

Clinical Prognostic Values of Vascular Endothelial Growth Factor, Microvessel Density, and p53 Expression in Esophageal Carcinomas

Vascular endothelial growth factor (VEGF) is known to play a key role in tumor angiogenesis. The tumor-suppressor gene p53 has been thought to regulate VEGF. We investigated the effect of VEGF on esophageal carcinoma and the correlation between VEGF and p53. Tissue samples were taken from 81 patients with esophageal carcinoma after surgery. VEGF and p53 expressions were examined by immunohistochemical staining. Microvessels in the tumor stained for CD34 antigen were also counted. VEGF and p53 expressions were observed in 51.3% (41/80) and 51.9% (41/79), respectively. The microvessel density was 70.9 ± 6.7 (mean \pm SE) in VEGF-positive group and 68.7 ± 5.1 in VEGF-negative group. However, no correlation was noted between VEGF and p53 expression. Whereas the tumor size, nodal status, depth of invasions, and tumor stage were associated with poor overall survival, VEGF expression or p53 expression was not. These results indicate that VEGF and p53 are highly expressed in esophageal carcinomas. Since the VEGF expression is not correlated with the p53 expression, microvessel density or clinicopathological findings, further studies with other angiogenic molecules are needed to determine the role in esophageal carcinomas.

Key Words : Endothelial Growth Factor; Microvessel Density; Genes, p53; Esophageal Neoplasms

Myung-Ju Ahn, Se-Jin Jang*,
Yong-Wook Park*, Jung-Hye Choi,
Ho-Suk Oh, Chul-Burm Lee[†],
Hong-Kyu Paik[‡], Chan-Kum Park*

Department of Internal Medicine, Department of Pathology*, Department of Thoracic Surgery[†], Department of General Surgery[‡], College of Medicine, Hanyang University, Seoul, Korea

Received : 23 July 2001
Accepted : 24 December 2001

Address for correspondence

Chan-Kum Park, M.D.
Department of Pathology, Hanyang University Hospital, 17 Handang-dong, Sungdong-gu, Seoul 133-791, Korea
Tel : +82.2-2290-8250, Fax : +82.2-2296-7502
E-mail : parkcg@hanyang.ac.kr

INTRODUCTION

Esophageal carcinoma is one of the most common malignancies in the world. Since the growth of tumor is relatively fast, patients with esophageal carcinoma generally have a worse prognosis than those with any other gastrointestinal tumors. It has been thought that several factors such as stage, histological grade, DNA ploidy, epidermal growth factor receptor, p53, and lymph node metastasis influence the survival. Angiogenesis, which is essential for tumor growth and metastasis, depends on the production of angiogenic factors by tumor cells and normal cells. Increased vascularity enhances the growth of the primary neoplasm and provides a greater chance for a hematogenous metastasis. It has been shown to have a prognostic value in several solid tumors such as breast, lung, prostate, cervical, and colon cancer (1-5). Previous studies also have demonstrated that the vascular density of a tumor directly correlates with metastasis and poor outcome in patients with solid tumors (9-14).

Vascular endothelial growth factor (VEGF) is an angiogenic factor that stimulates the growth of endothelial cells (15-17). It consists of four isoforms that have 121, 165, 189, and 206 amino acid residues and all four types of VEGF are secreted in abundance by many kinds of carcinoma cells (18). Recent

studies showed a positive correlation among the VEGF expression, tumor microvessel density (MVD), and tumor aggressiveness (19-21).

Alterations of the p53 tumor suppressor gene have known to be the most common genetic changes in solid tumors including esophageal carcinomas. Recent reports showed that mutations of p53 might be associated with angiogenesis by regulating the VEGF expression (22, 23).

To clarify the prognostic significance and relationship between VEGF expression and clinicopathological features and its correlation with microvessel density and p53 mutations, we retrospectively analyzed 81 primary esophageal carcinomas by immunohistochemical staining.

