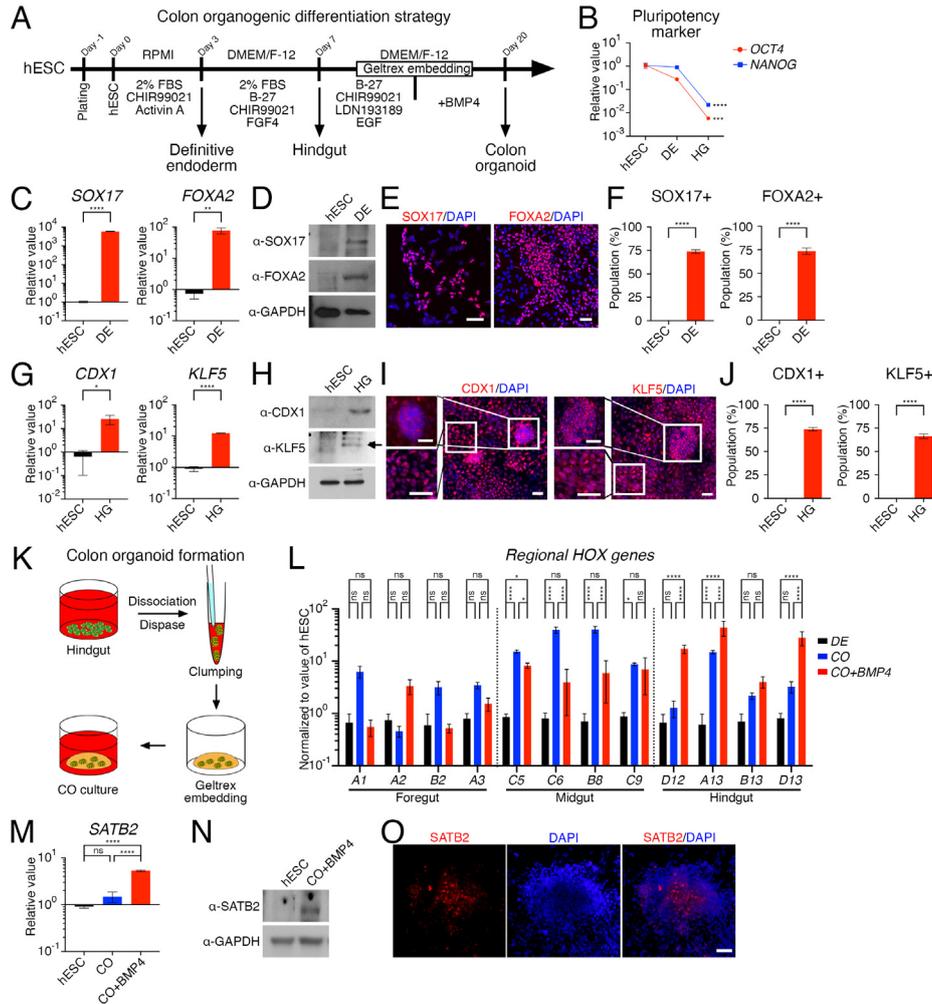
qPCR primers		
Gene	Forward	Reverse
<i>OCT4</i>	GGGGTTCTATTGGGAAGGTAT	TGTTGTCAGCTTCCTCCACC
<i>NANOG</i>	CAGAAGGCCTCAGCACCTAC	ATTGGAAGGTTCCCAGTCGG
<i>SOX17</i>	CAGGCTGCAGCGCCATGAGCAGCCCG	CTGGGGCGGATCCGGGACCTGTCACAC
<i>FOXA2</i>	CTTCAAGCACCTGCAGATTC	AGACCTGGATTCACCGTGT
<i>CDX1</i>	AGGACAAGTACCGCGTGGTCTA	CCTCTGAACGTATGGAGGAGGA
<i>KLF5</i>	GGTTGCACAAAAGTTTATAC	GGCTTGGCGCCCGTGTGCTTCC
<i>HOXA1</i>	TCCTGGAATACCCCATACTTAGCA	GCCGCCGCAACTGTTG
<i>HOXA2</i>	ACAGCGAAGGGAAATGTAAAAGC	GGGCCCCAGAGACGCTAA
<i>HOXB2</i>	TCCTTGGCCGTCTACTGGAA	AGTGGATTAACGCTAATTCAGTAATAC
<i>HOXA3</i>	TGCAAAAAGCGACCTACTACGA	CGTCGGCGCCCAAG
<i>HOXC5</i>	AGGTGCAGGCATCCAGGTACT	GGGTTGGCAGCCATGTCTAC
<i>HOXC6</i>	TTACCCCTGGATGCAGCGAATG	CCGCGTTAGGTAGCGATTGAAG
<i>HOXB8</i>	GTCGCCCACACAGCTCTTCCC	AATAGGAACCTCTTCCAGCTC
<i>BOXC9</i>	CAGCAAGCACAAAAGAGGAGAAGG	AGTTCCAGCGTCTGGTACTTGG
<i>HOXD12</i>	TGTGTGAGCGCAGTCTCTACAGA	CGGCCTCAGGTTGGAGAAG
<i>HOXA13</i>	AAATGTAAGTCCCAAGAGCA	ATCCGAGGGATGGGAGACC
<i>HOXB13</i>	CCACTGGCTGCTGGACTGTT	TATGACTGGGCCAGGTTCTTTG
<i>HOXD13</i>	CTGGGCTACGGCTACCACTTC	GCGATGACTTGAGCGCATT
<i>SATB2</i>	CCACCTTCCCAGCTTGATT	TTAGCCAGCTGGTGGAGACT
<i>PHOX2B</i>	CCAGTGCCAGCCCAATAGAC	TGGACAATAGCAAAGCGGTTG
<i>NDRG4</i>	ATGCTTTCATCCACTCACC	TTCACTGCTCTCTCCCGTTT
<i>LMX1A</i>	TCAGAAGGGTGATGAGTTGTCC	GGGGCGCTTATGGTCTTG
<i>CHGA</i>	CGGTTTTGAAGATGAACTCTCAG	GCTCTTCCACCGCTCTT
<i>NK1R</i>	GCCTGTTCTACTGCAAGTCCAC	CACAGATGACCACTTTGGTGGC
<i>NK3R</i>	GCCAAGAGAAAGGTTGTGAAGA	GTGTACATGCTGCTCTGGCG
<i>18S rRNA</i>	GGCCCTGTAATTGGAATGAG	GGCCCTGTAATTGGAATGAG

