

Short Communication

Successful surgical correction of anal atresia in a transgenic cloned piglet

Gab Sang Lee¹, Hye Soo Kim¹, So Hyun Lee¹, Dae Yong Kim^{3,4}, Kang Moon Seo⁵, Sang-Hwan Hyun⁶,
Sung Keun Kang^{1,3,4}, Byeong Chun Lee^{1,3,4,*}, Woo Suk Hwang^{1,2,3}

¹Departments of Theriogenology and Biotechnology, ⁴Pathology, and ⁵Surgery, College of Veterinary Medicine, Seoul National University, Seoul 151-742, Korea

²School of Agricultural Biotechnology, Seoul National University, Seoul 151-742, Korea

³Xenotransplantation Research Center, Seoul National University Hospital, Seoul 110-744, Korea

⁶Department of Veterinary Biotechnology, College of Veterinary Medicine, Chungbuk National University, Cheongju 361-763, Korea

Inbred strains of pig become indispensable for a wide range of biological studies. In biomedical science, it is generally accepted that somatic cell nuclear transfer (SCNT) technology with inbred strain of pig is essential for xenotransplantation. In this study, we observed the anal atresia in a cloned pig which was derived from fetal fibroblast of inbred miniature pig. A presumptive anal site of the cloned pig was excised and the rectum was sutured to apposed skin for treatment. This cloned piglet seemed to be normal with healthy status after surgery. This report can be useful for the treatment of anal atresia of cloned piglets.

Key words: anal atresia, cloned pig, miniature pig, nuclear transfer, transgenic pig

Successful somatic cell nuclear transfer (SCNT) of cultured cells, which was demonstrated in cattle [16], has provided an alternative for obtaining genetically modified pigs. McCreath *et al.* [11] reported the first success of obtaining gene-targeting sheep by using gene-targeted fibroblasts as a source of donor nuclei for SCNT. Since the birth of the first piglet from SCNT was reported in 2000 [14], tremendous progress has been made. Successful productions of pig resulting from random genetic modification *in vitro* followed by SCNT [5], as well as those with a specific modification (knock out) have been reported [9] by several groups in a short period. The production of cloned transgenic pigs is now in the transition from investigation to practical applications.

Anal atresia is a rarely recognized congenital disorder in the neonatal piglets, the incidence being estimated to be 0.1 to 1.0% [1]. A fistula develops in affected piglets between

anorectum and urethra and between anorectum and vagina, respectively. Congenital absence of the anus causes a build up of feces and consequent distension of the abdomen. The condition almost is invariably fatal within 2 to 3 days in males [12]. This report describes the successful treatment of anal atresia in a transgenic cloned piglet. It is known to be not worth attempting surgical repair local field because death invariably ensues without correct surgery and intensive care.

The SCNT and embryo transfer (ET): SCNT and ET were described in our previous study [4,9].

Treatment of anal atresia: For local anesthesia, 1 ml of 2% lidocaine solution (Xylocaine; Astra, Japan) was administered around presumptive anal region site. After, sufficient evacuation of the bowel, the rectal wall was apposed to the skin by interrupted sutures [6]. With assisting by pressure on the abdominal wall, gentle traction was used to draw the rectum caudal to the anus. Enrofloxacin 0.5 mg/kg/b.w. (Baytril; Bayer, Korea) was injected IM and surgery site is hip-bathed with 10% povidone-iodine up to 7 days post surgery (Fig. 1).

The piglet was nursied, and hydration status was assessed as normal. Antimicrobial and fluid therapies were continued for 7 days. The cloned piglet was seen to defecate normally. The transgenic cloned pig has been healthy and growing well without a history of complications for one year following surgery.

