

Clinical Features and Molecular Diagnosis of CATCH-22 Syndrome

Jung Yun Choi, MD^{1,2}, Jeong-Wook Seo, MD^{1,3}, Myoung Hee Kim, PhD⁴,
Eul Kyung Kim, MD³, Jung-Sun Kim, MD³, Ho Sung Kim, MD²,
Chong Heon Lee, DDS³, Hyangsuk Hur, PhD⁴, Eun Jung Bae, MD⁵,
Chung IL Noh, MD^{1,2} and Yong Soo Yun, MD^{1,2}

¹Heart Research Institute, Medical Research Center, Seoul National University, Seoul, ²Departments of Pediatrics and ³Pathology, Seoul National University College of Medicine, Seoul, ⁴Korea Research Institute of Bioscience and Biotechnology, KIST, Taejon, ⁵Department of Pediatrics, Sejong General Hospital, Buchon, Kyunggi-do, Korea

CATCH-22 증후군의 임상 소견 및 분자유전학적 진단

최정연^{1,2} · 서정욱^{1,3} · 김명희⁴ · 김을경³ · 김정선³ · 김호성²
이종현³ · 허향숙⁴ · 배은정⁵ · 노정일^{1,2} · 윤용수^{1,2}

국문초록

연구배경 : CATCH - 22 22q11 , , CATCH - 22 12
 . 방법 : CATCH - 22
 fluorescent in - situ hybridization(FISH) 22q11
 12 7가 ,
 , Southern blot . 결과 : 12
 7가 4가 가 7 , 1 3가
 가 5 . 8 Fallot 4 7 가 . 4
 . Southern blot 8
 6 . 결론 : CATCH - 22 Fallot 4
 , FISH Southern blot
 . (Korean Circulation J 1998;28(7):1077-1083)
 중심 단어 : CATCH - 22 . DiGeorge . 22q11 .

: 1998 6 22 / : 1998 7 21

Corresponding author : Jeong-Wook Seo, MD. 28 Yongon-Dong, Chongno-Gu, Seoul 110-799, Korea
 Department of Pathology, Seoul National University, College of Medicine, Seoul, Korea
 TEL : (02) 740-8268 · FAX : (02) 765-5600
 E-mail : jwseo@plaza.snu.ac.kr

Introduction

CATCH-22 is an acronym of cardiac defect, abnormal face, thymic hypoplasia, cleft palate, and hypocalcemia associated with microdeletion of chromosome 22q11.¹⁾ This syndrome encompasses heterogeneous groups of patients with DiGeorge syndrome (DGS),²⁾ velocardiofacial syndrome (VCFS),³⁾ and conotruncal anomaly face syndrome (CTAFS).⁴⁾ This syndrome is currently considered to be the second most common cause of congenital heart disease, preceded only by Down's syndrome.⁵⁾ In a survey in the northern England, deletion of 22q11 was estimated to account for 5% of all congenital heart defects and showed a minimum prevalence of 1 in 4000 births.⁵⁾ The prevalence of this syndrome in Korea and the type of associated cardiac lesions are not known, but in view of the relative incidence of congenital heart disease, it is estimated that there are at least ten thousand cases.

The important aspect of this syndrome is that the cardiac lesion is less severe and the mental retardation is milder than those in Down's syndrome.⁵⁾ Number of patients with 22q11 deletion, therefore, is expected as being far more than that of Down's syndrome in the clinical practice. Moreover, phenotypic and genotypic features are so variable and variability between monozygotic twins has been reported.⁶⁾ Several genes are being studied but no gene have been proven specific for this syndrome.⁷⁻¹⁰⁾ The fluorescent in situ hybridization (FISH) is currently understood as the most specific method to diagnose this genetic defect.¹¹⁾¹²⁾

We performed FISH study for metaphase chromosomes using a D22S75 probe. Twelve patients were collected to reveal genetic features of CATCH-22 syndrome. We report the clinical and genetic profile of those Korean cases of CATCH-22 syndrome. Eight cases among them were additionally studied by Southern blot analysis of genomic DNA using DGCR680 and pDH-1 probes.