Antibodies		
Target	Company	Application
SOX17	R&D system, AF1924	Western blotting, Immunofluorescence
FOXA2	Santa Cruz, sc-374376	Western blotting, Immunofluorescence
CDX1	Santa Cruz, sc-398470	Western blotting, Immunofluorescence
KLF5	NOVUS, NBP1-49538	Western blotting, Immunofluorescence
SATB2	Santa Cruz, sc-81376	Western blotting, Immunofluorescence
MUC2	Santa Cruz, sc-515032	Immunofluorescence
MUC3	Santa Cruz, sc-7315	Western blotting, Immunofluorescence
MUC4	Santa Cruz, sc-33654	Western blotting, Immunofluorescence
EpCAM	eBioscience, 14-5791-81	FACS, Immunofluorescence
CD133	Santa Cruz, sc-365537	Immunofluorescence
Villin	Santa Cruz, sc-58897	Western blotting, Immunofluorescence
DEFA5	NOVUS, NB110-60002	Western blotting
CHGA	NOVUS, NBP2-29428	Western blotting, Immunofluorescence
LGR5	Abcam, ab75732	Western blotting
TUJ1	Biologend, 802001	Immunofluorescence
KI-67	eBioscience, 14-5698-82	Immunofluorescence
SOX9	Cell signaling, 82630	Immunofluorescence
Serotonin	Immunostar, 20080	Immunofluorescence
P75 NGFR	Santa Cruz, sc-13577	FACS
HNK-1	Sigma Aldrich, C6680	FACS
GAPDH	Santa Cruz, sc-47724	Western blotting
Substance P	Abcam, ab288318	ELISA
CHGA	R&D systems, DY9098-05	ELISA

Table 1. Continued

Target	Company	Inhibitors	Application
Vapreotide	MCE, HY-P3802		NK1R antagonist
Spantide II	MCE, HY-P1722		NK1R antagonist



