

# NETosis in Autoimmune Diseases

Dong Hyun Sohn

Department of Microbiology and Immunology, Pusan National University School of Medicine, Yangsan, Korea

Neutrophils are the major antimicrobial cells of the innate immune system, which are recruited rapidly to the sites of infection and provide the primary defense against pathogens. Recent evidence suggests that neutrophils undergo a distinct cell death mechanism called NETosis, which not only contributes to the host defense, but also leads to severe pathological immune responses in cases of dysregulation. Here, we review the general features of NETosis as well as the generation of autoantigens and damage-associated molecular patterns by NETosis in autoimmune diseases. This review discusses the pathogenic role of NETosis in rheumatoid arthritis and systemic lupus erythematosus, where neutrophils may play a key role in the pathogenesis of these diseases, and suggest the possibility of neutrophil extracellular traps as biomarkers and therapeutic targets for the treatment of autoimmune diseases. (*J Rheum Dis* 2016;23:82-87)

**Key Words.** Neutrophil extracellular traps, NETosis, Citrullination, Damage-associated molecular patterns, Autoimmune diseases

## INTRODUCTION

Neutrophils are the most abundant leukocytes in mammals and form a crucial part of the innate immune system as a first line of defense against pathogens [1]. Upon infection, they are rapidly recruited to the sites of infection or inflammation by chemoattractants and inflammatory cytokines, which are secreted by tissue-resident macrophages and other sentinel cells [2]. In response to these molecules, neutrophils invade the infected tissues and play critical roles for clearance of the infection and recruitment of other immune cells. They possess multiple defense mechanisms, including phagocytosis, nicotinamide adenine dinucleotide phosphate (NADPH)-dependent oxidative burst, and degranulation of antimicrobial proteins to handle the infection and inflammation [3]. In 2004, it was reported that neutrophils have a distinct host defense mechanism to trap and kill pathogens, which are so called neutrophil extracellular traps (NETs) [4]. The release of NETs is a specialized form of neutrophil death, termed NETosis, which is dis-

tinct from apoptosis and necrosis [5]. This unique feature of neutrophils contributes to host defense, but also chronic inflammation and autoimmune diseases resulting from detrimental bystander damage. Here we focus on the generation of autoantigens and damage associated molecular patterns (DAMPs) by NETosis. This review will provide an overview of the contribution of NETosis to the pathogenesis of autoimmune diseases, such as rheumatoid arthritis (RA) and systemic lupus erythematosus (SLE). The involvement of NETosis in other diseases including antiphospholipid syndrome (APS), anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis, and cancer are discussed elsewhere [2,6].

## MAIN SUBJECTS

### NETosis

NETosis is a form of cell death unique to neutrophils that releases NETs, web-like structures of chromatin fiber and additional proteins. NETs are composed of decondensed chromatin with histones and various antimic-

Received : March 29, 2016, Revised : April 17, 2016, Accepted : April 17, 2016

Corresponding to : Dong Hyun Sohn, Department of Microbiology and Immunology, Pusan National University School of Medicine, 49 Busandaehak-ro, Mulgeum-eup, Yangsan 50612, Korea. E-mail : dhsohn@pusan.ac.kr

pISSN: 2093-940X, eISSN: 2233-4718

Copyright © 2016 by The Korean College of Rheumatology. All rights reserved.

This is a Free Access article, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

robial proteins such as neutrophil elastase (NE), myeloperoxidase (MPO), and cathepsin G [4,5]. Most of NET-associated proteins are of granular origin with several nuclear and cytoplasmic origins (Table 1) [7-10]. NETs serve as a physical barrier to prevent microbial dissemination by capturing and killing pathogens and increase the local concentration of antimicrobial effectors [11]. However, recent evidence suggests that NETs also have detrimental effects in autoimmune diseases by providing autoantigens and contributing to tissue damaging inflammation.