Materials and Methods

Patients

A total of twelve cases of congenital heart disease

with abnormal face were proven to have a deletion at the chromosome 22q11 by FISH analysis during the one-year period ending June 1997 (Table 1). Clinical diagnosis was performed at Seoul National University Children's Hospital (eight cases), Sejong General Hospital (four cases). Facial features were analyzed according to seven independent items (Table 1). Faces with positive result on four or more items were interpreted as abnormal face, faces with two or three positive items being equivocal. Cardiac diagnosis was confirmed by echocardiography, angiocardiography and/or surgery.

Fluorescent in situ hybridization for metaphase chromosome

Metaphase chromosome slides were prepared from culture of peripheral lymphocytes by a standard method which included exposure to a mitotic arrestant (colcemid), treatment with a hypotonic solution (0.075M KCl), fixation (3 : 1 mixture of methanol and glacial acetic acid) and dehydration (70%, 80%, 90% ethanol). DNA on slides was denatured in 70% formamide/2X saline sodium citrate (SSC) for 2 minutes at 70 °C, immediately dehydrated through a cold (-20 °C) ethanol series, and air dried. The hybridization mixture containing the digoxigenin-labeled D22S75 DGCR probe with D22S39 chromosome 22 control probe (Oncor, U.S.A.) was placed on denatured chromosome slides for in situ hybridization. After overnight hybridization at 37 °C in a moist chamber, slides were washed once for 5 minutes in 2X SSC at 72 °C, and once for 2 minutes in 1X phosphate buffered distilled water (PBD) at room temperature; they were incubated with anti-digoxigenin fluorescein in 1X PBD/5% BSA at 37 °C for 60 minutes and rinsed three times in 1X PBD (2 minutes each time). For amplification, slides were treated with rabbit anti-sheep antibodies at 37 °C for 30 minutes and washed in three changes of 1X PBD at room temperature (2 minutes each time). They were then incubated with FITC-rabbit antibody at 37 °C for 30 minutes, and then washed three times in 1X PBD. Slides were stained with propidium iodide and mounted with fluorescent mounting media (DA-KO, U.S.A.). For observation of

fluorescence signals in the chromosome, a fluorescence microscope equipped with appropriate fluorescence filter sets was used.

Production of probes for Southern hybridization

We used a mixture of two DNA probes, DGCR680 and pDH-1.¹³⁾¹⁴⁾ DGCR680 was obtained from human genomic DNA using two primers based on ADU breakpoint, where a DiGeorge syndrome patient had balanced translocation.¹⁵⁾ The other probe pDH1 was obtained by screening from a human liver cDNA library using DGCR680 as a probe. These together covered 1300 base pairs within the deleted sequence of chromosome 22¹⁶⁾ (Fig. 1). Details of our probes are described elsewhere.¹³⁾¹⁴⁾

Southern blot analysis

Genomic DNA extracted from peripheral blood of patients was digested with restriction enzyme HindIII. After Southern hybridization, the test probe signal at 18 kilobases and control signal at 16 kilobases were measured by densitometer.¹³⁾ The values representing locus copy number were obtained, standardized from quantitative analysis of the hybridization signals obtained with the 18kb fragment and compared to a low copy repeat intensity. It was decided that a value less than 1.50 indicated deletion.¹³⁾¹⁴⁾

Results

Of the twelve patients, eight cases were male and four cases were female ; their ages ranged from 3

month to 16 years (Table 1). Although all of twelve patients were initially suspected for CATCH-22 syndrome from the facial features observed by pediatric cardiologists, only seven cases were determined to have abnormal facial morphology according to our criteria using seven items (Table 1). A cleft palate was present in a patient and a high arched palate in six. Nine patients showed delayed development. Two cases had umbilical hernia. The main cardiac lesion of eight patients were tetralogy of Fallot (TOF) and seven of them had pulmonary atresia. Two cases had other anomalies in the ventricular outflow tract, being common arterial trunk or pulmonary stenosis. Two cases had a patent arterial duct or atrial septal defect(ASD) (Table 1).

All of twelve patients had positive result on FISH study. Eight cases among twelve patients were studied with Southern blot analysis. Six cases among eight patients were positive for Southern blot analysis.

There were six patients with positive FISH and positive Southern analysis. Five of them had typical facial features but one had an equivocal face. The cardiac lesion in every case was TOF and pulmonary atresia.

Both of two cases with positive FISH and negative Southern analyses had equivocal face and the cardiac lesions were TOF or ASD.