**Fig. 1.** Stepwise protocol for generating colonic organoids. (A) Schematic diagram showing the modification of previously reported protocols for colonic organoids, with or without bone morphogenetic protein (BMP) signaling stimulation. (B) Decrease in pluripotency markers *OCT4* and *NANOG* during differentiation process. (C) Expression level of mRNA for *SOX17* and *FOXA2* on day 3 of differentiation towards definitive endoderm (DE). (D) Immunoblot images of protein expression for *SOX17* and *FOXA2* on the differentiation stage of DE. GAPDH was used as an internal control. (E) Cells expressing a protein for *SOX17* and *FOXA2* on the DE stage. (F) Counting results for *SOX17* and *FOXA2* expressing cells from (E). (G) Expression level of mRNA for *CDX1* and *KLF5* on day 7 of differentiation towards hindgut (HG) cells. (H) Immunoblot images of protein expression for *CDX1* and *KLF5* on the differentiation stage of hindgut. GAPDH was used as an internal control. (I) Cells expressing a protein for *CDX1* and *KLF5* on the hindgut stage. (J) Counting results for *CDX1* and *KLF5* expressing cells from (I). (K) Schematic diagram showing the embedding process of hindgut cells that formed clusters into an artificial ECM matrix in clump form. (L) BMP4 stimulation during the embedding stage increases expression of hindgut *HOX* genes responsible for posterior development. (M and N) Increase in mRNA and protein expression of colon-restricted chromatin factor *SATB2* with BMP4 application. (O) Identification of a cell cluster expressing the *SATB2* protein in the inner center of the colonic organoid. Data expressed as mean  $\pm$  SD (n=9, biological repeat). p-values calculated by one-way ANOVA (B, L, M) or unpaired t-test (C, F, G, and J). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001, ns=not significant. All scale bars=50  $\mu$ m. DE: definitive endoderm, HG: hindgut, CO: colonic organoid.

software (Treestar, USA) was utilized to analyze the FACS data. The illustrations were generated using images obtained from biorender.com.

## Results

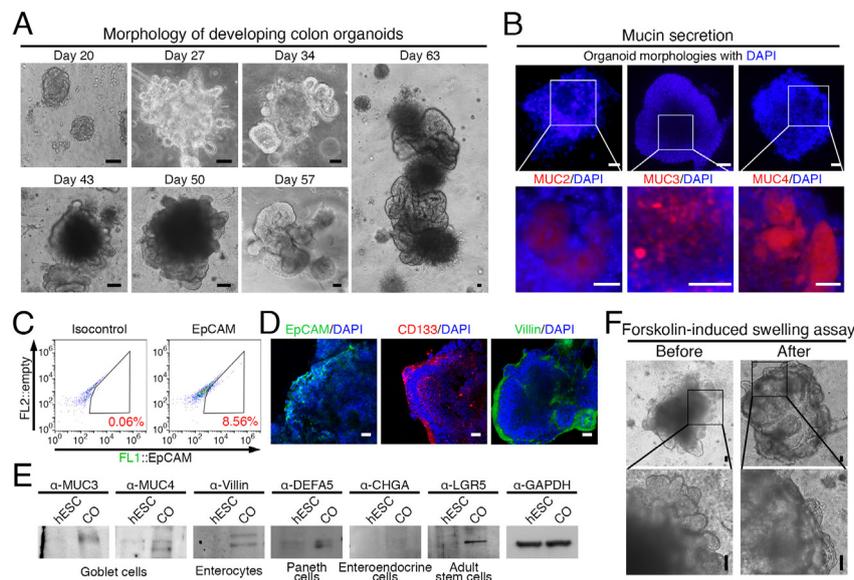
### Temporal regulation of BMP signaling modulates hindgut and colonic organoid differentiation

We developed a stepwise protocol for generating colon organoids by modifying previously reported protocols for colonic organoids, which either stimulated BMP signals (14) or did not (15) (Fig. 1A). The differentiation process was characterized by a decrease in the pluripotency markers *OCT4* and *NANOG* (Fig. 1B). By day 3 of differentiation towards DE, cells expressed both mRNA and protein for *SOX17* and *FOXA2* (Fig. 1C and 1D), with over 70% of cells expressing each protein (Fig. 1E and 1F). With additional differentiation, cells on day 7 expressed mRNA and protein for *CDX1* and *KLF5* (Fig. 1G and 1H) and differentiated into hindgut cells that formed cell clusters (Fig. 1I). The differentiation rate at this stage was similar to that of DE differentiation, with over 70% of cells expressing each protein (Fig. 1J). During the process of inducing the development of colonic organoids by em-

bedding hindgut cells that formed clusters into an artificial ECM matrix in clump form (Fig. 1K), the addition of BMP4 in the latter stages of embedding significantly increased the expression of hindgut hox genes, *HOXD12*, *HOXA13*, *HOXB13*, and *HOXD13*, which are responsible for posterior development, compared to hox gene families responsible for development of the foregut or midgut (Fig. 1L). The application of BMP4 to the colonic organoids also resulted in an increase in mRNA and protein expression of special AT-rich sequence-binding protein 2 (SATB2), a colon-restricted chromatin factor of the large intestine in the posterior region (Fig. 1M and 1N) (19). Additionally, a cell cluster expressing the SATB2 protein was identified in the inner center of the colonic organoid (Fig. 1O).