MATERIALS AND METHODS

Tissue samples and patients characteristics

A total of 81 patients with squamous cell carcinoma who underwent esophagectomy at Hanyang Medical Center between January 1989 and December 1999 were examined. Seventy-six were male and five were female. The patients ranged in age from 39 to 77 yr and the median age was 60

Table 1. Characteristics of patients with esophageal cancer

Characteristics	No. of patients (%)
Tumor stage (n=81)	
T1	4 (4.9)
T2	17 (21.0)
T3	42 (51.9)
T4	18 (22.2)
Nodal stage (n=81)	
N0	39 (48.1)
N1	42 (51.9)
Metastasis (n=81)	
M0	71 (87.7)
M1	10 (12.3)
Stage (n=81)	
I	3 (3.7)
IIA/B	28/4 (34.6/4.9)
III	36 (44.4)
IV	10 (12.3)
Grade (n=68)	
Well differentiated	13 (19.1)
Moderately differentiated	44 (64.7)
Poorly differentiated	11 (16.2)
Tumor size (n=79)	
<5 cm	46 (58.2)
>5 cm	33 (41.8)
Venous/lymphatic invasion (n=78)	
Negative	16 (20.5)
Positive	62 (79.5)
VEGF (n=80)	
Negative	39 (48.7)
Positive	41 (51.3)
p53 (n=79)	
Negative	38 (48.1)
Positive	41 (51.9)
Microvessel density (n=81)	
Median (range)	59.0 (13.3-179.0)
Relapse (n=81)	37/81 (45.7)
Death (n=81)	35/81 (43.2)
Overall survival (months) (n=81)	
Median (range)	11.8 (0.4-128)

yr. Cases of adenocarcinoma from Barrett's esophagus were excluded. Six cases of basaloid squamous cell carcinoma and one case of carcinosarcoma were included. Tumor staging was based on the pTNM pathological classification system. They included three patients with stage I, 28 with stage IIA, four with stage IIB, 36 with stage III, and ten with stage IV. Histological grades, tumor stages, lymph node metastasis, and the depth of invasion are shown in Table 1. Information concerning the date of initial diagnosis, other clinical characteristics, and death were obtained by a retrospective study. Subjects were followed up until any of the followings: the date of death, the last date they were known to be alive, or the end of the follow-up. Observations were censored either at the date of last follow-up or at the last date of the follow-up period if death had not occurred. The median duration of follow-up for surviving patients was 16.25 (3.9-128.0) months.

Immunohistochemical staining

The avidin-biotin complex (ABC) method was used for the immunostaining. Formalin-fixed paraffin-embedded tissue blocks were sectioned at 4- μ m thickness and immunostaining was performed according to following methods. Sections were deparaffinized in xylene for 10 min three times and rehydrated in serial graded alcohol in the following order: 100% for 5 min, 90% for 5 min, 70% for 5 min, and 50% for 5 min. Antigen retrieval was performed for VEGF and p53. For antigen retrieval, we microcooked the prepared 10 mM citrate buffer (pH 6.0) in a microwave for 3 min from the boiling point. The slides were put on the preheated citrate buffer and microwaved for 2 min three times. The container was put in a margin and cooled for 20 min at room temperature. The slides were washed in phosphate-buffered saline (PBS) once for 5 min. Endogenous peroxidase activity was blocked by 3% hydrogen peroxide in a 45 mL methanol solution for 15 min. The slides were washed in 1 \times PBS for 5 min three times. All the slides were preincubated with two drops of normal blocking solution (goat serum) at 37°C for 20 min (100 μ L/slide). Cautions should be exercised not to let the slides dry. The antibodies used were a mouse monoclonal IgG antibody (Santa Cruz Biotech, U.S.A.) at a 1:50 dilution for VEGF, a mouse monoclonal IgG antibody (Santa Cruz) at a 1:100 dilution for p53, and a mouse monoclonal antibody (Immunotech, Cedex, France) at a 1:100 dilution for CD 34. Each antibody was incubated overnight at 4°C and washed in 1 \times PBS for 5 min twice. Each of the biotinylated secondary antibodies was added for 30 min at 37°C followed by the avidin-biotinylated peroxidase complex (Immunotech) for additional 30 min at room temperature. After washing with PBS, the samples were stained by 3,3'-diaminobenzidine (Immunotech). Counterstaining was performed with hematoxylin for 30 sec.