Four cases were studied only by FISH method. The cardiac diagnoses were TOF, pulmonary stenosis, common arterial trunk and isolated patent arterial duct (PDA). The facial morphology of two cases had typical facial features.

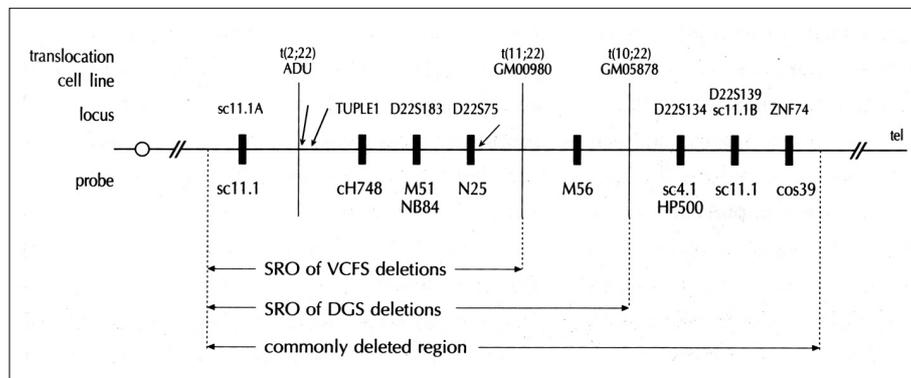


Fig. 1. Diagram of the chromosomal region at 22q11. Double arrow indicates the location of the probe (DGCR680 and pDH-1) and single arrow indicates the location of the probe for FISH study. Modified from Mulder et al.¹⁶⁾ (SRO : smallest region of overlap)

Table 1. Facial features, cardiac diagnosis and molecular findings of patients with CATCH-22 syndrome. Clinical decision of being "abnormal" was made when there are four or more positive findings. Clinical decision of being "equivocal" was made when there are one to three positive findings on their face. Every case showed positive reaction for the fluorescent in-situ hybridization study.

No.	Age/ Sex	Facial features*							Palate	Develop- ment	Ca/P**	Others	Clinical decision	Cardiac lesions***	Aortic arch	Southern blot
		1	2	3	4	5	6	7								
1	3m/M	-	+	+	+	+	+	+	High	Normal	8.6/6.2		Abnormal	TOF, PA	Rt	1.12
2	21m/M	+	-	+	+	+	+	+	High	Delayed			Abnormal	TOF	Lt	
3	15m/M	?	-	?	+	+	+	+	Normal	Delayed		Umb. hernia	Abnormal	PS	Rt	
4	3m/F	+	-	-	-	+	-	-	High	Normal			Equivocal	CAT	Lt	
5	9m/M	+	-	-	-	+	-	-	High	Delayed			Equivocal	PDA	Lt	
6	31m/M	?	?	+	+	+	+	?	High	Delayed	9.0/4.9	Umb. hernia	Abnormal	TOF, PA, PDA	Lt	1.00
7	30m/F	?	?	?	?	?	?	?	Normal	Delayed	8.9/6.6		Equivocal	TOF, PA	Rt	0.43
8	7m/F	-	-	+	-	-	+	-	Normal	Delayed			Equivocal	TOF, PA		2.32
9	2yr/F	?	?	?	+	+	+	-	Normal	Delayed			Equivocal	ASD		1.94
10	6m/M	-	-	+	+	+	+	-	High	Delayed			Abnormal	TOF, PA		0.49
11	16yr/M	+	+	+	-	-	+	+	Cleft	Delayed	8.3/7.1		Abnormal	TOF, PA	Lt	0.78
12	3yr/M	?	+	+	+	?	?	+	Normal	Normal	9.1/4.9		Abnormal	TOF, PA	Rt	1.21

*1, long face ; 2, flat malar area ; 3, depressed nose and narrow alae ; 4, retarded mandible ; 5, small A-shaped mouth ; 6, hypertelorism ; 7, bloating eyelid

**Ca/P, serum level of calcium and phosphorus

***TOF, Tetralogy of Fallot ; PA, pulmonary atresia ; ASD, atrial septal defect ; PS, pulmonary stenosis ; DORV, double outlet right ventricle ; CAT, common arterial trunk ; PDA, patent ductus arteriosus