### Colonic organoids generated by BMP treatment exhibit functional epithelial cells but lack enteroendocrine cells

After long-term culture, colonic organoids embedded in the ECM matrix exhibited an increase in volume as the ECM matrix was gradually removed (Fig. 2A). At week 6, Mucin proteins (MUC), detectable by MUC2, MUC3, and MUC4 antibodies, accumulated within the organoids (Fig.



**Fig. 2.** Characterization of long-term cultured colonic organoids embedded in an ECM matrix. (A) Morphological changes of colonic organoids during long-term culture. (B) Detection of Mucin proteins (MUC) within the organoids at week 6 using MUC2, MUC3, and MUC4 antibodies. (C) Representative FACS plot for EpCAM-expressing cells of colonic organoids. (D) Immunostaining confirmed the formation of an epithelial layer expressing EpCAM, CD133, or Villin on the outer layer of the organoids. (E) Detection of marker proteins representing various cell types within the epithelial layer, including goblet cells (MUC3, MUC4), enterocytes (Villin), Paneth cells (DEFA5), enteroendocrine cells (CHGA), and colonic stem cells (LGR5). (F) Swelling assay confirming the presence of functional epithelial cells in the organoids treated after forskolin. All scale bars=50  $\mu$ m. CO: colonic organoid.

2B), indicating the differentiation of epithelial cells, including goblet cells. Flow cytometry analysis of EpCAM-expressing cells revealed that approximately 8% of the cells were epithelial (Fig. 2C). Immunostaining confirmed the formation of an epithelial layer expressing EpCAM, CD133, or Villin on the outer layer of the organoids (Fig. 2D). To identify the various cell types within the epithelial layer, markers representing each cell type were examined. MUC3, MUC4, Villin, defensin alpha 5 (DEFA5), and leucine rich repeat containing G protein-coupled receptor 5 (LGR5) expression were detected (Fig. 2E), indicating the presence of goblet cells, enterocytes, Paneth cells, and colonic stem cells. However, chromogranin A (CHGA) expression, a marker for enteroendocrine cells, was not detected, suggesting that enteroendocrine cells were not present in the epithelial layer of the organoids (Fig. 2E). To assess the functionality of the epithelial cells based on their integrity, we treated the organoids with forskolin and performed a swelling assay based on CF transmembrane conductance regulator (CFTR) activation (20). As a result, we confirmed that the organoids swelled in response to forskolin treatment, indicating the presence of functional epithelial cells (Fig. 2F).