Since surgical correction, the cloned piglet seemed to be healthy and normal. No abdominal distension and difficulty in defecation were noted. To our knowledge, this is the first case of the anal atresia in cloned piglet. There are many reports on the congenital abnormalities of cloned mice and cattles [3,17]; however such data are sparse in cloned pigs [15]. We could not exclude any possibility that the anal atresia is directly related to SCNT since anal atresia occurs in normal piglet [12]. In the other hand, a previous study suggested that the atresia ani was genetically influenced by 2 major loci, 1 autosomal recessive and 1 autosomal

*Corresponding author

Tel: +82-2-880-1269; Fax: +82-2-884-1902

E-mail: bclee@snu.ac.kr

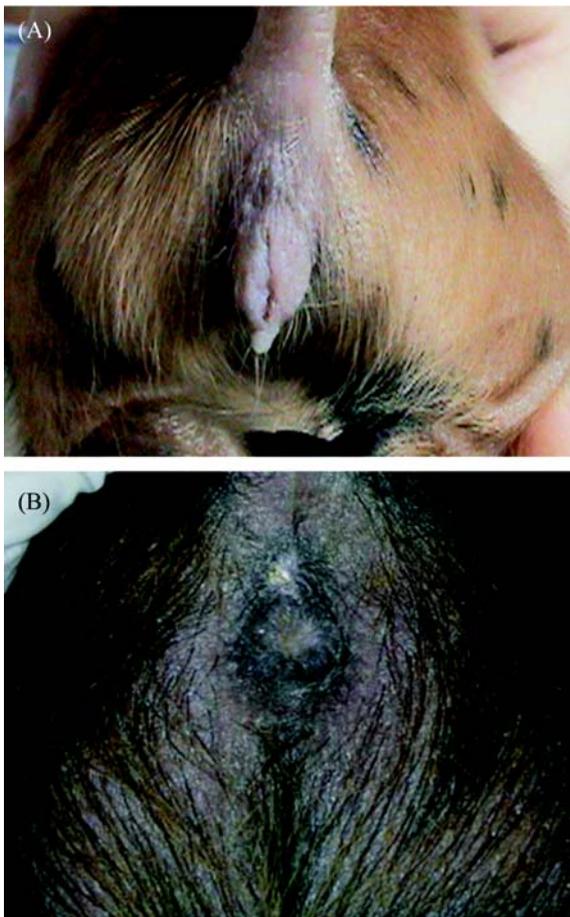


Fig. 1. The anal atresia in female cloned transgenic piglet before (A) and 40 days after surgical correction (B).

dominant [13]. Also the abnormality is known to be related with at least one or more recessive factor [2]. Additionally, mice that are homozygous for a targeted disruption of *Gli2* encoding a Shh-responsive transcription factor shows the imperforate anus and recto-urethral fistula [8]. In this study, the donor cells which provided a nuclear to SCNT embryos might have the high potential incidence for atresia ani, because this inbred miniature pigs are maintained for about 30 years [7]. In the previous study, anal atresia was observed in cloned pigs, but it is rarely seen in other reports related with pig somatic cell cloning [18]. We could not distinguish the reason of this malformation between the high incidence of original miniature pig line before inbreed and the genetic depression of inbreed strain. However, for xenotransplantation, pig inbred stocks for genetic modification to minimize graft rejection in the host would be advantageous over genetically diverse outbreed individuals. Therefore, the correction of abnormality in cloned pig is necessary. Also the screening of genetic defects or genetic uniformity is suggested before SCNT.

In that sense, it is critical that more numerous cases needed to be documented or investigated to evaluate

incidence and prevalence of congenital abnormalities in cloned piglets and compare those data to normally born piglets. This study shows the usefulness of aggressive surgical approach to keep the piglet alive. Cloned pigs must be cared and treated because they have invaluable potentials.

Acknowledgments

We thank Dr. Barry D. Bavister for his valuable editing of the manuscript. This study was supported by grants from the Ministry of Science and Technology (Top Scientist Fellowship) and Ministry of Agriculture and Forestry (Biogreen 21 #20050301-034-443-026-01-00), Korea. The authors acknowledge a graduate fellowship provided by the Ministry of Education through BK21 program.

