

## Review Article



# The Interplay between Host Immunity and Respiratory Viral Infection in Asthma Exacerbation

Ferdous Mohd Altaf Hossain <sup>1,2</sup>, Jin Young Choi<sup>1</sup>, Erdenebileg Uyangaa<sup>1</sup>,  
Seong Ok Park<sup>1</sup>, Seong Kug Eo <sup>1,\*</sup>

<sup>1</sup>College of Veterinary Medicine and Bio-Safety Research Institute, Chonbuk National University, Iksan 54596, Korea

<sup>2</sup>Faculty of Veterinary, Animal and Biomedical Sciences, Sylhet Agricultural University, Sylhet 3100, Bangladesh

## OPEN ACCESS

Received: Jun 9, 2019

Revised: Aug 22, 2019

Accepted: Aug 25, 2019

### \*Correspondence to

Seong Kug Eo

College of Veterinary Medicine and Bio-Safety Research Institute, Chonbuk National University, 79 Gobong-ro, Iksan 54596, Korea.  
E-mail: vetvirus@chonbuk.ac.kr

Copyright © 2019. The Korean Association of Immunologists

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

### ORCID iDs

Ferdous Mohd Altaf Hossain   
<https://orcid.org/0000-0003-4000-616X>  
Seong Kug Eo   
<https://orcid.org/0000-0001-9243-4268>

### Conflict of Interest

The funder had no role in the study design, data collection, data analysis, decision to publish, or preparation of the manuscript. The authors declare no potential conflict of interest.

### Abbreviations

AHR, airway hyper-responsiveness; AM, alveolar macrophages; APC, antigen-presenting cell; AR, airway reactivity; BAL, bronchoalveolar lavage; COPD, chronic

## ABSTRACT

Asthma is one of the most common and chronic diseases characterized by multidimensional immune responses along with poor prognosis and severity. The heterogeneous nature of asthma may be attributed to a complex interplay between risk factors (either intrinsic or extrinsic) and specific pathogens such as respiratory viruses, and even bacteria. The intrinsic risk factors are highly correlated with asthma exacerbation in host, which may be mediated via genetic polymorphisms, enhanced airway epithelial lysis, apoptosis, and exaggerated viral replication in infected cells, resulting in reduced innate immune response and concomitant reduction of interferon (types I, II, and III) synthesis. The canonical features of allergic asthma include strong Th2-related inflammation, sensitivity to non-steroidal anti-inflammatory drugs (NSAIDs), eosinophilia, enhanced levels of Th2 cytokines, goblet cell hyperplasia, airway hyper-responsiveness, and airway remodeling. However, the NSAID-resistant non-Th2 asthma shows a characteristic neutrophilic influx, Th1/Th17 or even mixed (Th17-Th2) immune response and concurrent cytokine streams. Moreover, inhaled corticosteroid-resistant asthma may be associated with multifactorial innate and adaptive responses. In this review, we will discuss the findings of various *in vivo* and *ex vivo* models to establish the critical heterogenic asthmatic etiologies, host-pathogen relationships, humoral and cell-mediated immune responses, and subsequent mechanisms underlying asthma exacerbation triggered by respiratory viral infections.

**Keywords:** Asthma exacerbation; Respiratory tract infection; Innate immunity; Adaptive immunity

## INTRODUCTION

Asthma is a frequent health concern globally associated with significant morbidity, hospitalization costs, and even fatal prognosis. Asthma is defined as a heterogeneous disease usually characterized by chronic airway inflammation (1) affecting almost 334 million people worldwide including up to 10% of adults and 30% of children, with an expected increase to around 400 million by the year 2025 (2). The heterogeneity of asthma depends on numerous risk factors, determinants, degree of airflow obstruction, hyper-responsiveness, severity and

obstructive pulmonary disease; DAMP, damage-associated molecular pattern; DC, dendritic cell; GC, glucocorticoids; HDM, house dust mite; ICAM-1, intercellular adhesion molecule-1; ICS, inhaled corticosteroids; ILC, innate lymphoid cell; IM, interstitial macrophages; iNOS, inducible nitric oxide synthase; IRF, IFN regulatory factor; LABA, long-acting  $\beta_2$  agonist; LPG, laboratory of genetics and physiology; MDA, melanoma differentiation-associated gene; NLR, Nod-like receptor; NSAIDs, non-steroidal anti-inflammatory drugs; OVA, chicken ovalbumin; PAMP, pathogen-associated molecular pattern; PRR, pattern recognition receptor; RIG, retinoic acid-inducible gene; RLR, retinoic acid-inducible gene-like receptor; RSV, respiratory syncytial virus; RV, rhinovirus

#### Author Contributions

Conceptualization: Hossain FM, Eo SK; Funding acquisition: Eo SK; Investigation: Eo SK; Validation: Choi JY, Uyangaa E, Park SO; Writing - original draft: Hossain FM, Eo SK; Writing - review & editing: Hossain FM, Choi JY, Uyangaa E, Park SO, Eo SK.

type of airway inflammation (3). Environmental triggers including allergens and pollen in the air and the diet, air pollutants, tobacco smoke, genetic differences, and respiratory viruses may exacerbate asthma, and play a vital role in worsening the outcomes (4). Approximately 100 major or minor genes may be associated with the development of asthma mediated via innate immunity, Th2 effector function, mucosal and lung functions along with airway remodeling (5). The immune responses and possible treatment strategies may be clear in classical (Th2 signature) asthma. However, in cases of severe and refractory asthma the multidimensional mechanisms and related immune response have yet to be defined. In non-Th2 response, the severity of augmented asthma and increased airway remodeling associated with poor prognosis have been correlated with a complex innate and adaptive immune response (6). Herein, the innate and adaptive immune signatures of asthma exacerbated by respiratory viral infection are discussed.

## ASTHMA EXACERBATION

Asthma exacerbation is characterized by higher and intense asthma severity, increased airway remodeling, a lower response to bronchodilator and anti-inflammatory treatments, resulting in pulmonary dysfunction. As a phenotypically heterogeneous disease, asthma involves multifaceted interactions between multiple risk factors and disease determinants (1,4). The diversity of asthma may be attributed to a complex interplay between risk factors (either intrinsic or extrinsic) and the specific disease determinants (viruses, bacteria, etc.). Rhinovirus (RV) is the major and most frequent determinant than respiratory syncytial virus (RSV), influenza, other corona viruses, or even pneumonia-inducing bacteria (7). Hospitalized patients manifesting asthma features carry a history of respiratory viral infections (mainly RVs) along with major extrinsic factors such as cigarette smoking, age, obesity, rhinosinusitis, pneumonic symptoms, and intolerance to non-steroidal anti-inflammatory drugs (NSAIDs) (8). The intrinsic risk factors (genetic polymorphisms, enhanced airway epithelial lysis, and apoptosis) are strongly correlated with exacerbation in asthmatic hosts, and exaggerated virus replication in infected cells may result in reduced innate immune response, and concomitant reduction in the expression of innate interferons (types I, II and III) (9,10). This subsection will highlight the etiology, the pathophysiological mechanism and ongoing treatment strategies targeting exacerbated asthma.

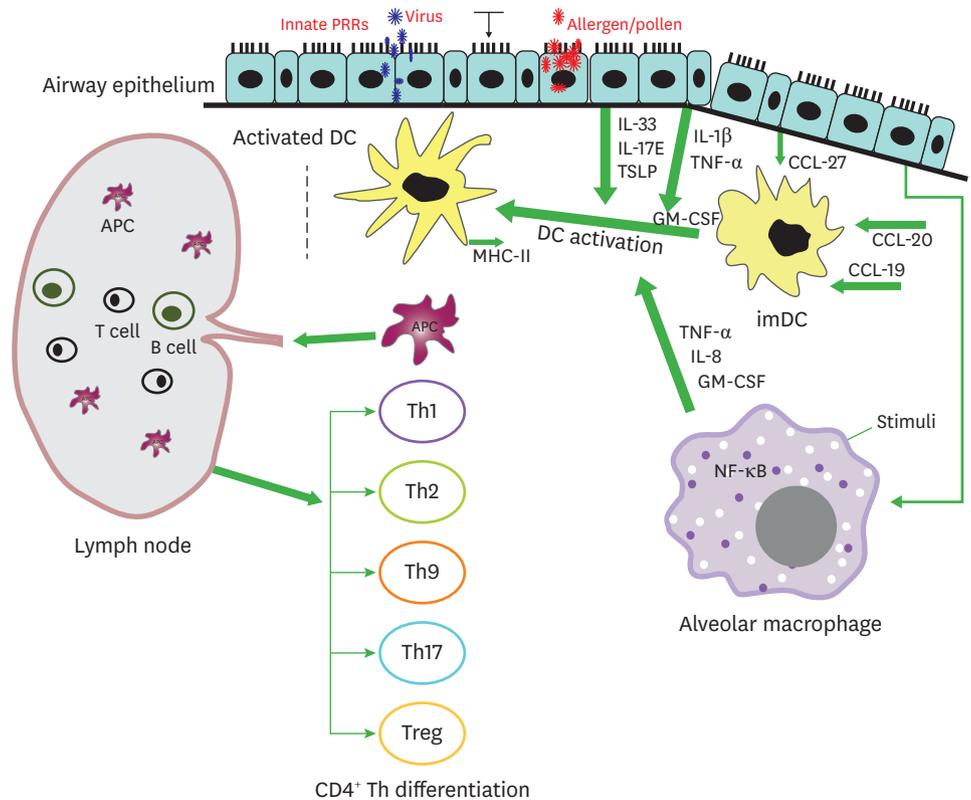
### Respiratory virus infections inducing asthma exacerbation

In addition to classical allergic asthma and delayed immune response, respiratory viral infections involved nearly 80% of cases diagnosed with hospitalized asthma exacerbation (11), including pediatric cases constituting nearly 63% of them (12). Among the respiratory viruses, RVs represent the most prevalent viral pathogens responsible for around 60% of virus-associated asthma exacerbations (7,13). Different murine models of experimental RV infection including chronic obstructive pulmonary disease (COPD), airway hyper-responsiveness (AHR) to histamine and subsequent lung inflammation were used to demonstrate the typical features of asthma (7,14). Emerging evidence supporting asthma exacerbation suggests a deficiency in antiviral immunity and loss of epithelial barrier integrity in individuals prone to severe refractory asthma (15). Although the exact mechanisms underlying asthma exacerbations have yet to be defined, studies suggest that RV infection of airway epithelium leads to cytokine interaction with allergic inflammation to overwhelm both innate and antigen-specific Th2 pathways resulting in enhanced Th2-related inflammation (7,16,17). Moreover, both in children and adults, a complex association between RV infection and asthma exacerbation may

also induce a non-Th2 immune response, along with increased AHR, mucus hypersecretion, airway remodeling, and respiratory failure (6).