### The reciprocal interaction between colon organoids and NCSCs promotes enterochromaffin cell differentiation

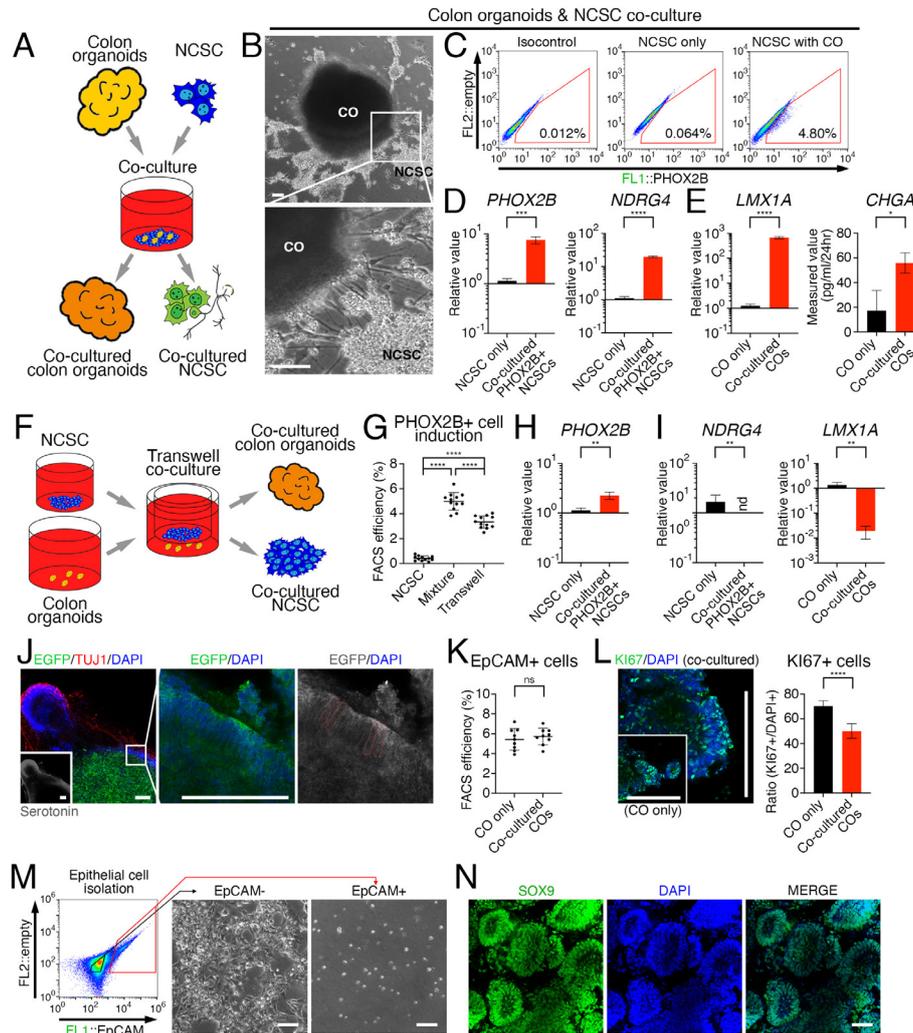
To investigate the reciprocal interaction between immature embryonic colonic tissue and the peripheral nervous system, we co-cultured colon organoids with NCSCs that were differentiated from PHOX2B::eGFP reporter hESCs (18) using a conventional method (21) (Fig. 3A). Within 7 days of co-culture, NCSCs showed morphological characteristics of neuronal differentiation (Fig. 3B), and about 5% of NCSCs expressed eGFP, indicating the expression of PHOX2B (Fig. 3C). FACS sorting was used to isolate NCSCs expressing PHOX2B after co-culture with colon organoids, which showed increased expression of *PHOX2B* and the enteric neuronal marker N-myc downstream-regulated gene 4 (*NDRG4*), compared to NCSCs not co-cultured with colon organoids (Fig. 3D). Meanwhile, colon organoids isolated after co-culture showed an enrichment of mRNA expression for enterochromaffin cell markers LIM homeobox transcription factor 1 alpha (*LMX1A*) and CHGA (Fig. 3E). To determine whether the reciprocal effects were due to direct contact or unidirectional influence, we attempted co-culture using transwells (Fig. 3F). The proportion of PHOX2B-expressing cells induced by co-culture with colon organoids decreased compared to NCSCs in direct contact with colon organoids, but still maintained an increased proportion compared to

NCSCs that were not co-cultured (Fig. 3G). The isolated PHOX2B-expressing cells were verified for their expression (Fig. 3H). In contrast, the expression of *NDRG4* and *LMX1A* in colon organoids co-cultured with NCSCs via transwells did not increase (Fig. 3I), unlike colon organoids in direct contact with NCSCs.

After co-culturing NCSCs and colon organoids directly for 10 days, differentiated neuronal cells expressing neuron-specific class III beta-tubulin (TUJ1) contained serotonin (Fig. 3J, lower panel), and the epithelial cells of colon organoids labeled with eGFP surrounded by the neuronal cells exhibited typical columnar morphology of colon epithelial cells (Fig. 3J). However, co-culture with NCSCs did not increase the number of epithelial cells expressing EpCAM (Fig. 3K). Instead, the proportion of Ki-67-expressing cells, which are proliferating cells, decreased within the colon organoids (Fig. 3L). When EpCAM-expressing cells were separated from the colon organoids after co-culture with NCSCs and re-plated, epithelial cells expressing EpCAM showed low efficiency for attachment and survival rates while non-epithelial cells that did not express EpCAM exhibited high survival and proliferation rates (Fig. 3M). After 10 days of re-cultivation, most rapidly proliferating cells from EpCAM-expressing cells expressed SOX9, an epithelial stem cell marker in the colon (22) (Fig. 3N).