NETosis can be initiated by diverse stimuli that can be broadly classified into microbes, microbe products, host factors, and others (Table 2) [3,12,13]. Neutrophil activation in response to these factors is mediated by a multi-

**Table 1.** Neutrophil extracellular trap-associated proteins [7-10]

Cellular localization	Protein name
Granules	Azurocidin
	Bactericidal/permeability-increasing protein (BPI)
	Cathelicidin/LL37
	Cathepsin G
	Defensins 1 and 3
	Gelatinase B/matrix metalloproteinase 9
	Gelatinase-associated lipocalin
	Lactotransferrin
	Lysozyme C
	Myeloperoxidase (MPO)
	Neutrophil elastase (NE)
	Proteinase 3 (PR3)
	Histone H2A
	Histone H2B
Nucleus	a) Histone H2B
	b) H2B-like histone
	Histone H3
	Histone H4
Cytoplasm	Myeloid nuclear differentiation antigen
	S100A8 (component of calprotectin)
	S100A9 (component of calprotectin)
	S100A12
Cytoskeleton	Actin ( $\beta$ and/or $\gamma$ )
	$\alpha$ -actinin (1 and/or 4)
	Cytokeratin-10
	Myosin-9
	Plastin-2
Peroxisomes	Catalase
Glycolytic enzymes	$\alpha$ -enolase
	Transketolase

step signaling cascade. First, binding of one of the above factors to its receptors on neutrophils induces calcium influx. Elevated calcium leads to protein kinase C (PKC)

**Table 2.** NETosis inducers [3,12,13]

Microbes
<i>Aspergillus fumigates</i> , <i>Aspergillus nidulans</i>
<i>Candida albicans</i> , <i>Candida glabrata</i>
<i>Cryptococcus gattii</i> , <i>Cryptococcus neoformans</i>
<i>Eimeria bovis</i>
<i>Enterococcus faecalis</i>
<i>Escherichia coli</i>
<i>Helicobacter pylori</i>
HIV-1
Influenza A virus
<i>Klebsiella pneumoniae</i>
<i>Lactococcus lactis</i>
<i>Leishmania amazonensis</i> , <i>Leishmania donovani</i>
<i>Listeria monocytogenes</i>
<i>Mannheimia haemolytica</i>
<i>Mycobacterium tuberculosis</i>
<i>Pseudomonas aeruginosa</i>
<i>Salmonella enteric</i>
<i>Serratia marcescens</i>
<i>Shigella flexneri</i>
<i>Staphylococcus aureus</i>
<i>Streptococcus</i> (group A), <i>Streptococcus dysgalactiae</i> , <i>Streptococcus pneumoniae</i>
<i>Toxoplasma gondii</i>
<i>Yersinia enterocolitica</i>
Microbe products
Glucose oxidase
Lipophosphoglycan (LPG)
Lipopolysaccharide (LPS)
Host factors
Anti-neutrophil antibodies
GM-CSF + C5a
HMGB1
IL-8
Immune complexes
MIP-2
Platelet activating factor (PAF)
TNF
Others
Calcium ionophore (ionomycin)
Hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> )
Monosodium urate (MSU) crystals
Nitric oxide (NO)
Phorbol myristate acetate (PMA)

C5a: complement 5a, GM-CSF: granulocyte macrophage colony stimulating factor, HIV: human immunodeficiency virus, HMGB1: high mobility group box 1, IL-8: interleukin 8, MIP-2: macrophage inflammatory protein 2, TNF: tumor necrosis factor.