Discussion

DiGeorge syndrome was the first to be recognized as a group of defects currently understood as CATCH-22 syndrome²⁾ and the association of cardiac anomalies in this immunologic defect was studied in 1972.¹⁷⁾ The hypocalcemic features, such as seizure, and cellular immune deficiency are seen early in life, but they become less apparent with age, probably due to secondary compensations. Frequent cardiac lesions are interrupted aortic arch (IAA), CAT, TOF with PA, TOF, and isolated VSD.⁵⁾ Velocardiofacial syndrome (VCFS) or Shprintzen syndrome, with its characteristic facial features comprising a prominent nose, broad nasal root, narrow palpebral fissures and retrognathia were reported.³⁾¹⁸⁾¹⁹⁾ Typical cases show a high incidence of craniofacial anomalies ; cleft palate (98%), pharyngeal hypotonia (90%), retrognathia (80%) and malar flatness (70%) ; the incidence of congenital heart disease has been reported as 82%²⁰⁾ ; Driscoll et al. and Shprintzen et al. reported that VSD and TOF were the common cardiac lesions.¹⁸⁾²¹⁾ The Japanese

group led by Takao²²⁾ was the first to recognize the major contribution of the phenotype to the patient population with the ventricular outflow tract defects. The prevalence of CTAFS, based on the facial features in a series of TOF, was 12.8%, but this figure was 48% in the subgroup of TOF associated with PA and major aortopulmonary collateral arteries.²²⁾ Among clinical cases of CTAFS, the most common cardiac defect was TOF (92%) ; half of these cases were associated with the PA and the systemic pulmonary collateral arteries.²³⁾

Within chromosome region 22q11, reported frequencies of association with deletion are 89% in DGS, 81% in VCFS, and 84% in CTAFS.²¹⁾²³⁾ But it is also possible that non-deletion patients have undetectable smaller deletions or point mutations within critical genes in this region.²⁴⁾ Deletions were also observed in 20-30% of such unselected, nonsyndromic patients with CAT, IAA, and TOF.²⁵⁾ Among patients requiring surgery for congenital heart disease, 5% to 10% may have this single genetic abnormality.²⁶⁾ Parental deletions are found in approximately 25% of patients with CATCH-22, though variable expression within a

family is well documented.¹⁾⁶⁾²⁷⁾ Among 40 parents of children with congenital heart disease, one of 14 fathers and five of 26 mothers had CTAFS and in one father and four mothers there was deletion.²³⁾

Our study aims to reveal the clinical significance of this genetic defect in the pediatric cardiological practice in Korea and to assess the significance of different genetic studies in the diagnosis of this syndrome. This study is the first report on the systematic study on this syndrome in Korea.

As is shown in Table 1, facial features of individual cases were so variable that no single criteria could be used as an indicator of this syndrome. But we could define an abnormal face when one had abnormalities on four or more items and equivocal when one had abnormalities on two or three items. The incidence of this disease, therefore, varies when we define this disease through the clinical examination of the facial features only. We could divide facial features into abnormal, equivocal or normal faces. But this classification system should be a simple screening tool for patients with congenital heart disease.

In view of the variability of phenotypic features observed in association with 22q11 deletions, we cannot precisely predict outcome on the basis of molecular studies at this time. The observed variability may reflect the amount of deleted genes in that critical region, or may be dependent upon genetic background, in utero environment, or parental origin of the deletion.²⁸⁾ Correlation between genotype and phenotype will require a detailed molecular analysis of the deleted region to determine which region or genes specify individual features of the phenotype. Further molecular studies involving the functional characteristics of probes D22S75, DGCR680 and pDH-1 should also be performed. Other tools for the molecular diagnoses include RFLP (Restriction Fragment Length Polymorphism), DNA dosage analysis in addition to the FISH or Southern blot analysis. But one of important aspect of the study would be those for their family members.

We conclude that CATCH-22 syndrome has variable facial, cardiac and genetic features, and the combined use of probes is recommended for a more accurate

diagnosis.

Summary

Background :

CATCH-22 syndrome is a common genetic disorder with features of cardiac defect, abnormal face, thymic hypoplasia, cleft palate, and hypocalcemia, along with microdeletion at chromosome 22. This study is to report twelve Korean patients with CAT-CH-22 syndrome diagnosed by the fluorescent in situ hybridization (FISH) method.