Evaluation of immunostaining for p53, VEGF, and microvessel density

All slides were coded and evaluated without knowledge of patients' identity or clinical status by two experienced pathologists. Each experiment was independently performed twice. More than 10% of nuclear staining was defined as positive for p53. VEGF can be expressed in various human tissues including esophageal squamous cells and stromal cells. To determine the expression status of VEGF in cancer cells, we examined adjacent normal squamous epithelium. In our experiment, VEGF expression was largely confined in basal part of the epithelium (Fig. 1) and was not exceed 30% of the squamous cells. Accordingly, we considered positive for VEGF if more than 30% of tumor cells stained in their cytoplasm with stronger intensity than nonspecific background staining.

The degree of angiogenesis was determined by the MVD in defined areas of tissue sections according to the method

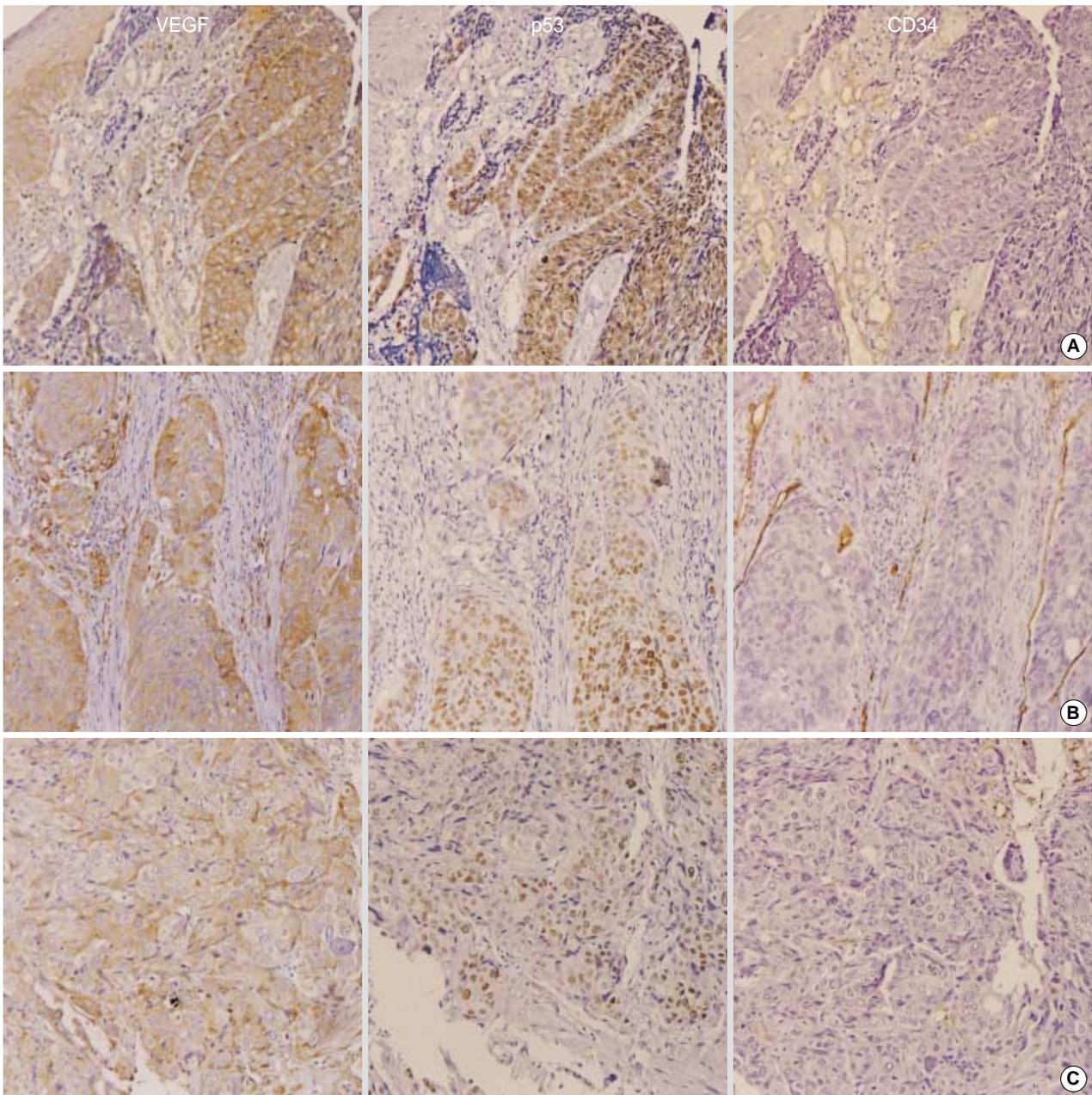


Fig. 1. A panel of immunohistochemical stainings for VEGF, p53, and CD34 in esophageal squamous cell carcinoma (original magnification, $\times 200$). (A) high VEGF and p53 expressions in the tumor cells are associated with a high vascular density visualized by CD34. (B) low VEGF and moderate p53 expressions are confined in the peripheral cells of the tumor nests with low vascular density. (C) moderate VEGF and p53 expressions are scattered in the tumor nests with low vascular density.