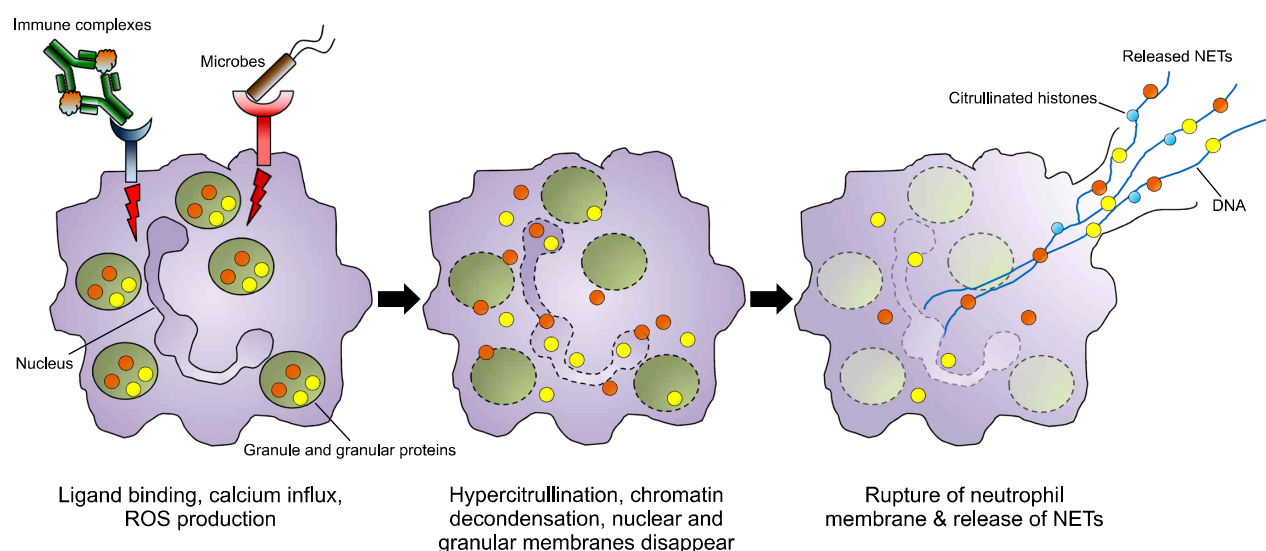
activation and phosphorylation of gp91<sup>phox</sup> (NOX2) by PKC allows the cytosolic and membrane-bound subunits of NADPH oxidase to assemble into a functional complex to produce reactive oxygen species (ROS) and nitric oxide (NO) [14,15]. For this reason, the most frequently used compound to induce in vitro NETosis is phorbol myristate acetate (PMA), a synthetic activator of the PKC family of enzymes [16].

The intracellular signaling induces several morphological changes in nuclear structure and granular membranes [5]. Initially, activated neutrophil undergoes chromatin decondensation and loses its characteristic multi-lobular structure of the nucleus. During NETosis, NE, a neutrophil-specific protease, is transferred from azurophilic granules to the nucleus and participates in histone modification and chromatin unfolding by degrading histone H1. In the next step, both nuclear and granular membranes disintegrate and additional granular proteins including MPO associate with chromatin. Eventually, the cell membrane ruptures and fully functional NETs are released from the cell (Figure 1) [3-5].

### Citrullination and NETosis

Protein citrullination is a post-translational modification of peptidyl arginine to citrulline, which is mediated by peptidylarginine deiminase (PAD) family of

enzymes. During NETosis, activation of NADPH oxidase is followed by activation of PAD4, which converts positively charged arginines in histones to neutral citrullines, thereby weakening the binding of histones to DNA and contributing to histone decondensation [17,18]. Histone citrullination by PAD4 is a prominent posttranslational modification in NETs and drives the major morphologic changes in NETosis [17,19,20]. Inhibition of PAD4 by Cl-amidine, a PAD inhibitor, disables NET formation [17] and neutrophils from mice deficient in PAD4 display impaired NETosis [20,21]. However, the implication of PAD4 in NETosis may be dependent on the stimulus. It has been reported that PMA induces NETosis without apparent activation of PAD4 and detectable histone citrullination [22]. It seems likely that PMA is qualitatively different from more physiologic stimuli to induce NETosis, because inflammatory and infection-related stimuli induce both histone citrullination and NETosis [22,23]. It is worthy to note that PAD4 is regulated by specific isoforms of PKC. While PMA activates PKC  $\alpha$ , an inhibitor of PAD4, many other stimuli including calcium ionophores activate PKC  $\zeta$ , an inducer of PAD4 [2,22]. Therefore, differential engagement of PKC isoforms induces differential histone citrullination by regulating PAD4 activation during NETosis.