**Clinical pathophysiology of asthma exacerbation**

RV-induced asthma is not only confined to the upper airway, but conclusive clinical evidence suggests that it is very common in the lower respiratory tract of children along with critical pneumonia signatures (8,14). In the respiratory airways, mucosal lining and cilia eventually trap airborne antigens and other harmful particles to prevent entry into the lung via mucociliary transport. Airway epithelium is exposed to foreign antigens, resulting in a cascade of secretory mechanisms such as mucus secretion, followed by secondary defense mechanisms mediated via cytokines and chemokines (18). The most cardinal features of asthma include inflammatory cellular infiltration, and mucus hyper-secretion along with AHR. Upon alveolar injury or pathogen invasion, the levels of patrolling immune cells, antigen-presenting cells (APCs), alveolar macrophages, and dendritic cells (DCs) are persistently elevated in conjunction with airway epithelium where leukocyte infiltration resolves subsequent inflammation (Fig. 1). The immune response starts with the activation of T lymphocytes by APCs, along with concomitant interaction between infiltrated cells, cytokines, chemokines and related mediators triggering and maintaining asthmatic airways (19). Differentiation



**Figure 1.** Interplay between innate immune cells in airways for adaptive immunity. The innate immune response in airway epithelium is initiated by contact with innocuous antigens, which triggers the expression of PRRs in lung epithelium. The activation of innate signaling receptors in airway epithelium results in migration of imDC to the mucosal epithelium via chemokine secretion from the epithelium. Following antigen stimulation, the AM are also activated to secrete GM-CSF, IL-8, and TNF-α that facilitate the activation of DCs. Moreover, epithelial cells release IL-1β and TNF-α, which induce the release of innate cytokines GM-CSF, IL-17E, TSLP, and IL-33 via binding to their respective receptors on the epithelium. The activation of airway DCs by the combined streaming of cytokines secreted by the epithelium promotes migration into draining lymph nodes, and induction of adaptive Th response. imDC, immature dendritic cells.

of Th0 cells into Th1, Th2, Th17, Th22, and Th9 cells is a complex phenomenon and poorly understood. Classical or eosinophilic asthma may exhibit a 'high Th2' molecular phenotype, and elevated serum IgE level indicating type 2 allergic mechanisms (20). The Th2 airway inflammation is characterized by eosinophilia and exhaled nitric oxide exacerbating the risk of asthma (21). By contrast, asthma exacerbation was characterized by Th1 response with neutrophil infiltration or even concurrent Th1-Th2-Th17 expression (22). Non-Th2 asthmatic pathophysiology has been linked to dysregulated innate immunity, including the activation of inflammasome pathways and development of airway neutrophilia (23). Recent studies investigating the mechanism of asthma exacerbation revealed a positive association between respiratory viral infections and activation of microRNAs and inflammasome-activating genes, and enhanced corticosteroid resistance (24,25)

### Treatments for asthma exacerbation

The efficacy of specific treatments and therapeutic agents targeting asthma has yet to be clearly defined. Most of the therapies are symptom-oriented and palliative based on severity. In general, current approaches include treatment with inhaled corticosteroids (ICS), long-acting  $\beta_2$  agonists, leukotriene receptor antagonists, inhaled IFNs, and anti-IgE therapies. Glucocorticoids (GC) are practicing as a mainstay to treat airway inflammatory disorders like rhinitis, chronic sinusitis, COPD and asthma. In addition to displaying well-known anti-inflammatory functions, GC is found to spare or enhance local innate response in respiratory epithelium (26). Treatment with anti-IL-5 (mepolizumab), anti-IL-13 (lebrikizumab), and even anti-IL4-R $\alpha$  (dupilumab) has been shown to reduce the levels of eosinophils and asthma exacerbation characterized by Th2 features (27). Generally, treatment of allergic airway inflammation, AHR, and respiratory virus-induced asthma with corticosteroids is ineffective, especially in RV-induced asthma exacerbation associated with neutrophil infiltration (11). Nearly 50% of cases showing exacerbated asthma involving neutrophil infiltration are not resolved with corticosteroid therapies (28). The etiology of corticosteroid-resistant asthma was strongly related to airway epithelial resistance via NF- $\kappa$ B and JAK activation, oxidative stress, and limited activity of transcriptional corepressor histone deacetylase-2 (29). Moreover, the ssDNA-like antisense agents including peptide-conjugated phosphorodiamidate oligomers (pleconaril and morpholino) are used as effective anti-enterovirus therapies against RVs by targeting highly conserved internal ribosome entry sites (30). Besides, inhaled IFNs including type I IFN (9), particularly IFN- $\beta$  (31) were used to alleviate respiratory virus-induced exacerbation of asthma symptoms (32). Therefore, appropriate therapies are urgently needed to manage asthma associated with non-eosinophilic or neutrophil-eosinophil mixed allergic asthma responses. Furthermore, corticosteroids diminish the response to CD4<sup>+</sup> Th9 cells in allergic asthma. The Th2-mediated asthma was controlled using corticosteroids via inhibition of the epithelial TARK (CCR4 ligand expressed by Th2 cells) (33). However, Th9 cells express functional CCR3, CCR6, and CXCR3, and corticosteroids may enhance CXCR3 ligand (IP-10) in asthmatic lungs, resulting in inadequate control of Th9-mediated asthma (34).

## SMALL ANIMAL MODELS FOR ASTHMA EXACERBATION INDUCED BY RESPIRATORY VIRAL INFECTION

Due to the logical limitations of human disease models, numerous small animal models, *in vitro* models, and mathematical equations have been widely used to study asthma pathogenesis. These models may have widened the scope of investigation into the

mechanistic pathways of asthma pathogenesis, and subsequent therapeutic discovery. Therefore, even though these models have some limitations, the hypothesis can be proved by analyzing all the models collectively and integrating their outcomes. In particular, respiratory virus-induced asthma exacerbation is a big concern, due to its clinical sequelae, and the intricate relationship between pathogenicity of respiratory viruses and defense mechanisms of model animals. Development of asthma models depends on the small animals (mice, rabbits, etc.) used and the viruses (RV, RSV, influenza, etc.) investigated. It also depends on the strain variation along with pre-exposure allergen sensitization (chicken ovalbumin [OVA], house dust mite [HDM], etc.). The results are similar to acute or chronic asthma models. Influenza virus infections in HDM- or OVA-sensitized mice usually induce critical features of asthma, whereas influenza and RSV models may fail occasionally due to the overwhelming cytokine response and canonical eosinophilic trafficking (35). Further, murine models, particularly those involving allergic inflammation (OVA, HDM) or RV-induced exacerbations are associated with varying levels of advantages and disadvantages from a scientific perspective (36). Many murine models of asthma associated with RV infection have failed, probably due to RV serotypes, a majority (90%) of which binds to human intercellular adhesion molecule-1 (ICAM-1) but not the mouse ICAM-1. By contrast, 10% of RVs such as RV-1B use different adhesion molecules and bind to the mouse low density lipoprotein receptors (17). Most of the murine models show inflammatory changes and typical asthma features resembling human bronchial obstruction, AHR, epithelial hypertrophy, goblet cell metaplasia, infiltration of innate cells, trafficking of Th cells and concurrent cytokine-chemokine interactions (37,38). Basically, models of acute and chronic inflammation are highly specific to allergens (OVA, HDM, cockroach extract, etc.), murine strains, sensitization, and virus exacerbation protocols used (39).

## INTERPLAY BETWEEN HOST IMMUNITY AND RESPIRATORY VIRAL INFECTION IN ASTHMA EXACERBATION

Respiratory viral infections linked to asthma pathogenesis may differ in outcomes, depending on the related risk factors. The common cold virus, RV, has emerged as the primary etiological agent associated with exacerbations and other pathological outcomes of asthma. After the onset of RV attacks, the typical or exacerbated asthma features develop following the activation of different innate responses. The critical pathogenesis of RV-exacerbated asthma is triggered by impaired respiratory epithelial function inducing the release of variable proinflammatory mediators to attract airway inflammatory cells like eosinophils, neutrophils, macrophages, and lymphocytes (40).

### Role of innate immunity in asthma exacerbation caused by respiratory viral infection

The host innate immune response is characterized by critical mechanisms for prompt detection, binding, and elimination of invading pathogens. The innate immune system is fortified with various pattern recognition receptors (PRRs) such as TLRs, retinoic acid-inducible gene (RIG)-like receptors (RLRs), and Nod-like receptors (NLRs) for recognition and response to pathogen-associated molecular patterns (PAMPs). Here we will focus on the mechanisms of different PRRs against respiratory viruses, and innate cells to orchestrate innate immune response in asthma development.

### *PRRs in allergic airway inflammation*

Recognition of allergens or respiratory viral infections (such as RVs) by PRRs is followed by enhanced secretion of inflammatory cytokines including types I and III IFNs (41). RV infection-increased numbers of subepithelial IFN/PRR-expressing inflammatory cells are related to RV loads in airways and concurrent severity of asthma (42). TLRs are transmembrane and noncatalytic PRRs expressed on macrophages, DCs, fibroblasts, epithelial or other non-immune cells (43). The initial innate responses are regulated by TLRs via activation of TRIF and MyD88 signaling pathways, ensuring the synthesis of inflammatory mediators via NF- $\kappa$ B pathway (44). The asthmatic conditions associated with neutrophilic trafficking are characterized by the increased threshold of IL-8 and IL-1 $\beta$ , and upregulation of TLR2 and TLR4 via MyD88 and TRIF pathways followed by NF- $\kappa$ B and MAPK signal activation. Besides, TLR2 and TLR4 are associated with lung dysfunction, and pediatric asthma (45). Following the initiation of TLR4 activation, pro-inflammatory cytokines are released to attract innate cells (macrophages, mast cells and neutrophils), and eventually leading to ROS production (46). TLR7 activation is strongly associated with the induction of Th1 axis, masking the development of classical Th2 immunity, and IgE assembly in response to allergen-induced airway inflammation or RV-exacerbated asthma (47). The RLR family comprises RIG-I, melanoma differentiation-associated gene (MDA) 5, and laboratory of genetics and physiology (LPG), which have been shown to mediate the innate immune response against double-stranded RNA (48). The RLR-induced signaling results in a robust production of type I IFN via activation of NF- $\kappa$ B, MAP kinase and IFN regulatory factor (48). Furthermore, RLRs (RIG-I and MDA5), which could be induced initially through TLR3/TRIF pathway, show the coordinated role with TLR3 in exhibiting innate defense to RV (41).

Currently, asthmatic features are highly linked to NLR-dependent maturation and secretion of pro-IL-1 $\beta$  and IL-18. NLRs can be activated by physical damage to the plasma membrane induced by respiratory viruses and host-derived damage-associated molecular patterns (DAMPs), which stimulates the caspase 1-activating platform (inflammasome) that plays a vital role in host defense against RV (25,49). RV infection activates NLRP3 and NLRC5, which leads to the activation of caspase 1 and the hyper-production of IL-1 $\beta$  (50). Thus, in response to allergens or respiratory virus infections, the PRRs (TLRs, RLRs, and NLRs) play a crucial role in the recognition of viruses, and subsequent induction of innate response mediated via inflammatory cytokines and stimulation of IFNs.