### Immature neuron-secreted substance P promotes enteroendocrine cell differentiation of colonic organoids

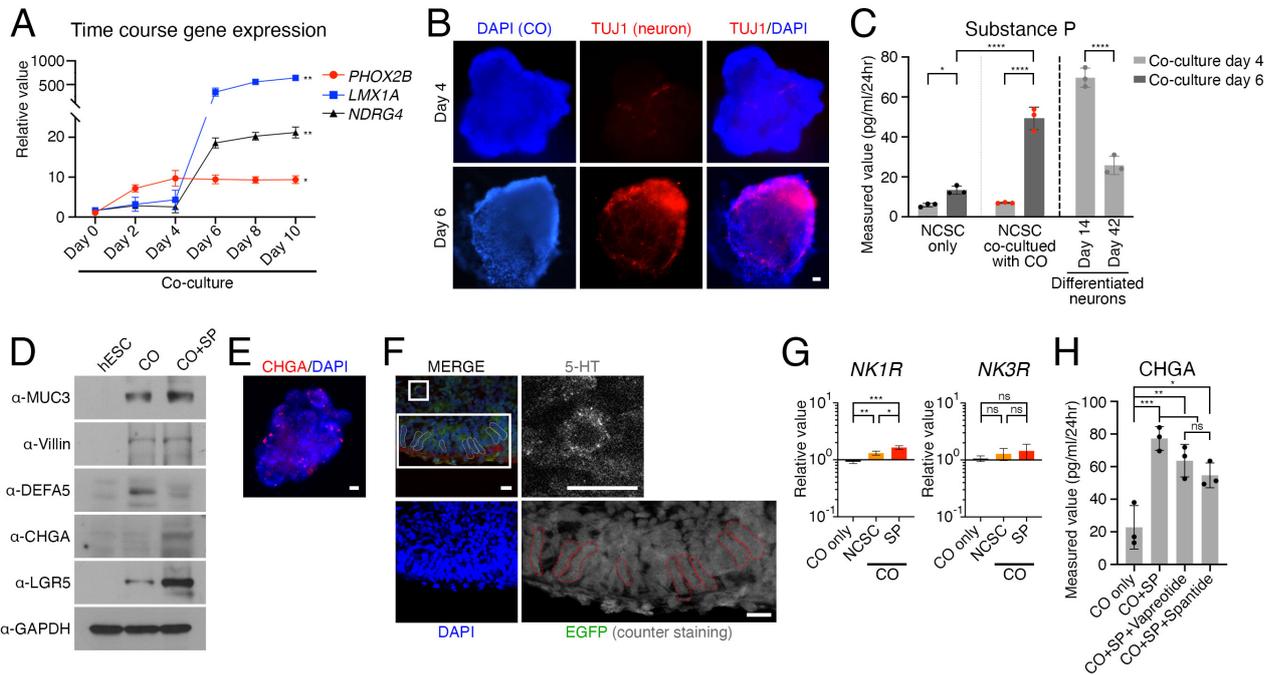
To investigate the influence of developing neurons on colonic organoids, we examined the time points during co-culture of NCSCs and colonic organoids at which *LMX1A* and *NDRG4* changed (Fig. 4A). Between days 4 and 6 of co-culture, we observed an increasing trend in both *NDRG4* in NCSC-derived neurons and *LMX1A* in colonic organoids. This coincided with a significant difference in TUJ1 expression, indicating neuronal differentiation of NCSCs (Fig. 4B). To further evaluate the impact of developing neurons on colonic organoids, we measured the quantity of a signaling molecule, substance P, which is spontaneously secreted by immature peptidergic neurons during early differentiation. We used ELISA to measure substance P levels in the co-culture medium on days 4 and 6. The result revealed a significant increase in substance P secretion on day 6 compared to day 4 in the NCSC-derived neurons co-cultured with colonic organoids (Fig. 4C). While the level of substance P was lower than that produced by immature neurons spontaneously secreting substance P alone, it was much higher than that of matured neurons or NCSCs not co-cultured with colonic organoids