**Figure 1.** NETosis. NETosis is initiated by binding of ligands such as immune complexes and microbes to their receptors on neutrophils, followed by calcium influx and reactive oxygen species (ROS) production. The intracellular signaling induces histone citrullination, chromatin decondensation, and disintegration of nuclear and granular membranes, which enables granular proteins to translocate to the nucleus. Subsequently, granular and cytoplasmic proteins are mixed with the chromatin. Finally, the neutrophil membrane ruptures and fully functional web-like neutrophil extracellular traps (NETs) are released.

## DAMPs and NETs

DAMPs are endogenous danger molecules released at the site of tissue damage under conditions of cell stress or tissue injury, which can initiate and perpetuate a non-infectious inflammatory response by activating the innate immune system [24]. DAMPs include intracellular proteins, such as histones, high-mobility group box 1 (HMGB1), and heat-shock proteins (HSPs), and purine metabolites, such as adenosine triphosphate (ATP) and uric acid, and mitochondrial components, such as formyl peptides and mitochondrial DNA [25]. Recognition of these DAMPs by pattern recognition receptors (PRRs) in innate immune system such as toll-like receptors (TLRs) and inflammasomes, induces inflammatory response [26]. Although DAMPs normally induce inflammatory responses to mediate host defense, they have also been implicated in multiple diseases including RA, atherosclerosis, sepsis, and cancer. [25,26].

NETs are composed of deleterious substances including histones and HMGB1, and these NET-derived DAMPs, in turn, can induce NETosis [27-29]. It has been reported that HMGB1 and histones released by injured hepatocytes stimulate NETosis through TLR4 and TLR9, and the development of NETs subsequently exacerbates inflammatory liver injury [27]. In addition, citrullinated histones and their immune complexes function as DAMPs, and citrullinated histone-containing immune complexes induce NETosis [30]. Therefore, tissue damage initiates the release of DAMPs which activate neutrophils to form NETs and, in turn, this NET formation generates additional DAMPs to exacerbate inflammatory responses through this vicious cycle.

## NETosis in rheumatoid arthritis

RA is a systemic autoimmune disease associated with chronic inflammation, mainly in the synovial joints, resulting in joint damage and loss of function [31]. The excessive and prolonged activation of the innate immune system contributes to the pathogenesis of autoimmune diseases including RA by establishing chronic inflammatory response. NETosis is a potent defense mechanism of neutrophils to protect human from invading pathogens [4]. However, dysregulation or defects of NETosis can lead to severe pathological consequences. Aberrant NETosis or impaired clearance of NETs plays a role in contributing to inflammation and tissue damage in autoimmune diseases [2]. NETs may promote production of autoantibodies by providing a source of citrullinated

autoantigens, thereby stimulating inflammatory responses in RA. Neutrophils from RA patients have displayed significantly enhanced NETosis, compared to neutrophils from healthy controls, and NET-releasing neutrophils have been shown to infiltrate RA synovial tissue [7]. Furthermore, RA sera and immunoglobulin (Ig) fractions from RA patients with high levels of anti-citrullinated protein antibody (ACPA) significantly induced NETosis and this effect was recapitulated when neutrophils were exposed to inflammatory cytokines interleukin-17A and tumor necrosis factor (TNF). More importantly, NETs augmented inflammatory responses and citrullinated autoantigens implicated in RA pathogenesis were externalized during NETosis. Citrullinated autoantigens can be produced by active extracellular PADs in RA which are released during neutrophil cell death [32]. Further support for the pathogenic role of NETosis in RA has also been reported. Citrullinated histones derived from NETosis reacted with ACPA-positive RA IgG and the immune complexes containing citrullinated histones activated macrophage to produce inflammatory cytokines and propagated NETosis [30]. These results suggest that accelerated NETosis perpetuates pathogenic mechanisms in RA.