### *Innate response of airway epithelial cells*

Airway epithelium, the first-line barrier to external environment and maintenance of internal milieu, expresses multiple PRRs for rapid detection and response via release of microbial PAMPs or DAMPs under cellular stress, tissue damage, necrosis, and cellular death (5,51). Airway epithelium detects hRV infection via TLRs, RIG-I and MDA5, followed by activation of signaling pathways that lead to the synthesis of pro-inflammatory cytokines and type I IFNs (41,52). Upon RV infection, airway epithelium may express type I IFN- $\beta$  and type III IFN- $\lambda$  resulting in defective epithelial apoptosis during exacerbation (9,10,53). Airway epithelium orchestrates and triggers airway inflammation via release of CCL2 and CCL20 in response to allergen stimuli. The airway epithelium activates the immunomodulatory role of DCs via release of multiple cytokines (TSLP, IL-25, IL-33, SCF, GM-CSF, etc.) following activation of TLRs in response to external stimuli and allergens (54). Simultaneous activation of DCs may induce the release of CCL17 and CCL22 mediating Th2 response via CCR4, followed by the synthesis of RANTES, Eotaxin-1, and Eotaxin-2 to attract eosinophils via CCR3 activation pathway. Specifically, TSLP promotes growth and differentiation of basophils to induce IgE

class switching controlled by IL-4; however, IL-17E (IL-25) and IL-33 may activate nuocytes to induce simultaneous expression of IL-5 and IL-13 for eosinophilic trafficking, goblet cell metaplasia, and AHR development triggered by the virus (55). In addition to cytokine synthesis, the respiratory epithelium may produce DAMPs (ATP, uric acid, ROS, etc.) to regulate the immune response in atopic conditions or even a single exposure to allergens due to activation of DCs and other innate cells via NF- $\kappa$ B signaling pathway (56). Moreover, in response to antigen exposure, the airway epithelium secretes prostaglandin E<sub>2</sub> to diminish DC reactivity by binding to the EP4 receptor, and thus facilitate the suppression of airway inflammation (57).

#### *Eosinophils*

In allergic inflammation and respiratory virus-induced asthma, the infiltration of key innate cells including eosinophils and neutrophils explains asthma pathogenesis individually or via mixed immune response (58). Eosinophils play an innate antiviral role via production and secretion of neurotoxin, ROS, and eosinophilic cationic proteins, cytokines and lipid mediators. Thus, eosinophils exhibit multiple roles such as epithelial barrier function, intrinsic interaction between innate and adaptive immunity, and tissue remodeling (59). Following RV infection, the infiltrated eosinophils also secrete cationic proteins and leukotrienes, thereby resulting in AHR upon activation of eosinophils following the secretion of RANTES by the airway epithelium (15). However, increased levels of nasal or BAL eosinophils are related to allergic airway inflammation, and respiratory viral infections triggering either acute or chronic asthma (17,38,39,60). In human respiratory infections, and various murine models of asthma, the eosinophils act as APCs to activate naïve CD4<sup>+</sup> Th cells via surface markers MHC-II and CD40 in response to stimulation by cytokines including IFN- $\gamma$ , GM-CSF, IL-3, IL-4, and IL-5 (61).

#### *Neutrophils*

Airway neutrophilic trafficking occurs early as a first-line defense after the onset of respiratory virus-infections and concomitant asthma exacerbation (52). Neutrophils release chemotactic factors and granular proteins to activate monocytes and macrophages in the airway and induce an innate response (62). Despite the innate host defense, the activated neutrophils exhibit immunoregulatory roles to secrete ROS, dsDNA and elastase containing neutrophil extracellular traps (NETs) via programmed cellular death known as NETosis (63). In adults, the exacerbation of virus-induced asthma is characterized by an influx of sputum neutrophils and neutrophil *elastase*, responsible for airway obstruction, mucus secretion, and lower respiratory tract symptoms (64). Moreover, the neutrophil-mediated dsDNA may enhance the expression of Th2 cytokines (IL-4, IL-5, and IL-13) in nasal or BAL fluid, which mediate the classical type 2 immune response (60,63). In acute asthma exacerbations by RVs, neutrophil infiltration occurs most predominantly in sputum or lungs, and is strongly induced by IL-8 and G-CSF (17). Several murine models suggest classical and neutrophil-eosinophil interaction in BAL and inflamed asthmatic lungs (7,14,38); however, the exact mechanism underlying the role of neutrophils in asthma has yet to be established.

#### *Macrophages*

Macrophages, DCs, and monocytes constitute the mononuclear phagocytic network in the host innate response to airborne antigens. The macrophages are the most abundant innate immune cells in the airway lumen, and are involved in the early phase of host defense against RV exposure (37). In asthma pathogenesis, macrophages play a vital role in coordinating adaptive immunity, regulation of airway inflammation, and subsequent tissue repair (52). In

early allergy response, the influx of very short-lived proinflammatory monocytes, interstitial macrophages (IM) and airway macrophages (AM) appears to mediate early- and late-phase activation and differentiation (65). The innate response of resident AM is initiated by the secretion of IL-10, and IL-12 (anti-Th2) cytokines (66). In contrast, the IM regulates the DC function via IL-10 followed by neutrophil infiltration and subsequent attenuation of Th2 response to allergen exposure and other determinants of airway inflammation (66). RV infection also induces the release of IFN- $\alpha$  from AM to maintain the antiviral status of the airway epithelium and facilitate homeostasis of the lung (67). Thus, the allergen-primed macrophages play a protective role against tissue damage, and the asthmatic mice show increased allergic inflammation and AHR upon depletion of those macrophages. Several *in vivo* murine and *ex vivo* models corroborated the intense induction of CCL11 and IL-4 expression in the AM via phenotypic changes in response to allergen and RV exposures (37). The classical (M1) polarized macrophages utilize transcription factors STAT1/NF- $\kappa$ B, and are activated via IFN- $\gamma$ , GM-CSF, LPS, to secrete IL-1, IL-8, IL-12p40, inducible nitric oxide synthase (iNOS), IFN- $\gamma$ , TNF- $\alpha$ , and MIP-1 $\alpha$ . In contrast, the M2-polarized macrophages express a different set of phagocytic receptors, and are activated by M-CSF, IL-4, IL-8, and IL-13 resulting in the secretion of arginase, IL-10, and TGF- $\beta$ , using the transcription factor STAT3/STAT6 (68). A few murine and human models also demonstrated the sequential differentiation of M2-polarized macrophages in both RV-induced acute and chronic asthma via CCL11, and CCL17-dependent eosinophilic expression (37,69).

#### $\gamma\delta$ -T cells

In asthma, the  $\gamma\delta$ T cells are strongly associated with epithelial tissue homeostasis, modulating both antiviral innate and cell-mediated immune responses. They facilitate a neutrophil-mediated host defense mechanism against respiratory infection (70). The murine asthma models were used to demonstrate the role of  $\gamma\delta$ T cells in AHR, and prime the antiviral innate response to RV-induced asthma exacerbation (71). Moreover, in animal models of OVA-induced allergic asthma, the  $\gamma\delta$ Treg associated with immunosuppressive function express both Foxp3 and TCR $\gamma\delta$  (72). In humans, upon RV or allergen exposure, the critical features of asthma such as increased viral load, airway obstruction, AHR, and exaggerated eosinophilia in BAL, are strongly associated with  $\gamma\delta$ T-cell population in the airway and the blood (71). In the murine asthma model, the  $\gamma\delta$ T cells attenuate allergic airway inflammation. By contrast, a few reports suggested the insignificant effect of anti- $\gamma\delta$ TCR treatment on pulmonary viral load, IFN- $\beta$  response, and RV-specific serum IgG (71). Therefore, multidimensional *ex vivo* and *in vivo* models are needed to elucidate the immunoregulatory roles of  $\gamma\delta$ T-cells.

#### Innate lymphoid cells (ILCs)

In airway mucosal tissues, ILCs play a critical role in host defense and tissue homeostasis, and facilitate the interaction between innate cellular signals and adaptive immune response (73). Based on the response to innate cellular signals (TSLP, IL-17A, IL-33, IL-1 $\beta$ , and TGF- $\beta$ ) generated by the asthmatic airway epithelium, ILCs can be subdivided into ILCs-1, 2, and 3, depending on the cytokines expressed (74). Cytokine synthesis in ILCs may be linked to macrophage polarization (M1 or M2) to orchestrate asthma pathogenesis (75). The roles of ILCs in asthmatic airways vary depending on the type of differentiation, cytokine production, and asthma pathogenesis. Usually, ILCs express and secrete high levels of Th1, Th2 and Th17 cytokines, whereas ILC2 shows an eosinophilic Th2 asthmatic signature via induction of IL-4, IL-5, IL-9, and IL-13 in response to IL-25 and IL-33 secreted by the asthmatic epithelium (76). However, ILC1 and ILC3 coordinate with other innate cells, or neutrophils although a few reports suggested that ILC1 secretes IFN- $\gamma$  via T-bet as an innate complement to CD4<sup>+</sup> Th1

cells (77). Besides, in some animal and human models, ILC3 secretes IL-17A, TNF- $\alpha$ , and IL-22 to exacerbate asthma pathogenesis (78).

### Role of IFNs in innate immunity during asthma

The innate IFNs in asthma boost host antiviral defenses, resulting in reduced RV load and RV-induced acute exacerbation, although the precise mechanisms are unknown. Based on the tropism for different receptors, the type I IFNs consist of a single IFN- $\beta$ , and 14 IFN- $\alpha$  subtypes; the type II includes the IFN- $\gamma$  subtype, and the type III IFNs comprise IFN- $\lambda$  1, 2, 3, and 4 subtypes. Type I and III IFNs share a similar role in innate anti-viral immunity (79). Usually, airway epithelium and hematopoietic cells produce type I and III IFNs, whereas type II IFN (IFN- $\gamma$ ) secretion is restricted to Th1, NKT,  $\gamma\delta$ T cells, macrophages, DCs, and NK cells.

#### Type I IFNs

Notably, type I IFNs are only induced after RV exposure to airway epithelium or *in vitro* stimulation of coherent cells with either type I IFNs or specific TLR agonists. The type I IFNs signal via IFNAR in conjunction with ISG induction to yield an anti-RV phenotype (80). Type I ( $\alpha/\beta$ ) IFNs in airway epithelium or sub-epithelial innate cells (macrophages) play a critical role in combating RV loads and replication due to enhanced apoptosis of infected cells, IgE production, and concurrent exacerbation of Th2 responses in allergen-RV-induced asthma by inhibiting the production of CCL17, eotaxin-1, and eotaxin-2 (60,81). Basically, type I IFNs (IFN- $\alpha/\beta$ ) suppress Th2 cytokines via inhibition of IL-5 synthesis by CD4<sup>+</sup> T cells, and interfere with the infiltration of eosinophils and CD4<sup>+</sup> T cells into RV-infected areas (9). Despite the different mechanisms involved, type I and III IFNs share a similar role in the innate response against viral infections (79). A guinea pig model has been used to demonstrate the inhibitory role of type I IFN against the development of eosinophilic asthma after repeated OVA challenges (82). Similar mechanisms in human RV-infected lung epithelium treated with type I (IFN- $\beta$ ) and type III IFN- $\lambda$ 1 reduced the viral load and limited the viral replication (83).