of Weidner et al. (24). All slides were coded and evaluated by an experienced pathologist without knowledge of patient's identity or clinical status. Each microvessel counting was performed twice. Each slide was first scanned at $\times 100$ magnification to determine three "hot spots" defined as areas with the maximum number of microvessels. The slides were then examined at $\times 200$ magnification. Microvessels were counted within the area defined in each of the three hot spots. Areas

of staining with no discrete breaks were counted as a single vessel. Microvessel density was estimated by adding the number of vessels in each of the three hot spots and then expressed as the mean number of vessels.

Statistical analysis

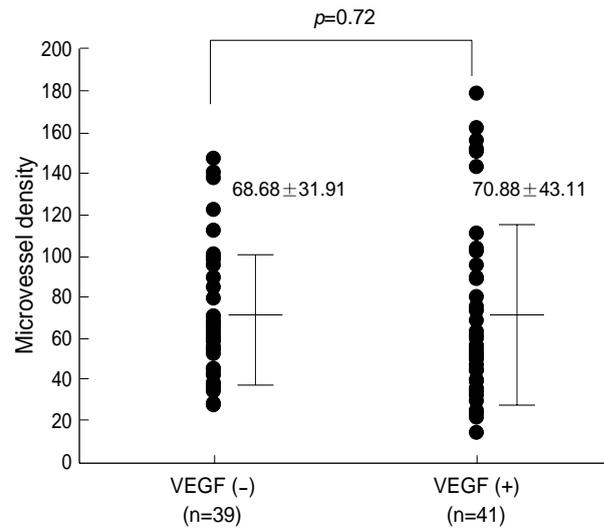
Statistical significance was evaluated using the Mann-Whit-

Table 2. VEGF (n=80) and p53 (n=79) expression in esophageal cancer patients

	VEGF (-)	VEGF (+)	p value	p53 (-)	p53 (+)	p value
Age (years)						
<60	19	21	0.823	19	20	0.914
>60	20	20		19	21	
Sex						
Male	36	39	0.603	33	41	0.016
Female	3	2		5	0	
p53						
Negative	16	22	0.358			
Positive	21	19				
VEGF						
Negative				16	21	0.358
Positive				22	19	
Tumor size						
<5 cm	24	22	0.645	22	23	0.862
>5 cm	15	17		15	17	
Grade						
WD*	8	5	0.564	8	5	0.354
MD	20	24		19	24	
PD	6	5		7	4	
Depth of invasion						
T1	3	1	0.458	2	1	0.453
T2	9	8		10	7	
T3/T4	27	32		26	33	
Lymph node						
Negative	19	19	0.832	19	17	0.447
Positive	20	22		19	24	
Metastasis						
Negative	34	36	0.933	35	34	0.220
Positive	5	5		3	7	
Stage						
I/IIA/IIIB	17	18	0.978	19	14	0.153
III/IV	22	23		19	27	
Venous/lymphatic invasion						
Negative	5	10	0.204	7	9	0.835
Positive	32	30		28	32	
Relapse						
No	20	23	0.666	24	20	0.199
Yes	19	18		14	21	
Death						
No	24	22	0.476	19	26	0.229
Yes	15	19		19	15	