**Fig. 3.** Reciprocal interaction between colonic organoid and NCSC-derived peripheral neurons. (A) Schematic representation of co-culture of colon organoids with neural crest stem cells (NCSCs). (B) Representative images of NCSCs showing neuronal differentiation after 7 days of co-culture with colon organoids. (C) Representative FACS plot for NCSCs expressing PHOX2B after co-culture with colonic organoids. (D) Increased expression of *PHOX2B* and *NDRG4* in FACS-isolated NCSCs after co-culture with colon organoids. (E) Enrichment of mRNA expression for *LMX1A* and *CHGA* in colon organoids after co-culture with NCSCs. (F) Schematic representation of transwell co-culture system. (G) Percentage of PHOX2B-expressing NCSCs induced by co-culture with colon organoids using direct contact or transwells. (H) Verification of PHOX2B-expressing cells after co-culture with colon organoids. (I) Expression of *NDRG4* and *LMX1A* in colon organoids after co-culture with NCSCs using transwells. (J) Differentiated neuronal cells expressing TUJ1 containing serotonin (lower panel) and epithelial cells of colon organoids labeled with eGFP. Columnar morphology of epithelial cells are indicated by dotted red line in the right panel. (K) Proportion of EpCAM-expressing epithelial cells in colon organoids after co-culture with NCSCs. (L) Representative immunostaining image of Ki-67-expressing proliferating cells within colon organoids after co-culture with NCSCs, and the counting result. (M) Representative FACS plot for the isolation of EpCAM-expressing cells of colonic organoid, and representative images for attached cells after re-plating. (N) SOX9 expression in proliferating cells from EpCAM-expressing cells after 10 days of re-cultivation. Data expressed as mean  $\pm$  SD (n=9, 12 for G, biological repeat). p-values calculated by unpaired t-test (D, E, H, I, K, L) or one-way ANOVA (G). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001, ns=not significant. All scale bars=50  $\mu$ m. CO: colonic organoid.

(Fig. 4C). Upon direct administration of substance P to colonic organoids, we observed a significant decrease in the expression of DEFA5 protein, while the expression of CHGA protein increased (Fig. 4D). Interestingly, we

found that cells expressing CHGA were scattered throughout the organoid instead of being limited to the epithelial layer (Fig. 4E). Additionally, the epithelial cells outside of the organoid maintained their columnar morphology,



**Fig. 4.** Developing neurons influence colonic organoids. (A) Tracking the expression changes of *PHOX2B* and *NDRG4* in NCSCs and *LMX1A* in colonic organoids according to the co-culture period of colonic organoids and NCSCs. (B) Immunostaining images of NCSC-derived neurons expressing TUJ1 as a key difference between co-culture time points of 4 and 6 days. (C) Significant increase in substance P in the co-culture medium on day 6 compared day 4, as measured by ELISA. (D) Expression of colonic epithelial cell marker proteins of colonic organoid after substance P administration were validated by immunoblotting. (E) Scattering distribution of cells expressing CHGA in the colonic organoid was confirmed using immunofluorescence staining. (F) Presence of enteroendocrine cells detectable by 5-HT within the colonic organoid, and columnar epithelial cell morphology (red dotted line). (G) The expression of *NK1R* and *NK3R* mRNA in colonic organoids was validated using qRT-PCR, with or without treatment with substance P. (H) Protein expression levels of CHGA in colonic organoids were measured using ELISA after inhibition of *NK1R* with specific antagonists. Data expressed as mean  $\pm$  SD (n=s, biological repeat). p-values calculated by two-way ANOVA (A and C) or one-way ANOVA (G and H). \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001, ns=not significant. Scale bars=25  $\mu$ m (B and E) or 10  $\mu$ m (F). CO: colonic organoid, SP: substance P.

and we were able to confirm the presence of enteroendocrine cells detectable by 5-hydroxytryptamine (5-HT) within the organoid (Fig. 4F) (23). Notably, the expression of neurokinin 1 receptor (*NK1R*), which is known as a receptor for substance P (24), increased in colonic organoids co-cultured with NCSC-derived neurons or treated with Substance P (Fig. 4G). To investigate whether the interaction between peripheral neurons and colonic organoids due to a conventional mechanism of substance P via *NK1R*, we utilized *NK1R* antagonists for inhibition. However, the increase in CHGA protein level was not significantly attenuated (Fig. 4H). The findings suggest that substance P plays an important role in regulating the expression of *NK1R* and cells expressing CHGA in colonic organoids, but it does not selectively target the cells expressing *NK1R* in a conventional state.