#### Type II IFN

Type II IFN (IFN- $\gamma$ ) mimics a wide range of immunomodulatory activities compared with type I IFNs, with minimal antiviral innate response. IFN- $\gamma$  upregulates the expression of MHC class I and II on the innate cells, particularly macrophages, and neutralizes the antigens presented by mast cells, the critical mediators of allergic airway inflammation. Moreover, IFN- $\gamma$  may downregulate CCR3 expression (via eotaxin-1 and -2 receptors) and CXCL9 during induction, and the progenitor hematopoietic cells are regressed during the differentiation and activation of eosinophils (84). Several *in vivo* murine models also echoed the regulatory roles of allergen-induced asthma pathogenesis, and IFN- $\gamma$ -receptor-ablated (GRKO) mice showed inhibition of prolonged eosinophilia via recruitment of CD4<sup>+</sup> T cells and IL-5 induction (85). Besides, IFN- $\gamma$  plays an important role in iNOS synthesis, resulting in IgE-mediated degranulation of mast cells, and inhibition of hyperplasia and hypertrophy of airway smooth muscle cells (86). Due to the anti-IgE class switching ability of IFN- $\gamma$ , neutralization of inhaled allergen via Fc $\gamma$ R-mediated endocytosis may epitomize the allergen capture mechanism of macrophages, and subsequent apoptosis of T cells and eosinophils via influx of proapoptotic proteins, caspases and Bax (87). Thus, the compelling inhibitory role of IFN- $\gamma$  in allergic asthma demonstrates a definitive therapeutic approach, suggesting the need for further mechanistic studies using both *in vivo* and *ex vivo* models.

#### Type III IFNs

The innate roles of IFN- $\lambda$  in RV-induced airway inflammation are mediated via a regulatory

response in severe and exacerbated asthma (10). Type III IFNs (IFN- $\lambda$  1 and IFN- $\lambda$  2/3) display similar features, and share analogous roles in innate immunity after RV infection. RSV infection induces type III IFNs which activates Jak-Stat pathways to exhibit antiviral effect through ISG gene expression (88). Upon viral infection, TLR3, TLR4, TLR7, TLR8, and TLR9 mediate IFN- $\lambda$  induction, and TLR3-mediated signaling is associated with RV-induced asthma severity and concurrent deficiency of IFN- $\lambda$  (89). The *ex vivo* RV infection of isolated human bronchial epithelial cells and BAL cells derived from asthmatic subjects result in impaired IFN- $\lambda$  synthesis (90). Moreover, the beneficial effect of IFN- $\lambda$  in allergic asthma is clearly demonstrated using a murine model, where IL-28R $\alpha^{-/-}$  mice develop enhanced airway inflammation, increased mucus production, and a Th2-Th17-dependent cytokine response along with a reduction in IFN- $\gamma$  and IL-12-expressing Th1 cells (91). Therefore, IFN- $\lambda$ , despite lacking the multi-dimensional activity of type I IFNs, represents a potential candidate in combating RV-induced asthma exacerbation. Additional immunomodulatory mechanisms of type III IFNs should be investigated in different *in vivo* models.

### Role of adaptive immunity in asthma exacerbation caused by respiratory viral infection

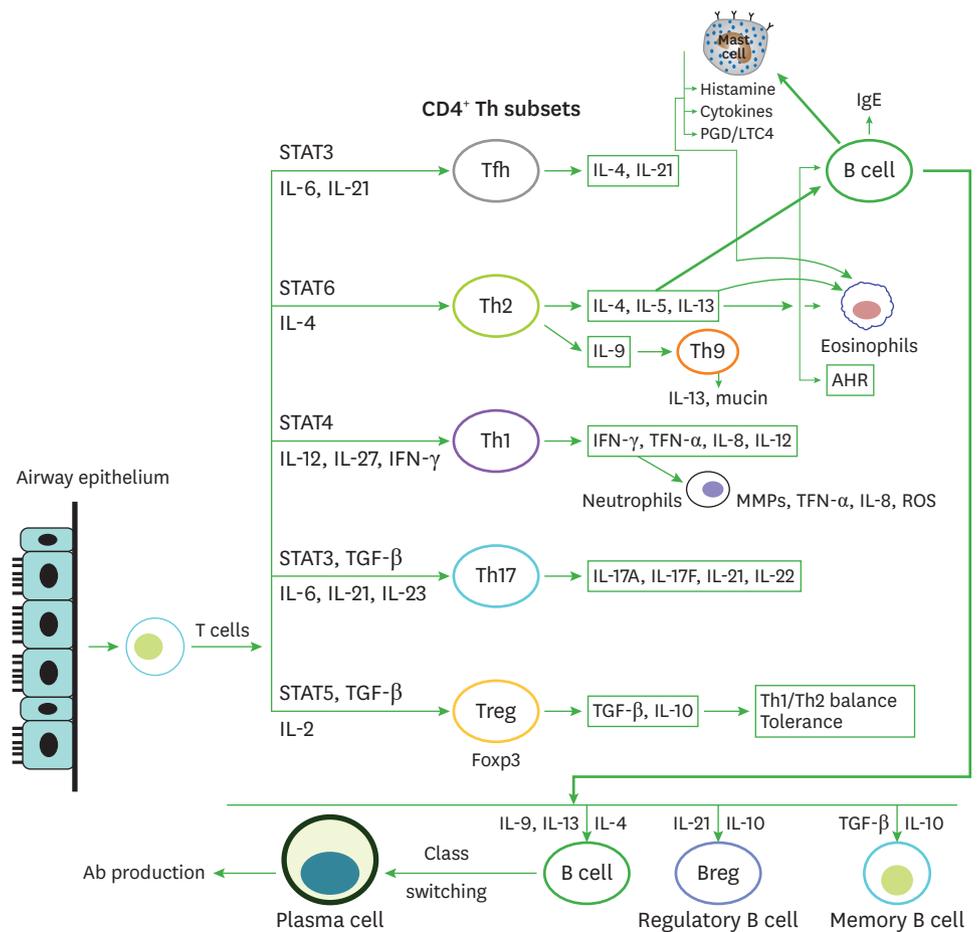
Cell-mediated immunity represents a comprehensive and more refined parameter for the recognition of self- and non-self-antigens, allergens and pathogens via regulation of interplay between APCs and lymphocytes (T and B cells). Thus, the adaptive immune response enhances antigen-specific immunological mechanisms to generate effector memory T cells and lead to concurrent immune homeostasis in the host.

#### B cells

The B cells have been the focus of attention following the detection of a series of B-cell-mediated immune responses in asthma, multiple sclerosis, and autoimmune diseases (92). In allergic inflammation or respiratory virus-induced asthma, B cells induce Ig synthesis effectively to overwhelm asthma progression. Isotype switching of B cells to allergen-specific IgE synthesis is highly dependent on IL-4 and IL-13 (via interaction with BCR), and provides the critical triggers during airway inflammation (93). The allergen-specific IgE interacts with low-affinity Fc $\epsilon$ RII (CD23) receptor on B cells, and enhances both allergen-specific B and T cell-mediated recall in inflamed lungs (94). A cockroach allergen-induced allergy model showed a B cell-dependent prolongation of chronic pathogenesis mediated via Th2 phenotypes (95). The effector function of B cells is attributed to Be1 (B cells primed by Th1 cells) and Be2 (B cells primed with Th2 cells) cells, resulting in the differentiation of Th0 cells to IFN- $\gamma$ -secreting Th1 and IL-4-dependent Th2 cells, respectively (96). Moreover, B lymphocytes induced T-cell-independent AHR and airway inflammation. The regulatory B cells manifested reversible inhibitory effects against allergic asthma via CD4<sup>+</sup>FoxP3<sup>+</sup>Treg cells in murine models (97).

#### T cells

The role of T cells in allergic airways and asthma exacerbation is mediated via complex mechanisms due to the different patterns of T cell subsets and effector functions involved in airway inflammation, remodeling, mucous cell metaplasia, intense and airway reactivity (AR), and less in prognosis. Notably, the function of CD4<sup>+</sup> T cells is well known in allergic asthma based on their cytokine profiles; however, the CD8<sup>+</sup> T cells also display an important protective or suppressive role in lungs. However, the functional role (regulatory/NK/proinflammatory) of CD8<sup>+</sup> T cells needs to be characterized adequately. In inhaled allergic asthma, the CD8<sup>+</sup> T cells exhibit proinflammatory, protective, and regulatory functions (83).



**Figure 2.** Contribution of CD4<sup>+</sup> Th cells to airway inflammation. APCs including DCs process antigen (allergens/ viruses) and present them on naïve T cells, which subsequently induces the differentiation of naïve T cells into Th1, Th2, Th17, and Tregs cells, depending on the microenvironment orchestrated by the cytokine milieu. Following exposure to allergens, the Th2 response facilitates the accumulation of eosinophils in the airway via production of IL-4, IL-5, IL-9, and IL-13, whereas Th1 response is linked to mixed (eosinophil-neutrophil) inflammation via induction of IFN- $\gamma$ , IL-8, TNF- $\alpha$ . Th17 cells mediate neutrophilic inflammation that is frequently observed in severe refractory asthma. CD4<sup>+</sup>Foxp3<sup>+</sup> Tregs secreting IL-10 and TGF- $\beta$  coordinate the tolerance to inflammation and balance of Th1/Th2 response.

After the onset of allergic inflammation or exacerbation with respiratory viruses, the CD4<sup>+</sup> T helper (Th) cells display an adaptive immune response, and differentiate into effector T cell subsets secreting various cytokines (**Fig. 2**).

In asthma pathogenesis, a robust and complex mechanism of differentiation and initiation of Th1, Th2, and Th17 cytokines or even Th9 response has been established using *in vivo* and *in vitro* models (17,38,58,60). In airway inflammation and allergic asthma, Th1 and Th2 responses are highly restricted via activation of transcription factors T-bet and GATA3, respectively (98) with subsequent cytokine production (99). The canonical Th2 response is mainly characterized by the production of IL-4, IL-5, and IL-13 (99); however, the IFN- $\gamma$ -restricted Th1 response is proved to be amalgamated with Th2 cytokines (IL-4, IL-5, and IL-13), in chronic allergic responses (100). The exact mechanism underlying the differentiation of Th1 or Th2 response in atopic asthma or respiratory virus-induced exacerbation remains controversial, despite the balancing mechanisms of pro-inflammatory and suppressive Tregs (100). Moreover, Tregs provide insight into T-cell programming and flexibility during the

activation of transcription factors to augment or repress cytokine gene expression during a stable Th cell response.