## NETosis in systemic lupus erythematosus

NETosis is also involved in SLE by providing a source of autoantigens and inducing inflammatory responses. SLE is a severe autoimmune disease that affects skin and multiple organs, including kidney, lung, heart, and brain. It is characterized by autoantibodies to DNA, histones, ribonucleoproteins (RNPs) such as Ro and La, and ANCA antigens that are all NET components [2,33]. Recent evidence suggests that NETosis is associated with SLE. It has been reported that the clearance of NETs by serum endonuclease deoxyribonuclease I (DNase I) is essential for the progression of this disease. NET degradation was impaired in a subset of SLE patients due to the presence of DNase I inhibitors and anti-NET antibodies which prevent the access of DNase I to NETs [34]. In addition, SLE patients who have impaired NET degradation were at higher risk of developing nephritis. Further investigation has revealed that a low-density granulocyte subset expanded in SLE patients is prone to NETosis, and netting neutrophils infiltrate affected skin and kidneys, where they expose immunostimulatory molecules such as LL37 and dsDNA [35]. Furthermore, immune complexes that include the antimicrobial peptide LL37, human neu-

trophil peptide (HNP), and self-DNA, which are released during NETosis, activated plasmacytoid dendritic cells (pDCs) via TLR9 leading to interferon- $\alpha$  (IFN- $\alpha$ ) production [29,36]. The released IFN- $\alpha$  further activates pDCs by priming neutrophils to autoantibodies that induce NETosis. A recent study highlights the importance of NETs enriched in oxidized mitochondrial DNA in the pathogenesis of SLE [37]. Ribonucleoprotein immune complexes induce oxidized mitochondrial DNA release that elicits interferon-induced gene expression, and mitochondrial reactive oxygen species (ROS) scavengers suppress lupus-like disease in vivo [37].

### NETs as biomarkers and potential therapeutic targets

NETs may be useful biomarkers for NET-driven autoimmune diseases. For example, decreased NET degradation is associated with clinical manifestations in SLE [38]. Decreased NET degradation even preceded an increase in SLE Disease Activity Index (SLEDAI) score and proteinuria. Therefore, these results suggest that NET degradation can be used as a biomarker for predicting and assessing disease activity and renal involvement in SLE. However, this measurement only detects the relative changes of NETs among samples. Furthermore, the clinical heterogeneity of SLE may limit the usefulness of NETs as biomarkers for SLE. Future studies will need to establish standard and validated methods for measuring NETs as biomarkers for autoimmune diseases.

The inhibition of NET formation could be a potential therapeutic target for the treatment of autoimmune diseases. Treatment of ROS scavengers such as N-acetyl-cysteine (NAC) and MitoTEMPO blocks NET formation and suppresses SLE [6,37]. In addition, anti-malarials and DNase I may target NETs to reduce disease activity in SLE [6], and PAD inhibitors which target citrullination and NETs modulate disease activity in collagen induced arthritis and murine lupus [6,39]. Therefore, NETs may provide potential therapeutic targets to treat autoimmune diseases.

## CONCLUSION

Since the discovery of NETosis, it has come into the spotlight because of its potential pathogenic role in autoimmune diseases. NETs play a key role in host defense by trapping and killing microbes if they are released and cleared at the right place and time, otherwise dysregulation of the formation and clearance of NETs has detri-

mental effects by providing autoantigens and DAMPs. Although much progress has been made in recent years, it is still unclear why NETs from different stimuli are distinct, and why autoantibodies against NETs found in each autoimmune disease are unique. For example, the autoantibodies detected in RA patients are ACPA and rheumatoid factor (RF), whereas anti-nuclear antibodies (ANA), such as anti-dsDNA antibody, are the major autoantibodies found in SLE patients. In addition, NETosis induced by sterile inflammatory mediators such as immune complexes and TNF, which are involved in autoimmune diseases, is distinct from NETosis induced by microbial exposure. A better understanding of the molecular mechanisms regulating the release and clearance of NETs, and downstream signaling pathways that induce NETosis may allow for the development of new biomarkers and therapeutics for autoimmune diseases.

## ACKNOWLEDGMENTS

This work was supported by a 2-Year Research Grant of Pusan National University. I thank Christopher Rhodes and Annie HaEun Cho for proofreading.

## CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

## REFERENCES

1. Kolaczowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol* 2013;13:159-75.
2. Branzk N, Papayannopoulos V. Molecular mechanisms regulating NETosis in infection and disease. *Semin Immunopathol* 2013;35:513-30.
3. Pruchniak MP, Kotuła I, Manda-Handzlik A. Neutrophil extracellular traps (Nets) impact upon autoimmune disorders. *Cent Eur J Immunol* 2015;40:217-24.
4. Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, et al. Neutrophil extracellular traps kill bacteria. *Science* 2004;303:1532-5.
5. Fuchs TA, Abed U, Goosmann C, Hurwitz R, Schulze I, Wahn V, et al. Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol* 2007;176:231-41.
6. Barnado A, Crofford LJ, Oates JC. At the bedside: neutrophil extracellular traps (NETs) as targets for biomarkers and therapies in autoimmune diseases. *J Leukoc Biol* 2016; 99:265-78.
7. Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, Gizinski A, Yalavarthi S, Knight JS, et al. NETs are a source of citrullinated autoantigens and stimulate inflammatory

- responses in rheumatoid arthritis. *Sci Transl Med* 2013; 5:178ra40.
8. Urban CF, Ermer D, Schmid M, Abu-Abed U, Goosmann C, Nacken W, et al. Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against *Candida albicans*. *PLoS Pathog* 2009;5: e1000639.
  9. Kessenbrock K, Krumbholz M, Schönermarck U, Back W, Gross WL, Werb Z, et al. Netting neutrophils in autoimmune small-vessel vasculitis. *Nat Med* 2009;15:623-5.
  10. Remijsen Q, Kuijpers TW, Wirawan E, Lippens S, Vandenaabee P, Vanden Berghe T. Dying for a cause: NETosis, mechanisms behind an antimicrobial cell death modality. *Cell Death Differ* 2011;18:581-8.
  11. Papayannopoulos V, Zychlinsky A. NETs: a new strategy for using old weapons. *Trends Immunol* 2009;30:513-21.
  12. Cheng OZ, Palaniyar N. NET balancing: a problem in inflammatory lung diseases. *Front Immunol* 2013;4:1.
  13. Vorobjeva NV, Pinegin BV. Neutrophil extracellular traps: mechanisms of formation and role in health and disease. *Biochemistry (Mosc)* 2014;79:1286-96.
  14. Raad H, Paclet MH, Boussetta T, Kroviarski Y, Morel F, Quinn MT, et al. Regulation of the phagocyte NADPH oxidase activity: phosphorylation of gp91phox/NOX2 by protein kinase C enhances its diaphorase activity and binding to Rac2, p67phox, and p47phox. *FASEB J* 2009;23:1011-22.
  15. Hakikim A, Fuchs TA, Martinez NE, Hess S, Prinz H, Zychlinsky A, et al. Activation of the Raf-MEK-ERK pathway is required for neutrophil extracellular trap formation. *Nat Chem Biol* 2011;7:75-7.
  16. Kaplan MJ, Radic M. Neutrophil extracellular traps: double-edged swords of innate immunity. *J Immunol* 2012; 189:2689-95.
  17. Wang Y, Li M, Stadler S, Correll S, Li P, Wang D, et al. Histone hypercitrullination mediates chromatin decondensation and neutrophil extracellular trap formation. *J Cell Biol* 2009;184:205-13.
  18. Wright HL, Moots RJ, Edwards SW. The multifactorial role of neutrophils in rheumatoid arthritis. *Nat Rev Rheumatol* 2014;10:593-601.
  19. Neeli I, Khan SN, Radic M. Histone deimination as a response to inflammatory stimuli in neutrophils. *J Immunol* 2008;180:1895-902.
  20. Li P, Li M, Lindberg MR, Kennett MJ, Xiong N, Wang Y. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J Exp Med* 2010; 207:1853-62.
  21. Hemmers S, Teijaro JR, Arandjelovic S, Mowen KA. PAD4-mediated neutrophil extracellular trap formation is not required for immunity against influenza infection. *PLoS One* 2011;6:e22043.