Method :

Clinical features were analyzed according to the FISH result and the Southern blot analysis using new probes DGCR680 and pDH-1 was performed to correlate with the clinical findings and FISH results. Twelve patients were studied by FISH method and eight of them were studied by Southern blot analysis.

Results :

Seven patients had typical facial features for CATCH-22 syndrome, but five patients had equivocal face, although they were originally suspected to have the conotruncal face. The main cardiac lesion of eight patients were tetralogy of Fallot (TOF) and seven of them had pulmonary atresia. Two cases had other anomalies in the ventricular outflow tract, being common arterial trunk or pulmonary stenosis. Two cases had a patent arterial duct or atrial septal defect (ASD). All of twelve patients had positive result on FISH study. Among eight patients with positive FISH study, six cases were positive for Southern blot analysis.

Conclusion :

We conclude that CATCH-22 syndrome has variable facial, cardiac and genetic features, and the combined use of probes is recommended for a more accurate diagnosis.

KEY WORDS : CATCH-22 syndrome · Conotruncal anomaly face syndrome · DiGeorge syndrome · Chromosome 22q11 · Congenital heart disease.

Acknowledgment

The authors are indebted to Drs. Hye Soon Kim, Hong Ryang Kil, Soon Sung Park, Seong Ho Kim, Se Jung Sohn, Mee Hye Oh, Shi Joon Yoo and Heung Jae Lee who actively participated in our collection of clinical cases. Drs. Dong Soo Kim and Soo Kyung Choi helped our cytogenetic study. We are also grateful to Ms. Sung Hee Hong and GiJin Kim for their valued technical assistance with cytogenetic study.

1996 (96 - B00 -
02 - 001 - 004)