The most unpredictable (Th1 or Th17) features of severe asthma relate to the Th17 cell repertoire induced by IL-6, IL-22, and TGF- $\beta$  mediated via expression of the transcription factor ROR- $\gamma$ t in several *in vivo* and *ex vivo* models of Th17 cells secreting IL-17A and IL-17F, and IL-25 (101). Recently, the Th17 cells were identified as prominent immune cells in airway inflammation of tissues during severe and chronic allergic asthma associated with steroid resistance. The most prominent and critical cytokine IL-17A binding to the IL-17R complex in the airway epithelium may act via increased NF- $\kappa$ B activation and induction of neutrophil chemokine, CXCL8 (102). The increased interest in the role of Th17 cells in severe asthma has been driven by their simultaneous appearance in classical Th2-mediated inflammation. Therefore, the Th17 immune responses can be orchestrated either alone or depending on Th1 or Th2 features, and also reciprocally with Th2 response (103). In addition to these unpredictable phenomena, IL-17A acts as a negative regulator of asthma by decreasing eosinophilic infiltration into the airways, AR, and expression of CCL5, CCL11, and CCL17 (104), and IL-17A neutralization exaggerated Th2-mediated eosinophilic response (105). Therefore, several *in vivo* and *ex vivo* models of allergic airway inflammation demonstrated the varying and conflicting roles of IL-17A and Th17 cells. Conversely, a murine model reinforced the synergistic signaling pattern of IL-23, and Th17 cells leading to increased eosinophilic Th2 response (106). Therefore, the precise and possible Th17-mediated mechanisms involved in allergic inflammation and RV-induced asthma exacerbation, are unknown.

The features of Th17 cell-dependent asthma triggered a new debate following the possible cross-talk between IL-17A and IL-22. Displaying both pro- and anti-inflammatory features, IL-22 is secreted from Th1, Th22, and Th17 cells with dominant IL-6, and TNF- $\alpha$ -dependent traits (107). A few reports suggested that CD4<sup>+</sup> CCR6<sup>+</sup> Th17 cells manifested the IL-22-secreting cellular phenotypes; however, lack of IL-22 resulted in a decreased Th2 response (107). Other studies reported contrasting asthmatic features associated with the induction of IL-17E, and IL-13 upon IL-22 neutralization in OVA-challenged asthma (108). Taken together, due to the pro- and anti-inflammatory roles of IL-22, several studies postulate a distinct role of CD4<sup>+</sup>Th22 cells in asthma progression.

Currently, the canonical asthma signatures have been used to address new concepts involving the critical differentiation of IL-9-dependent CD4<sup>+</sup>Th9 cells from Th2 cells (109). Previously, Th9 cells were treated as classical Th2 cells, due to their capacity to induce IL-9-dependent mast cell differentiation, goblet cell metaplasia, and IL-13 production (110). A few *ex vivo* models demonstrated Th9 cellular characterization upon IL-4, and TGF- $\beta$  priming (109). Thus, in general, the CD4<sup>+</sup>Th9 cells induce Th2-like asthma features characterized by bronchial hyper-reactivity, mucus production, and airway eosinophilia following allergen exposure. However, the roles of Th2 and Th9 cells in asthma pathophysiology have been demonstrated adequately using eosinophil-deficient mice models (111). Therefore, an investigation into the Th9 cellular migration in asthmatic lungs is needed to elucidate the mechanism distinct from the Th2 response.

Allergic asthma or virus-induced severe airway inflammation resulting in Th1, Th2, Th17, Th22, Th9 or mixed inflammation can be controlled by Tregs to maintain airway tolerance against inhaled antigens (112). Usually, anti-inflammatory IL-10 plays a vital role in Treg

function. Moreover, TGF- $\beta$  also plays a bystander role in immune suppression via complex pathways to induce airway remodeling, and maintenance of immune homeostasis (113). Both thymic-derived and virus-induced CD4<sup>+</sup>FoxP3<sup>+</sup>Treg cells (iTregs/nTregs) disrupt the regulatory mechanism by inhibiting the effector T cell function, and the immunomodulatory role of subsequent APCs in airways, via IL-10 and TGF- $\beta$  (114). Moreover, upon adaptive transfer, the antigen-specific CD4<sup>+</sup>CD25<sup>+</sup> Tregs may also initiate allergic inflammation and AHR suppression via IL-10-dependent mechanism (115), and CD4<sup>+</sup>CD25<sup>+</sup> Tregs induced the expression of the chemokine receptors CCR4 and CCR8 (116). Differentiation of highly secretory IL-10-dependent Tregs via ICOS-ICOS ligand pathway in asthmatic subjects was triggered by higher doses of inhaled allergens following chronic exposure, suggesting that repeated stimulation of airway DCs may strongly elevate IL-10 levels (117,118).

In our preceding studies using the Japanese encephalitis virus mice model, we reported the roles of CCR5 chemokine receptor in the recruitment of CD4<sup>+</sup>FoxP3<sup>+</sup>Tregs during CNS inflammation (119). Similarly, we also found a diminished population of CD4<sup>+</sup>FoxP3<sup>+</sup>Tregs in CCR5-deficient mice following asthma exacerbation induced via allergen-sensitized RV-1B (unpublished personal data). Therefore, the CCR5 chemokine receptor also plays a role in the initiation and maintenance of CD4<sup>+</sup>Tregs. Although CD4<sup>+</sup> Tregs received extensive attention, studies elucidating the role of CD8<sup>+</sup> T cells, NK,  $\gamma\delta$  T cells, B cells, mast cells, and various APC populations in asthma are needed.

## CONCLUSION AND FUTURE PERSPECTIVES

In asthma pathogenesis, a robust and complex cytokine connection has been established using numerous *in vivo* and *in vitro* models, demonstrating the differentiation and initiation mechanisms mediated via Th1, Th2, and Th17 or even Th9 response. The specific mechanism underlying the differentiation of Th1 or Th2 response in atopic asthma or respiratory virus-induced exacerbation (Th17 or Th17/Th2 mixed response) is intriguing, as the balancing roles of pro-inflammatory and suppressive Tregs suggest new paradigms and mechanisms associated with asthma prognosis.

Respiratory viral infections, particularly those associated with RVs, are the foremost cause of morbidity and mortality in exacerbated asthma due to poor prognosis, and resistance to corticosteroids. Patients diagnosed with allergic asthma are highly susceptible to respiratory virus and exhibit severe non-canonical asthma pathogenesis. Impaired antiviral immune responses are associated with severe disease burden, economic loss, disease severity and even mortality. Moreover, the specific anti-viral strategies are still under development in the absence of clear-cut efforts to prevent or combat virus-initiated asthma exacerbations. In addition to traditional ICS, the introduction of IL-4R $\alpha$  is effective in Th2 asthma response by inhibiting the expression of IL-4 and IL-13. Moreover, mepolizumab, reslizumab, benralizumab, and dupilumab are also effective against classical asthma; however, they are associated with a poor response against virus-induced asthma characterized by a non-Th2 immune response. Different disease models enhance our understanding of the molecular epidemiology and mechanisms underlying disease progression. They provide opportunities for the development of antiviral therapies against specific targets for effective management and prevention. Hence, to delineate the virus-induced mechanisms exacerbating asthma for the development of appropriate antiviral therapeutics, extensive and sophisticated efforts are needed using effective *in vivo* and *ex vivo* models.

## ACKNOWLEDGMENTS

This study was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) grant funded by The Ministry of Education Science and The Ministry of Science and ICT (MSIT), Republic of Korea (2017R1E1A2A01077188, 2018R1A2B2001873, 2019R1A6A1A03033084, <http://www.nrf.re.kr>).

## REFERENCES

- Anderson GP. Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogeneous disease. *Lancet* 2008;372:1107-1119.  
[PUBMED](#) | [CROSSREF](#)
- Pawankar R. Allergic diseases and asthma: a global public health concern and a call to action. *World Allergy Organ J* 2014;7:12.  
[PUBMED](#) | [CROSSREF](#)
- Zedan M, Attia G, Zedan MM, Osman A, Abo-Elkheir N, Maysara N, Barakat T, Gamil N. Clinical asthma phenotypes and therapeutic responses. *ISRN Pediatr* 2013;2013:824781.  
[PUBMED](#) | [CROSSREF](#)
- Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. *Nat Med* 2012;18:716-725.  
[PUBMED](#) | [CROSSREF](#)
- Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol* 2015;16:45-56.  
[PUBMED](#) | [CROSSREF](#)
- Agache I. Non-eosinophilic asthma endotypes. *Curr Treat Options Allergy* 2015;2:257-267.  
[CROSSREF](#)
- Jackson DJ, Johnston SL. The role of viruses in acute exacerbations of asthma. *J Allergy Clin Immunol* 2010;125:1178-1187, quiz 1188-quiz 1189.  
[PUBMED](#) | [CROSSREF](#)
- Miller EK, Lu X, Erdman DD, Poehling KA, Zhu Y, Griffin MR, Hartert TV, Anderson LJ, Weinberg GA, Hall CB, et al. Rhinovirus-associated hospitalizations in young children. *J Infect Dis* 2007;195:773-781.  
[PUBMED](#) | [CROSSREF](#)
- Wark PA, Johnston SL, Bucchieri F, Powell R, Puddicombe S, Laza-Stanca V, Holgate ST, Davies DE. Asthmatic bronchial epithelial cells have a deficient innate immune response to infection with rhinovirus. *J Exp Med* 2005;201:937-947.  
[PUBMED](#) | [CROSSREF](#)
- Contoli M, Message SD, Laza-Stanca V, Edwards MR, Wark PA, Bartlett NW, Keadze T, Mallia P, Stanciu LA, Parker HL, et al. Role of deficient type III interferon-lambda production in asthma exacerbations. *Nat Med* 2006;12:1023-1026.  
[PUBMED](#) | [CROSSREF](#)
- Green RM, Custovic A, Sanderson G, Hunter J, Johnston SL, Woodcock A. Synergism between allergens and viruses and risk of hospital admission with asthma: case-control study. *BMJ* 2002;324:763.  
[PUBMED](#) | [CROSSREF](#)
- Khetsuriani N, Kazerouni NN, Erdman DD, Lu X, Redd SC, Anderson LJ, Teague WG. Prevalence of viral respiratory tract infections in children with asthma. *J Allergy Clin Immunol* 2007;119:314-321.  
[PUBMED](#) | [CROSSREF](#)
- Hu X, Li X, Hu C, Qin L, He R, Luo L, Tang W, Feng J. Respiratory syncytial virus exacerbates OVA-mediated asthma in mice through C5a-C5aR regulating CD4<sup>+</sup>T cells immune responses. *Sci Rep* 2017;7:15207.  
[PUBMED](#) | [CROSSREF](#)
- Message SD, Laza-Stanca V, Mallia P, Parker HL, Zhu J, Keadze T, Contoli M, Sanderson G, Kon OM, Papi A, et al. Rhinovirus-induced lower respiratory illness is increased in asthma and related to virus load and Th1/2 cytokine and IL-10 production. *Proc Natl Acad Sci U S A* 2008;105:13562-13567.  
[PUBMED](#) | [CROSSREF](#)
- Busse WW, Lemanske RF Jr, Gern JE. Role of viral respiratory infections in asthma and asthma exacerbations. *Lancet* 2010;376:826-834.  
[PUBMED](#) | [CROSSREF](#)