  22. Neeli I, Radic M. Opposition between PKC isoforms regulates histone deimination and neutrophil extracellular chromatin release. *Front Immunol* 2013;4:38.
  23. Parker H, Dragnow M, Hampton MB, Kettle AJ, Winterbourn CC. Requirements for NADPH oxidase and myeloperoxidase in neutrophil extracellular trap formation differ depending on the stimulus. *J Leukoc Biol* 2012;92:841-849.
  24. Chen GY, Nuñez G. Sterile inflammation: sensing and reacting to damage. *Nat Rev Immunol* 2010;10:826-37.
  25. Piccinini AM, Midwood KS. DAMPening inflammation by modulating TLR signalling. *Mediators Inflamm* 2010; 2010:672395.
  26. Takeuchi O, Akira S. Pattern recognition receptors and inflammation. *Cell* 2010;140:805-20.
  27. Huang H, Tohme S, Al-Khafaji AB, Tai S, Loughran P, Chen L, et al. Damage-associated molecular pattern-activated neutrophil extracellular trap exacerbates sterile inflammatory liver injury. *Hepatology* 2015;62:600-14.
  28. Mitroulis I, Kambas K, Chrysanthopoulou A, Skendros P, Apostolidou E, Kourtzelis I, et al. Neutrophil extracellular trap formation is associated with IL-1 $\beta$  and autophagy-related signaling in gout. *PLoS One* 2011;6:e29318.
  29. Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, et al. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med* 2011;3:73ra20.
  30. Sohn DH, Rhodes C, Onuma K, Zhao X, Sharpe O, Gazitt T, et al. Local Joint inflammation and histone citrullination in a murine model of the transition from preclinical autoimmunity to inflammatory arthritis. *Arthritis Rheumatol* 2015;67:2877-87.
  31. Firestein GS. Evolving concepts of rheumatoid arthritis. *Nature* 2003;423:356-61.
  32. Spengler J, Lugonja B, Ytterberg AJ, Zubarev RA, Creese AJ, Pearson MJ, et al. Release of active peptidyl arginine deiminases by neutrophils can explain production of extracellular citrullinated autoantigens in rheumatoid arthritis synovial fluid. *Arthritis Rheumatol* 2015;67:3135-45.
  33. Tsokos GC. Systemic lupus erythematosus. *N Engl J Med* 2011;365:2110-21.
  34. Hakikim A, Fürnrohr BG, Amann K, Laube B, Abed UA, Brinkmann V, et al. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc Natl Acad Sci U S A* 2010;107:9813-8.
  35. Villanueva E, Yalavarthi S, Berthier CC, Hodgins JB, Khandpur R, Lin AM, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol* 2011;187:538-52.
  36. Lande R, Ganguly D, Facchinetti V, Frasca L, Conrad C, Gregorio J, et al. Neutrophils activate plasmacytoid dendritic cells by releasing self-DNA-peptide complexes in systemic lupus erythematosus. *Sci Transl Med* 2011;3:73ra19.
  37. Lood C, Blanco LP, Purmalek MM, Carmona-Rivera C, De Ravin SS, Smith CK, et al. Neutrophil extracellular traps enriched in oxidized mitochondrial DNA are interferogenic and contribute to lupus-like disease. *Nat Med* 2016;22: 146-53.
  38. Leffler J, Gullstrand B, Jönsen A, Nilsson JÅ, Martin M, Blom AM, et al. Degradation of neutrophil extracellular traps co-varies with disease activity in patients with systemic lupus erythematosus. *Arthritis Res Ther* 2013;15: R84.
  39. Willis VC, Gizinski AM, Banda NK, Causey CP, Knuckley B, Cordova KN, et al. N- $\alpha$ -benzoyl-N5-(2-chloro-1-iminoethyl)-L-ornithine amide, a protein arginine deiminase inhibitor, reduces the severity of murine collagen-induced arthritis. *J Immunol* 2011;186:4396-404.