References

1. **Furukawa T, Sawaguchi S, Ohkawa H.** Basic studies on anorectal malformations I. An examination of swine as an experimental animal model. *J Jpn Soc Pediatr Surg* 1982, **18**, 793-801.
2. **Hori T, Giuffra E, Andersson L, Ohkawa H.** Mapping loci causing susceptibility to anal atresia in pigs, using a resource pedigree. *J Pediatr Surg* 2001, **36**, 1370-1374.
3. **Humpherys D, Eggan K, Akutsu H, Friedman A, Hochedlinger K, Yanagimachi R, Lander ES, Golub TR, Jaenisch R.** Abnormal gene expression in cloned mice derived from embryonic stem cell and cumulus cell nuclei. *Proc Natl Acad Sci USA* 2002, **99**, 12889-12894.
4. **Hwang WS, Lee BC, Lee CK, Kang SK.** Human embryonic stem cells and therapeutic cloning. *J Vet Sci* 2005, **6**, 87-96.
5. **Hyun S, Lee G, Kim D, Kim H., Lee S, Nam D, Jeong Y, Kim S, Yeom S, Kang S, Han J, Lee B, Hwang W.** Production of nuclear transfer-derived piglets using porcine fetal fibroblasts transfected with the enhanced green fluorescent protein. *Biol Reprod* 2003, **69**, 1060-1068.
6. **Kersjes AW.** Atlas of Large Animal Surgery. pp. 50, Williams & Wilkins, Philadelphia, 1985.
7. **Kim YB, Huh ND, Koren HS, Amos DB.** Natural killing (NK) and antibody-dependent cellular cytotoxicity (ADCC) in specific pathogen-free (SPF) miniature swine and germfree piglets. I. Comparison of NK and ADCC. *J Immunol* 1980, **125**, 755-762.
8. **Kimmel SG, Mo R, Hui CC.** New mouse models of congenital anorectal malformations. *J Pediatr Surg* 2000, **35**, 227-230.
9. **Lai L, Kolber-Simonds D, Park KW, Cheong HT, Greenstein JL, Im GS, Samuel M, Bonk A, Rieke A, Day BN, Murphy CN, Carter DB, Hawley RJ, Prather RS.** Production of alpha-1,3-galactosyltransferase knockout pigs by nuclear transfer cloning. *Science* 2002, **295**, 1089-1092.
10. **Lee GS, Kim HS, Hyun SH., Lee SH., Jeon HY., Nam DH, Jeong YW, Kim S, Kim JH., Han JY, Ahn Curie, Kang**

- SK, Lee BC, Hwang WS.** Production of transgenic cloned piglets from genetically transformed fetal fibroblasts selected by green fluorescent protein. *Theriogenology* 2005, **63**, 973-991.
11. **McCreath KJ, Howcroft J, Campbell KH, Colman A, Schnieke AE, Kind AJ.** Production of gene-targeted sheep by nuclear transfer from cultured somatic cells. *Nature* 2000, **405**, 1066-1069.
12. **Ohkawa H, Sawaguchi S, Kaneko M.** Basic studies on anorectal malformations. II. Consideration on the significance of swine imperforate anus as a therapeutic model. *J Jpn Soc Pediatr Surg* 1984, **20**, 923-929.
13. **Ohkawa H, Sawaguchi S, Kaneko M.** Basic studies on anorectal malformations. III Studies on heredity of swine anorectal anomalies. *J Jpn Soc Pediatr Surg* 1986, **22**, 511-518.
14. **Polejaeva IA, Chen SH, Vaught TD, Page RL., Mullins J, Ball S, Dai Y, Boone J, Walker S, Ayares DL, Colman A, Campbell KH.** Cloned pigs produced by nuclear transfer from adult somatic cells. *Nature* 2000, **407**, 86-90.
15. **Prather RS, Hawley RJ, Carter DB, Lai L, Greenstein JL.** Transgenic swine for biomedicine and agriculture. *Theriogenology* 2003, **59**, 115-123.
16. **Sims M, First NL.** Production of calves by transfer of nuclei from cultured inner cell mass cells. *Proc Natl Acad Sci USA* 1994, **91**, 6143-6147.
17. **Young LE, Sinclair KD, Wilmut I.** Large offspring syndrome in cattle and sheep. *Rev Reprod* 1998, **3**, 155-163.
18. **Walker SC, Shin T, Zaunbrecher GM, Romano JE, Johnson GA, Bazer FW, Piedrahita JA.** A highly efficient method for porcine cloning by nuclear transfer using in vitro-matured oocytes. *Cloning Stem Cells* 2002, **4**, 105-112.