16. Yilmaz İ. Is Asthma-COPD overlap an asthma phenotype or a COPD phenotype? *Tuberk Toraks* 2018;66:78-79.  
[PUBMED](#) | [CROSSREF](#)
17. Bartlett NW, Walton RP, Edwards MR, Aniscenko J, Caramori G, Zhu J, Glanville N, Choy KJ, Jourdan P, Burnet J, et al. Mouse models of rhinovirus-induced disease and exacerbation of allergic airway inflammation. *Nat Med* 2008;14:199-204.  
[PUBMED](#) | [CROSSREF](#)
18. Whittsett JA, Alenghat T. Respiratory epithelial cells orchestrate pulmonary innate immunity. *Nat Immunol* 2015;16:27-35.  
[PUBMED](#) | [CROSSREF](#)
19. Barnes PJ. Immunology of asthma and chronic obstructive pulmonary disease. *Nat Rev Immunol* 2008;8:183-192.  
[PUBMED](#) | [CROSSREF](#)
20. Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, Arron JR, Koth LL, Fahy JV. T-helper type 2-driven inflammation defines major subphenotypes of asthma. *Am J Respir Crit Care Med* 2009;180:388-395.  
[PUBMED](#) | [CROSSREF](#)
21. Ramsahai JM, Hansbro PM, Wark PA. Mechanisms and management of asthma exacerbations. *Am J Respir Crit Care Med* 2019;199:423-432.  
[PUBMED](#) | [CROSSREF](#)
22. Shi YH, Shi GC, Wan HY, Jiang LH, Ai XY, Zhu HX, Tang W, Ma JY, Jin XY, Zhang BY. Coexistence of Th1/Th2 and Th17/Treg imbalances in patients with allergic asthma. *Chin Med J (Engl)* 2011;124:1951-1956.  
[PUBMED](#)
23. Simpson JL, Grissell TV, Douwes J, Scott RJ, Boyle MJ, Gibson PG. Innate immune activation in neutrophilic asthma and bronchiectasis. *Thorax* 2007;62:211-218.  
[PUBMED](#) | [CROSSREF](#)
24. Kim RY, Horvat JC, Pinkerton JW, Starkey MR, Essilfie AT, Mayall JR, Nair PM, Hansbro NG, Jones B, Haw TJ, et al. MicroRNA-21 drives severe, steroid-insensitive experimental asthma by amplifying phosphoinositide 3-kinase-mediated suppression of histone deacetylase 2. *J Allergy Clin Immunol* 2017;139:519-532.  
[PUBMED](#) | [CROSSREF](#)
25. Kim RY, Pinkerton JW, Essilfie AT, Robertson AA, Baines KJ, Brown AC, Mayall JR, Ali MK, Starkey MR, Hansbro NG, et al. Role for nlrp3 inflammasome-mediated, il-1beta-dependent responses in severe, steroid-resistant asthma. *Am J Respir Crit Care Med* 2017;196:283-297.  
[PUBMED](#) | [CROSSREF](#)
26. Zhang N, Truong-Tran QA, Tancowny B, Harris KE, Schleimer RP. Glucocorticoids enhance or spare innate immunity: effects in airway epithelium are mediated by CCAAT/enhancer binding proteins. *J Immunol* 2007;179:578-589.  
[PUBMED](#) | [CROSSREF](#)
27. Busse W, Chupp G, Nagase H, Albers FC, Doyle S, Shen Q, Bratton DJ, Gunsoy NB. Anti-IL-5 treatments in patients with severe asthma by blood eosinophil thresholds: Indirect treatment comparison. *J Allergy Clin Immunol* 2019;143:190-200.e20.  
[PUBMED](#) | [CROSSREF](#)
28. Macedo P, Hew M, Torrego A, Jouneau S, Oates T, Durham A, Chung KF. Inflammatory biomarkers in airways of patients with severe asthma compared with non-severe asthma. *Clin Exp Allergy* 2009;39:1668-1676.  
[PUBMED](#) | [CROSSREF](#)
29. Papi A, Contoli M, Adcock IM, Bellettato C, Padovani A, Casolari P, Stanciu LA, Barnes PJ, Johnston SL, Ito K, et al. Rhinovirus infection causes steroid resistance in airway epithelium through nuclear factor κB and c-Jun N-terminal kinase activation. *J Allergy Clin Immunol* 2013;132:1075-1085.e6.  
[PUBMED](#) | [CROSSREF](#)
30. Stone JK, Rijnbrand R, Stein DA, Ma Y, Yang Y, Iversen PL, Andino R. A morpholino oligomer targeting highly conserved internal ribosome entry site sequence is able to inhibit multiple species of picornavirus. *Antimicrob Agents Chemother* 2008;52:1970-1981.  
[PUBMED](#) | [CROSSREF](#)
31. Cakebread JA, Xu Y, Grainge C, Kehagia V, Howarth PH, Holgate ST, Davies DE. Exogenous IFN-β has antiviral and anti-inflammatory properties in primary bronchial epithelial cells from asthmatic subjects exposed to rhinovirus. *J Allergy Clin Immunol* 2011;127:1148-1154.e9.  
[PUBMED](#) | [CROSSREF](#)
32. Barnes PJ. Targeting cytokines to treat asthma and chronic obstructive pulmonary disease. *Nat Rev Immunol* 2018;18:454-466.  
[PUBMED](#) | [CROSSREF](#)

33. Kaminuma O, Ohtomo T, Mori A, Nagakubo D, Hieshima K, Ohmachi Y, Noda Y, Katayama K, Suzuki K, Motoi Y, et al. Selective down-regulation of Th2 cell-mediated airway inflammation in mice by pharmacological intervention of CCR4. *Clin Exp Allergy* 2012;42:315-325.  
[PUBMED](#) | [CROSSREF](#)
34. Kara EE, Comerford I, Bastow CR, Fenix KA, Litchfield W, Handel TM, McColl SR. Distinct chemokine receptor axes regulate Th9 cell trafficking to allergic and autoimmune inflammatory sites. *J Immunol* 2013;191:1110-1117.  
[PUBMED](#) | [CROSSREF](#)
35. Samarasinghe AE, Woolard SN, Boyd KL, Hoselton SA, Schuh JM, McCullers JA. The immune profile associated with acute allergic asthma accelerates clearance of influenza virus. *Immunol Cell Biol* 2014;92:449-459.  
[PUBMED](#) | [CROSSREF](#)
36. Rydell-Törmänen K, Johnson JR. The applicability of mouse models to the study of human disease. *Methods Mol Biol* 2019;1940:3-22.  
[PUBMED](#) | [CROSSREF](#)
37. Nagarkar DR, Bowman ER, Schneider D, Wang Q, Shim J, Zhao Y, Linn MJ, McHenry CL, Gosangi B, Bentley JK, et al. Rhinovirus infection of allergen-sensitized and -challenged mice induces eotaxin release from functionally polarized macrophages. *J Immunol* 2010;185:2525-2535.  
[PUBMED](#) | [CROSSREF](#)
38. Tan HT, Hagner S, Ruchti F, Radzikowska U, Tan G, Altunbulakli C, Eljaszewicz A, Moniuszko M, Akdis M, Akdis CA, et al. Tight junction, mucin, and inflammasome-related molecules are differentially expressed in eosinophilic, mixed, and neutrophilic experimental asthma in mice. *Allergy* 2019;74:294-307.  
[PUBMED](#) | [CROSSREF](#)
39. Kumar RK, Herbert C, Foster PS. The “classical” ovalbumin challenge model of asthma in mice. *Curr Drug Targets* 2008;9:485-494.  
[PUBMED](#) | [CROSSREF](#)
40. Sajjan U, Wang Q, Zhao Y, Gruenert DC, Hershenson MB. Rhinovirus disrupts the barrier function of polarized airway epithelial cells. *Am J Respir Crit Care Med* 2008;178:1271-1281.  
[PUBMED](#) | [CROSSREF](#)
41. Slater L, Bartlett NW, Haas JJ, Zhu J, Message SD, Walton RP, Sykes A, Dahdaleh S, Clarke DL, Belvisi MG, et al. Co-ordinated role of TLR3, RIG-I and MDA5 in the innate response to rhinovirus in bronchial epithelium. *PLoS Pathog* 2010;6:e1001178.  
[PUBMED](#) | [CROSSREF](#)
42. Zhu J, Message SD, Mallia P, Keadze T, Contoli M, Ward CK, Barnathan ES, Mascelli MA, Kon OM, Papi A, et al. Bronchial mucosal IFN- $\alpha/\beta$  and pattern recognition receptor expression in patients with experimental rhinovirus-induced asthma exacerbations. *J Allergy Clin Immunol* 2019;143:114-125.e4.  
[PUBMED](#) | [CROSSREF](#)
43. Goulopoulou S, McCarthy CG, Webb RC. Toll-like receptors in the vascular system: Sensing the dangers within. *Pharmacol Rev* 2016;68:142-167.  
[PUBMED](#) | [CROSSREF](#)
44. Yamamoto M, Sato S, Hemmi H, Hoshino K, Kaisho T, Sanjo H, Takeuchi O, Sugiyama M, Okabe M, Takeda K, et al. Role of adaptor TRIF in the MyD88-independent toll-like receptor signaling pathway. *Science* 2003;301:640-643.  
[PUBMED](#) | [CROSSREF](#)
45. Tizaoui K, Kaabachi W, Hamzaoui K, Hamzaoui A. Association of single nucleotide polymorphisms in toll-like receptor genes with asthma risk: a systematic review and meta-analysis. *Allergy Asthma Immunol Res* 2015;7:130-140.  
[PUBMED](#) | [CROSSREF](#)
46. Zuo L, Lucas K, Fortuna CA, Chuang CC, Best TM. Molecular regulation of toll-like receptors in asthma and COPD. *Front Physiol* 2015;6:312.  
[PUBMED](#) | [CROSSREF](#)
47. Frotscher B, Anton K, Worm M. Inhibition of IgE production by the imidazoquinoline resiquimod in nonallergic and allergic donors. *J Invest Dermatol* 2002;119:1059-1064.  
[PUBMED](#) | [CROSSREF](#)
48. Yoneyama M, Kikuchi M, Matsumoto K, Imaizumi T, Miyagishi M, Taira K, Foy E, Loo YM, Gale M Jr, Akira S, et al. Shared and unique functions of the DExD/H-box helicases RIG-I, MDA5, and LGP2 in antiviral innate immunity. *J Immunol* 2005;175:2851-2858.  
[PUBMED](#) | [CROSSREF](#)