## Discussion

In this study, we developed a comprehensive protocol for generating colon organoids that closely resemble key features of the developing colon. The protocol involves inducing differentiation of cells towards definitive endoderm, followed by embedding hindgut cells in an artificial extracellular matrix (ECM), and stimulating BMP4 to initiate colonic differentiation. Our hypothesis was that BMP4 plays a crucial role in determining the orientation of cells, particularly the epithelial lining, instead of in overall gut tube formation since it is expressed symmetrically only in the gut tube and is not detectable in the mesentery (25-27). Our findings revealed that adding BMP4 instead of previously used BMP2 (15) during the later stage of the culture period induced the formation of epithelial cells with posterior gut tube specification (19, 28). These organoids exhibited functional epithelial cells such as goblet cells,

enterocytes, Paneth cells, and colonic stem cells, as evidenced by the presence of markers such as MUC3, MUC4, Villin, DEFA5, and LGR5 (29, 30). Additionally, the organoids responded to forskolin treatment, indicating the presence of functional epithelial cells (20). Despite these promising results, enteroendocrine cells were not detected in the epithelial layer of the organoids. Thus, further optimization was required to promote their differentiation.

To investigate the interaction between colonic organoids and the peripheral nervous system, we co-cultured colonic organoids with NCSCs. We observed that NCSCs differentiated into enteric NCSCs expressing PHOX2B and neurons expressing TUJ1 (18), while colonic organoids showed abundant mRNA expression for the enterochromaffin cell markers *LMX1A* and *CHGA*. This suggests that the interaction between colon organoids and NCSC-derived neurons promotes enteric chromaffin cell differentiation, further implying the potential role of the enteric nervous system in gut development (12). Interestingly, co-cultured colon organoids and NCSCs spontaneously differentiated into peripheral neurons expressing PHOX2B without any specific stimulation, modeling the behavior of migrating NCSCs naturally committing to enteric fate when approaching the developing colon along the dorso-ventral track *in vivo* (21, 31). Moreover, substance P, which was secreted from early differentiated neurons in co-culture, has been found to promote intestinal epithelial differentiation. This observation suggests that immature early differentiated neurons possess the ability to spontaneously release neuropeptides nonsynaptically and contribute to tissue formation during development (32-35).

However, our study has limitations of oversimplifying the complex interaction between developing immature gut and differentiating peripheral neurons. The enteric nervous system in the colon is composed of various types of neurons and enteric glial cells (36), forming complex neural networks in the submucosal and myenteric plexuses (37). To clarify the impact of interactions between immature organoids and developing neurons on the functional maturation of the gut epithelium and the functional specification of neural cells, it is necessary to differentiate the various types of neural cell constituents and conduct further investigations into the precise mechanisms underlying nonsynaptic transmission of substance P.

Despite certain limitations, our research sheds light on the crucial role of immature neurons in the process of development. Our study indicates that even though the colon requires fewer enterochromaffin cells compared to the small intestine, peripheral nerve innervation is essential for the presence of the limited number of enterochromaffin

cells in the colon. Our results also suggest that irregular neural activation or dysfunctional nerve cells that regulate neuropeptides and neurotransmitters such as substance P may lead to abnormal increases in enterochromaffin cells in the rectum of irritable bowel syndrome (IBS) patients (38-41). Currently, there is a dearth of research models for neurological disorders like IBS, and our colonic organoids innervated by neurons may prove invaluable for studying relevant mechanisms and developing therapeutic interventions to regulate neuropeptides and neurotransmitters. Moreover, considering the role of the microbiome in gut health, further studies may explore the interaction between colon organoids and the microbiota in the gut. Understanding the interplay between the microbiota and the nervous system may help unravel the complex mechanisms underlying the development of gastrointestinal disorders and facilitate the development of more effective treatments (42, 43).

Overall, our findings have far-reaching implications for the fields of developmental biology and tissue engineering. The stepwise protocol we used to generate colon organoids can be optimized further to facilitate the differentiation of other cell types in the colonic epithelial cells. Additionally, investigating the interaction between colon organoids and the peripheral nervous system can deepen our understanding of the mechanisms that regulate gut development, function, and diseases. Future studies may lead to the development of novel therapeutic interventions for gastrointestinal disorders and enhance our knowledge of the complexities of organ development and function.

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### Author Contributions

Y.H.C., I.Y.C., and C.E.S.: performing experiments, data analysis, assembly, interpretation, and writing manuscript; C.P., S.K.L., and J.H.K.: performing experiments and data analysis; S.H.S.: performing experiments; S.L. and J.H.P.: study design; Y.J.K.: conception, study design, performing experiments, data analysis, assembly, interpretation, and writing a manuscript.

### Potential Conflict of Interest

The authors have no conflicting financial interest.

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