REFERENCES

- 1) Wilson DI, Burn J, Scambler P, Goodship J. *DiGeorge syndrome: Part of CATCH 22*. *J Med Genet* 1993;30:852-6.
- 2) DiGeorge AM. *Congenital absence of the thymus and its immunologic consequences: Concurrence with congenital hypothyroidism*. In *Birth Defects. White plains: March of Dimes;1968. p.116-21*.
- 3) Shprintzen RJ, Golderg RB, Young D, Wolford L. *The velocardio-facial syndrome: A clinical and genetic analysis*. *Pediatrics* 1981;67:167-72.
- 4) Takao A, Ando M, Cho K, Kinouchi A, Murakami Y. *Etiologic categorization of common congenital heart disease*. In: *Van Praagh R, Takao A, editors. Etiology and Morphogenesis of Congenital Heart Disease. New York: Futura;1980. p.253-69*.
- 5) Burn J, Wilson DI, Cross I, Atif U, Scambler P, Takao A, et al. *The clinical significance of 22q11 deletion*. In: *Clark EB, Markwald RR, Takao A, editors. Developmental Mechanisms of Heart Disease. Armonk NY: Futura;1995. p.559-67*.
- 6) Goodship J, Cross I, Scambler P, Burn J. *Monozygotic twins with chromosome 22q11 deletion and discordant phenotype*. *J Med Genet* 1995;32:746-8.
- 7) Morrow B, Goldberg R, Carlson C, Gupta RD, Sirotkin H, Collins J, et al. *Molecular definition of the 22q11 deletions in velocardio-facial syndrome*. *Am J Hum Genet* 1995;56:1391-403.
- 8) Halford S, Wadey R, Roberts C, Daw SCM, Whiting JA, O'Donnell H, et al. *Isolation of putative transcriptional regulator from the region of 22q11 deleted in DiGeorge syndrome, Shprintzen syndrome and familial congenital heart disease*. *Hum Molec Genet* 1993;12:2099-107.
- 9) Pizzuti A, Novelli G, Mari A, Ratti A, Colosimo A, Amati F, et al. *Human homologue sequences to the Drosophila dishevelled (sic) segmentpolarity gene are deleted in the DiGeorge syndrome*. *Am J Hum Genet* 1966;58:722-9.
- 10) Demczuk S, Thomas G, Aurias A. *Isolation of a novel gene from the DiGeorge syndrome critical region with homology to Drosophila gdl and to human LAMC1 genes*. *Hum Molec Genet* 1996;5:633-8.
- 11) Demczuk S, Levy A, Aubry M, Croquette MF, Philip N, Prieur M, et al. *Excess of deletions of maternal origin in the DiGeorge/velocardio-facial syndromes: A study of 22 new patients and review of the literature*. *Hum Genet* 1995;96:9-13.
- 12) Kim HS, Kim HS, Rho JI, Choi JY, Yum YS, Kim JS, et al. *Clinical study of CATCH-22*. *J Korean Pediatr Soc* 1995;38:1603-9.
- 13) Hur HS. *Molecular genetic studies on the gene disrupted by a balanced translocation associated with DiGeorge syndrome (DGS)*. Chungnam University 1997. (Thesis)
- 14) Hur HS, Kim YJ, Noh JI, Seo JW, Kim MH. *Diagnosis of CATCH-22 syndrome by molecular genetic analysis*. 1998. (in preparation)
- 15) Budarf ML, Collins J, Gong W, Roe B, Wang Z, Bailey LC, et al. *Cloning a balanced translocation associated with DiGeorge syndrome and identification of a disrupted candidate gene*. *Nature Genet* 1995;10:269-78.
- 16) Mulder MP, Wilke M, Langeveld A, Wilming LG, Hagenmeijer A, van Drunen E, et al. *Positional mapping of loci in the DiGeorge critical region at chromosome 22q11 using a new marker (D22S183)*. *Hum Genet* 1995;96:133-41.
- 17) Freedom RM, Rosen FS, Nadas AS. *Congenital cardiovascular disease and anomalies of the third and fourth pharyngeal pouch*. *Circulation* 1972;46:165-72.
- 18) Shprintzen RJ, Golderg RB, Young D, Wolford L. *The velocardio-facial syndrome: A clinical and genetic analysis*. *Pediatrics* 1981;67:167-72.
- 19) Kelly D, Goldberg R, Wilson D, Lindsay E, Carey A, Goodship J, et al. *Confirmation that the velocardio-facial syndrome is associated with haplo-insufficiency of genes at chromosome 22q11*. *Am J Med Genet* 1993;45:308-12.
- 20) Goldberg R, Motzkin B, Marion R, Scambler PJ, Shprintzen RJ. *Velocardio-facial syndrome: A review of 120 patients*. *Am J Med Genet* 1993;45:313-9.
- 21) Driscoll DA, Goldmuntz E, Emanuel BS. *Detection of 22q11 deletions in patients with conotruncal cardiac malformations, DiGeorge, velocardiofacial, and conotruncal anomaly face syndromes*. In: *Clark EB, Markwald RR, Takao A, editors. Developmental Mechanisms of Heart Disease. New York: Futura;1995. p.560-75*.
- 22) Kinouchi A. *A study of specific peculiar facial features of conotruncal anomaly*. *J Tokyo Women Med Coll* 1980;50:396-409.
- 23) Matsuoka R, Takao A, Kimura M, Imamura S, Kondo C, Joho K, et al. *Confirmation that the conotruncal anomaly face syndrome is associated with a deletion within 22q11.2*. *Am J Med Genet* 1994;53:285-9.
- 24) Driscoll DA, Salvin J, Sellinger B, Budarf M, McDonald-McGinn D, Zackai EH, et al. *Prevalence of 22q11 microdeletions in DiGeorge and velocardiofacial syndromes: Implications for genetic counselling and prenatal diagnosis*. *J Med Genet* 1993;30:813-7.
- 25) Goldmuntz E, Driscoll D, Budarf ML, Zackai EH, McDonald DM, McGinn JA, et al. *Microdeletions of chromosomal region 22q11 in patients with congenital conotruncal cardiac defects*. *J Med Genet* 1993;30:807-12.
- 26) Payne RM, Johnson MC, Grant JW, Strauss AW. *Toward a molecular understanding of congenital heart disease*. *Circulation* 1995;91:494-504.
- 27) De Silva D, Duffty P, Booth P, Auchterlonie I, Morrison N, Dean JCS. *Familial studies in chromosome*

- 22q11 deletion: Further demonstration of phenotypic heterogeneity. Clin Dysmorph 1995;4:294-303.*
- 28) Halford S, Wade R, Roberts C, Daw SCM, Whiting JA, O'Donnell H, *et al.* *CATCH 22: Can molecular genetics*

explain the phenotype? In: Clark EB, Markwald RR, Takao A, editors. Developmental Mechanisms of Heart Disease. New York: Futura;1995. p.577-80.