49. Tan KS, Ong HH, Yan Y, Liu J, Li C, Ong YK, Thong KT, Choi HW, Wang DY, Chow VT. *In vitro* model of fully differentiated human nasal epithelial cells infected with rhinovirus reveals epithelium-initiated immune responses. *J Infect Dis* 2018;217:906-915.  
[PUBMED](#) | [CROSSREF](#)
50. Triantafylou K, Vakakis E, Richer EA, Evans GL, Villiers JP, Triantafylou M. Human rhinovirus recognition in non-immune cells is mediated by Toll-like receptors and MDA-5, which trigger a synergetic pro-inflammatory immune response. *Virulence* 2011;2:22-29.  
[PUBMED](#) | [CROSSREF](#)
51. Boldogh I, Bacsi A, Choudhury BK, Dharajiyi N, Alam R, Hazra TK, Mitra S, Goldblum RM, Sur S. ROS generated by pollen NADPH oxidase provide a signal that augments antigen-induced allergic airway inflammation. *J Clin Invest* 2005;115:2169-2179.  
[PUBMED](#) | [CROSSREF](#)
52. Zhu J, Message SD, Qiu Y, Mallia P, Keadze T, Contoli M, Ward CK, Barnathan ES, Mascelli MA, Kon OM, et al. Airway inflammation and illness severity in response to experimental rhinovirus infection in asthma. *Chest* 2014;145:1219-1229.  
[PUBMED](#) | [CROSSREF](#)
53. Contoli M, Ito K, Padovani A, Poletti D, Marku B, Edwards MR, Stanciu LA, Gnesini G, Pastore A, Spanevello A, et al. Th2 cytokines impair innate immune responses to rhinovirus in respiratory epithelial cells. *Allergy* 2015;70:910-920.  
[PUBMED](#) | [CROSSREF](#)
54. Xiao C, Puddicombe SM, Field S, Haywood J, Broughton-Head V, Puxeddu I, Haitchi HM, Vernon-Wilson E, Sammut D, Bedke N, et al. Defective epithelial barrier function in asthma. *J Allergy Clin Immunol* 2011;128:549-556.e1-12.  
[PUBMED](#) | [CROSSREF](#)
55. Chang YJ, Kim HY, Albacker LA, Baumgarth N, McKenzie AN, Smith DE, Dekruyff RH, Umetsu DT. Innate lymphoid cells mediate influenza-induced airway hyper-reactivity independently of adaptive immunity. *Nat Immunol* 2011;12:631-638.  
[PUBMED](#) | [CROSSREF](#)
56. Ckless K, Hodgkins SR, Ather JL, Martin R, Poynter ME. Epithelial, dendritic, and CD4<sup>+</sup> T cell regulation of and by reactive oxygen and nitrogen species in allergic sensitization. *Biochim Biophys Acta* 2011;1810:1025-1034.  
[PUBMED](#) | [CROSSREF](#)
57. Schmidt LM, Belvisi MG, Bode KA, Bauer J, Schmidt C, Suchy MT, Tsikas D, Scheuerer J, Lasitschka F, Gröne HJ, et al. Bronchial epithelial cell-derived prostaglandin E2 dampens the reactivity of dendritic cells. *J Immunol* 2011;186:2095-2105.  
[PUBMED](#) | [CROSSREF](#)
58. Fahy JV. Type 2 inflammation in asthma--present in most, absent in many. *Nat Rev Immunol* 2015;15:57-65.  
[PUBMED](#) | [CROSSREF](#)
59. Bankova LG, Dwyer DF, Yoshimoto E, Ualiyeva S, McGinty JW, Raff H, von Moltke J, Kanaoka Y, Frank Austen K, Barrett NA. The cysteinyl leukotriene 3 receptor regulates expansion of IL-25-producing airway brush cells leading to type 2 inflammation. *Sci Immunol* 2018;3:eaat9453.  
[PUBMED](#) | [CROSSREF](#)
60. Jackson DJ, Makrinioti H, Rana BM, Shamji BW, Trujillo-Torralba MB, Footitt J, Jerico Del-Rosario, Telcian AG, Nikonova A, Zhu J, et al. IL-33-dependent type 2 inflammation during rhinovirus-induced asthma exacerbations *in vivo*. *Am J Respir Crit Care Med* 2014;190:1373-1382.  
[PUBMED](#) | [CROSSREF](#)
61. Akuthota P, Wang H, Weller PF. Eosinophils as antigen-presenting cells in allergic upper airway disease. *Curr Opin Allergy Clin Immunol* 2010;10:14-19.  
[PUBMED](#) | [CROSSREF](#)
62. Snelgrove RJ, Patel DF, Patel T, Lloyd CM. The enigmatic role of the neutrophil in asthma: Friend, foe or indifferent? *Clin Exp Allergy* 2018;48:1275-1285.  
[PUBMED](#) | [CROSSREF](#)
63. Toussaint M, Jackson DJ, Swieboda D, Guedán A, Tsourouktsoglou TD, Ching YM, Radermecker C, Makrinioti H, Aniscenko J, Bartlett NW, et al. Host DNA released by NETosis promotes rhinovirus-induced type-2 allergic asthma exacerbation. *Nat Med* 2017;23:681-691.  
[PUBMED](#) | [CROSSREF](#)
64. Busse WW. A role for neutrophils in asthma exacerbations. *Nat Med* 2017;23:658-659.  
[PUBMED](#) | [CROSSREF](#)

65. Lee YG, Jeong JJ, Nyenhuis S, Berdyshev E, Chung S, Ranjan R, Karpurapu M, Deng J, Qian F, Kelly EA, et al. Recruited alveolar macrophages, in response to airway epithelial-derived monocyte chemoattractant protein 1/CCL2, regulate airway inflammation and remodeling in allergic asthma. *Am J Respir Cell Mol Biol* 2015;52:772-784.  
[PUBMED](#) | [CROSSREF](#)
66. Nomura H, Sato E, Koyama S, Haniuda M, Kubo K, Nagai S, Izumi T. Histamine stimulates alveolar macrophages to release neutrophil and monocyte chemotactic activity. *J Lab Clin Med* 2001;138:226-235.  
[PUBMED](#) | [CROSSREF](#)
67. Subauste MC, Jacoby DB, Richards SM, Proud D. Infection of a human respiratory epithelial cell line with rhinovirus. Induction of cytokine release and modulation of susceptibility to infection by cytokine exposure. *J Clin Invest* 1995;96:549-557.  
[PUBMED](#) | [CROSSREF](#)
68. Hong JY, Chung Y, Steenrod J, Chen Q, Lei J, Comstock AT, Goldsmith AM, Bentley JK, Sajjan US, Hershenson MB. Macrophage activation state determines the response to rhinovirus infection in a mouse model of allergic asthma. *Respir Res* 2014;15:63.  
[PUBMED](#) | [CROSSREF](#)
69. Chung Y, Hong JY, Lei J, Chen Q, Bentley JK, Hershenson MB. Rhinovirus infection induces interleukin-13 production from CD11b-positive, M2-polarized exudative macrophages. *Am J Respir Cell Mol Biol* 2015;52:205-216.  
[PUBMED](#) | [CROSSREF](#)
70. Nakasone C, Yamamoto N, Nakamatsu M, Kinjo T, Miyagi K, Uezu K, Nakamura K, Higa F, Ishikawa H, O'Brien RL, et al. Accumulation of gamma/delta T cells in the lungs and their roles in neutrophil-mediated host defense against pneumococcal infection. *Microbes Infect* 2007;9:251-258.  
[PUBMED](#) | [CROSSREF](#)
71. Glanville N, Message SD, Walton RP, Pearson RM, Parker HL, Laza-Stanca V, Mallia P, Kebabdz T, Contoli M, Kon OM, et al.  $\gamma\delta$ T cells suppress inflammation and disease during rhinovirus-induced asthma exacerbations. *Mucosal Immunol* 2013;6:1091-1100.  
[PUBMED](#) | [CROSSREF](#)
72. Yang X, Zhang JH, Deng WS, Li CQ. Imbalance of  $\gamma\delta$ T17/ $\gamma\delta$ Treg cells in the pathogenesis of allergic asthma induced by ovalbumin. *Braz J Med Biol Res* 2018;51:e7127.  
[PUBMED](#) | [CROSSREF](#)
73. Kim SW, Kim JH, Park CK, Kim TJ, Lee SY, Kim YK, Kwon SS, Rhee CK, Yoon HK. Effect of roflumilast on airway remodelling in a murine model of chronic asthma. *Clin Exp Allergy* 2016;46:754-763.  
[PUBMED](#) | [CROSSREF](#)
74. Hammad H, Lambrecht BN. Barrier epithelial cells and the control of type 2 immunity. *Immunity* 2015;43:29-40.  
[PUBMED](#) | [CROSSREF](#)
75. Kim J, Chang Y, Bae B, Sohn KH, Cho SH, Chung DH, Kang HR, Kim HY. Innate immune crosstalk in asthmatic airways: Innate lymphoid cells coordinate polarization of lung macrophages. *J Allergy Clin Immunol* 2019;143:1769-1782.e11.  
[PUBMED](#) | [CROSSREF](#)
76. Verma M, Liu S, Michalec L, Sripada A, Gorska MM, Alam R. Experimental asthma persists in IL-33 receptor knockout mice because of the emergence of thymic stromal lymphopoietin-driven IL-9<sup>+</sup> and IL-13<sup>+</sup> type 2 innate lymphoid cell subpopulations. *J Allergy Clin Immunol* 2018;142:793-803.e8.  
[PUBMED](#) | [CROSSREF](#)
77. Saenz SA, Siracusa MC, Perrigoue JG, Spencer SP, Urban JF Jr, Tocker JE, Budelsky AL, Kleinschek MA, Kastelein RA, Kambayashi T, et al. IL25 elicits a multipotent progenitor cell population that promotes T(H)2 cytokine responses. *Nature* 2010;464:1362-1366.  
[PUBMED](#) | [CROSSREF](#)
78. Bando JK, Gilfillan S, Song C, McDonald KG, Huang SC, Newberry RD, Kobayashi Y, Allan DS, Carlyle JR, Cella M, et al. The tumor necrosis factor superfamily member RANKL suppresses effector cytokine production in group 3 innate lymphoid cells. *Immunity* 2018;48:1208-1219.e4.  
[PUBMED](#) | [CROSSREF](#)
79. Ivashkiv LB, Donlin LT. Regulation of type I interferon responses. *Nat Rev Immunol* 2014;14:36-49.  
[PUBMED](#) | [CROSSREF](#)
80. Bhakta NR, Christenson SA, Nerella S, Solberg OD, Nguyen CP, Choy DF, Jung KL, Garudadri S, Bonser LR, Pollack JL, et al. IFN-stimulated gene expression, type 2 inflammation, and endoplasmic reticulum stress in asthma. *Am J Respir Crit Care Med* 2018;197:313-324.  
[PUBMED](#) | [CROSSREF](#)

81. Edwards MR, Regamey N, Vareille M, Kieninger E, Gupta A, Shoemark A, Saglani S, Sykes A, Macintyre J, Davies J, et al. Impaired innate interferon induction in severe therapy resistant atopic asthmatic children. *Mucosal Immunol* 2013;6:797-806.  
[PUBMED](#) | [CROSSREF](#)
82. Kikkawa Y, Sugiyama K, Obara K, Hirata H, Fukushima Y, Toda M, Fukuda T. Interferon-alpha inhibits airway eosinophilia and hyperresponsiveness in an animal asthma model [corrected]. *Asia Pac Allergy* 2012;2:256-263.  
[PUBMED](#) | [CROSSREF](#)
83. Baraldo S, Contoli M, Bazzan E, Turato G, Padovani A, Marku B, Calabrese F, Caramori G, Ballarin A, Snijders D, et al. Deficient antiviral immune responses in childhood: distinct roles of atopy and asthma. *J Allergy Clin Immunol* 2012;130:1307-1314.  
[PUBMED](#) | [CROSSREF](#)
84. Fulkerson PC, Zimmermann N, Brandt EB, Muntel EE, Doepker MP, Kavanaugh JL, Mishra A, Witte DP, Zhang H, Farber JM, et al. Negative regulation of eosinophil recruitment to the lung by the chemokine monokine induced by IFN-gamma (Mig, CXCL9). *Proc Natl Acad Sci U S A* 2004;101:1987-1992.  
[PUBMED](#) | [CROSSREF](#)
85. Coyle AJ, Tsuyuki S, Bertrand C, Huang S, Aguet M, Alkan SS, Anderson GP. Mice lacking the IFN-gamma receptor have impaired ability to resolve a lung eosinophilic inflammatory response associated with a prolonged capacity of T cells to exhibit a Th2 cytokine profile. *J Immunol* 1996;156:2680-2685.  
[PUBMED](#)
86. Boehm U, Klamp T, Groot M, Howard JC. Cellular responses to interferon-gamma. *Annu Rev Immunol* 1997;15:749-795.  
[PUBMED](#) | [CROSSREF](#)
87. Sehra S, Pynaert G, Tournoy K, Haegeman A, Matthys P, Tagawa Y, Pauwels R, Grooten J. Airway IgG counteracts specific and bystander allergen-triggered pulmonary inflammation by a mechanism dependent on Fc gamma R and IFN-gamma. *J Immunol* 2003;171:2080-2089.  
[PUBMED](#) | [CROSSREF](#)
88. Spann KM, Tran KC, Chi B, Rabin RL, Collins PL. Suppression of the induction of alpha, beta, and lambda interferons by the NS1 and NS2 proteins of human respiratory syncytial virus in human epithelial cells and macrophages [corrected]. *J Virol* 2004;78:4363-4369.  
[PUBMED](#) | [CROSSREF](#)
89. Yang K, Puel A, Zhang S, Eidenschenk C, Ku CL, Casrouge A, Picard C, von Bernuth H, Senechal B, Plancoulaine S, et al. Human TLR-7-, -8-, and -9-mediated induction of IFN-alpha/beta and -lambda is IRAK-4 dependent and redundant for protective immunity to viruses. *Immunity* 2005;23:465-478.  
[PUBMED](#) | [CROSSREF](#)
90. Sykes A, Edwards MR, Macintyre J, del Rosario A, Bakhsoliani E, Trujillo-Torralbo MB, Kon OM, Mallia P, McHale M, Johnston SL. Rhinovirus 16-induced IFN- $\alpha$  and IFN- $\beta$  are deficient in bronchoalveolar lavage cells in asthmatic patients. *J Allergy Clin Immunol* 2012;129:1506-1514.e6.  
[PUBMED](#) | [CROSSREF](#)
91. Lasfar A, Zloza A, Cohen-Solal KA. IFN-lambda therapy: current status and future perspectives. *Drug Discov Today* 2016;21:167-171.  
[PUBMED](#) | [CROSSREF](#)
92. Cannons JL, Lu KT, Schwartzberg PL. Lymph node choreography: B cells take the lead. *Nat Immunol* 2012;13:630-632.  
[PUBMED](#) | [CROSSREF](#)
93. Altin J, Shen C, Liston A. Understanding the genetic regulation of IgE production. *Blood Rev* 2010;24:163-169.  
[PUBMED](#) | [CROSSREF](#)
94. Sallmann E, Reininger B, Brandt S, Duschek N, Hoflehner E, Garner-Spitzer E, Platzer B, Dehlink E, Hammer M, Holcman M, et al. High-affinity IgE receptors on dendritic cells exacerbate Th2-dependent inflammation. *J Immunol* 2011;187:164-171.  
[PUBMED](#) | [CROSSREF](#)
95. Lindell DM, Berlin AA, Schaller MA, Lukacs NW. B cell antigen presentation promotes Th2 responses and immunopathology during chronic allergic lung disease. *PLoS One* 2008;3:e3129.  
[PUBMED](#) | [CROSSREF](#)
96. Harris DP, Haynes L, Sayles PC, Duso DK, Eaton SM, Lepak NM, Johnson LL, Swain SL, Lund FE. Reciprocal regulation of polarized cytokine production by effector B and T cells. *Nat Immunol* 2000;1:475-482.  
[PUBMED](#) | [CROSSREF](#)
97. Lund FE, Randall TD. Effector and regulatory B cells: modulators of CD4<sup>+</sup> T cell immunity. *Nat Rev Immunol* 2010;10:236-247.  
[PUBMED](#) | [CROSSREF](#)

98. Nakamura Y, Ghaffar O, Olivenstein R, Taha RA, Soussi-Gounni A, Zhang DH, Ray A, Hamid Q. Gene expression of the GATA-3 transcription factor is increased in atopic asthma. *J Allergy Clin Immunol* 1999;103:215-222.  
[PUBMED](#) | [CROSSREF](#)
99. Lambrecht BN, Hammad H, Fahy JV. The cytokines of asthma. *Immunity* 2019;50:975-991.  
[PUBMED](#) | [CROSSREF](#)
100. Ngoc PL, Gold DR, Tzianabos AO, Weiss ST, Celedón JC. Cytokines, allergy, and asthma. *Curr Opin Allergy Clin Immunol* 2005;5:161-166.  
[PUBMED](#) | [CROSSREF](#)
101. Alcorn JF, Crowe CR, Kolls JK. Th17 cells in asthma and COPD. *Annu Rev Physiol* 2010;72:495-516.  
[PUBMED](#) | [CROSSREF](#)
102. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. *Annu Rev Immunol* 2009;27:485-517.  
[PUBMED](#) | [CROSSREF](#)
103. Choy DF, Hart KM, Borthwick LA, Shikotra A, Nagarkar DR, Siddiqui S, Jia G, Ohri CM, Doran E, Vannella KM, et al. Th2 and Th17 inflammatory pathways are reciprocally regulated in asthma. *Sci Transl Med* 2015;7:301ra129.  
[PUBMED](#) | [CROSSREF](#)
104. Schnyder-Candrian S, Togbe D, Couillin I, Mercier I, Brombacher F, Quesniaux V, Fossiez F, Ryffel B, Schnyder B. Interleukin-17 is a negative regulator of established allergic asthma. *J Exp Med* 2006;203:2715-2725.  
[PUBMED](#) | [CROSSREF](#)
105. Barlow JL, Flynn RJ, Ballantyne SJ, McKenzie AN. Reciprocal expression of IL-25 and IL-17A is important for allergic airways hyperreactivity. *Clin Exp Allergy* 2011;41:1447-1455.  
[PUBMED](#) | [CROSSREF](#)
106. Wakashin H, Hirose K, Maezawa Y, Kagami S, Suto A, Watanabe N, Saito Y, Hatano M, Tokuhisa T, Iwakura Y, et al. IL-23 and Th17 cells enhance Th2-cell-mediated eosinophilic airway inflammation in mice. *Am J Respir Crit Care Med* 2008;178:1023-1032.  
[PUBMED](#) | [CROSSREF](#)
107. Besnard AG, Sabat R, Dumoutier L, Renaud JC, Willart M, Lambrecht B, Teixeira MM, Charron S, Fick L, Erard F, et al. Dual role of IL-22 in allergic airway inflammation and its cross-talk with IL-17A. *Am J Respir Crit Care Med* 2011;183:1153-1163.  
[PUBMED](#) | [CROSSREF](#)
108. Takahashi K, Hirose K, Kawashima S, Niwa Y, Wakashin H, Iwata A, Tokoyoda K, Renaud JC, Iwamoto I, Nakayama T, et al. IL-22 attenuates IL-25 production by lung epithelial cells and inhibits antigen-induced eosinophilic airway inflammation. *J Allergy Clin Immunol* 2011;128:1067-1076.e1-6.  
[PUBMED](#) | [CROSSREF](#)
109. Veldhoen M, Uyttenhove C, van Snick J, Helmby H, Westendorf A, Buer J, Martin B, Wilhelm C, Stockinger B. Transforming growth factor-beta 'reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing subset. *Nat Immunol* 2008;9:1341-1346.  
[PUBMED](#) | [CROSSREF](#)
110. Matsuzawa S, Sakashita K, Kinoshita T, Ito S, Yamashita T, Koike K. IL-9 enhances the growth of human mast cell progenitors under stimulation with stem cell factor. *J Immunol* 2003;170:3461-3467.  
[PUBMED](#) | [CROSSREF](#)
111. Saeki M, Kaminuma O, Nishimura T, Kitamura N, Mori A, Hiroi T. Th9 cells elicit eosinophil-independent bronchial hyperresponsiveness in mice. *Allergol Int* 2016;65 Suppl:S24-S29.  
[PUBMED](#) | [CROSSREF](#)
112. Choi IS. Immune tolerance by induced regulatory T cells in asthma. *Allergy Asthma Immunol Res* 2012;4:113-115.  
[PUBMED](#) | [CROSSREF](#)
113. Vignali DA, Collison LW, Workman CJ. How regulatory T cells work. *Nat Rev Immunol* 2008;8:523-532.  
[PUBMED](#) | [CROSSREF](#)
114. Andersson J, Tran DQ, Pesu M, Davidson TS, Ramsey H, O'Shea JJ, Shevach EM. CD4<sup>+</sup> FoxP3<sup>+</sup> regulatory T cells confer infectious tolerance in a TGF-beta-dependent manner. *J Exp Med* 2008;205:1975-1981.  
[PUBMED](#) | [CROSSREF](#)
115. Kearley J, Robinson DS, Lloyd CM. CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells reverse established allergic airway inflammation and prevent airway remodeling. *J Allergy Clin Immunol* 2008;122:617-624.e6.  
[PUBMED](#) | [CROSSREF](#)
116. Sather BD, Treuting P, Perdue N, Miazgowiec M, Fontenot JD, Rudensky AY, Campbell DJ. Altering the distribution of Foxp3<sup>+</sup> regulatory T cells results in tissue-specific inflammatory disease. *J Exp Med* 2007;204:1335-1347.  
[PUBMED](#) | [CROSSREF](#)

117. Akbari O, Freeman GJ, Meyer EH, Greenfield EA, Chang TT, Sharpe AH, Berry G, DeKruyff RH, Umetsu DT. Antigen-specific regulatory T cells develop via the ICOS-ICOS-ligand pathway and inhibit allergen-induced airway hyperreactivity. *Nat Med* 2002;8:1024-1032.  
[PUBMED](#) | [CROSSREF](#)
118. Ali FR, Oldfield WL, Higashi N, Larché M, Kay AB. Late asthmatic reactions induced by inhalation of allergen-derived T cell peptides. *Am J Respir Crit Care Med* 2004;169:20-26.  
[PUBMED](#) | [CROSSREF](#)
119. Kim JH, Patil AM, Choi JY, Kim SB, Uyangaa E, Hossain FM, Park SY, Lee JH, Eo SK. CCR5 ameliorates Japanese encephalitis via dictating the equilibrium of regulatory CD4<sup>+</sup>Foxp3<sup>+</sup> T and IL-17<sup>+</sup>CD4<sup>+</sup> Th17 cells. *J Neuroinflammation* 2016;13:223.  
[PUBMED](#) | [CROSSREF](#)