*WD: well differentiated; MD: moderately differentiated; PD: poorly differentiated.

ney U test for independent groups. Survival curves were calculated using the Kaplan-Meier method and compared with other prognostic variables using the log-rank test. Correlation between variables was assessed by the Pearson's coefficient (r). Univariate analysis and multivariate stepwise Cox's regression analyses were performed to identify prognostic factors for survival. All statistical analyses were two-sided at a significance level of $p=0.05$, and performed using SPSS 10^R statistical software.

**Fig. 2.** Microvessel density in VEGF-positive and VEGF-negative esophageal carcinomas.

RESULTS

The expression of VEGF was identified mainly in the cytoplasm of the cancer cells, and 41 (51.3%) of the 80 cases were evaluated as VEGF-positive. The representative data are shown in Fig. 1. When the patients were divided into two groups, that is, VEGF-positive and negative groups, there were no significant differences in clinicopathological findings between the two groups according to the results from the χ^2 analysis (Table 2). Since all of the pT4 cases underwent palliative resection, cases of esophagectomy with thoracotomy for radical lymphadenectomy in pT1b to pT3 cases were selected and compared. The average number of metastatic lymph nodes at surgery in this group of patients was 1.17 in the patients with VEGF-negative tumors and 2.18 in those with VEGF-positive tumors and there was no statistical significance ($p=0.46$). The recurrence rate was 48.7% (19 of 39) in the VEGF-negative group and 43.9% (18 of 41) in the VEGF-positive group.

Eighty-one cases were evaluated for MVD. The range of MVD in esophageal cancer was 13.3-179.0 and the median number was 59.0. The representative data are shown in Fig. 1. The mean MVD in VEGF-negative group was 68.68 ± 31.91 (mean \pm SE) and that in VEGF-positive group was 70.88 ± 43.11 (mean \pm SE). MVD in the patients with VEGF-positive tumors was higher than in those with VEGF-negative tumors, but the difference was not significant ($p=0.72$) (Fig. 2).

The p53 expression was evaluated in 79 of 81 patients. Forty-one of 79 (51.9%) were p53 positive. There were no significant differences in various clinicopathological findings, such as age, tumor size, histological grade, depth of invasion, lymph node metastasis, stage, and venous or lymphatic invasion between p53-positive and negative groups. The recur-

rence rate and death rate were 36.8% (14 of 38) and 50.0% (19 of 38) in the p53-negative group and 51.2% (21 of 41) and 36.5% (15 of 41) in the p53-positive group, respectively ($p=0.199$; 0.229) (Table 2). When we analyzed the correlation between the VEGF and p53 expression, there was no association between two groups.

By univariate analysis, depth of invasion, lymph node metastasis, stage, tumor size, and distant metastasis were correlated with overall survival (Table 3). Among those variables,

Table 3. Univariate analysis of factors that influence the overall survival

Factors	Patients (n)	Median survival (m)	<i>p</i> value
Depth of invasion (n=81)			
T1/T2	21		0.044
T3/T4	60	27.4	
Lymph node (n=81)			
Negative	39	60.4	0.040
Positive	42	27.9	
Metastasis (n=81)			
Negative	71	43.5	0.009
Positive	10	4.4	
Stage (n=81)			
I/IIA/IIB	35	60.4	0.034
III/IV	46	19.2	
Tumor size (n=79)			
<5 cm	46	53.9	0.053
>5 cm	33	17.8	
VEGF (n=80)			
Negative	39	53.9	0.313
Positive	41	39.6	
p53 (n=79)			
Negative	38	27.9	0.414
Positive	41	39.8	

the presence of distant metastasis was the most significant factor for the survival. For the patients with distant metastasis, the median survival was only 4.4 months, compared with 43.5 months for patients without metastasis. However, we could not find any difference in overall survival between VEGF-positive and negative groups ($p=0.3129$) (Fig. 3A). Same findings were noted in the p53 expression ($p=0.4144$) (Fig. 3B).

Since MVD is a continuous variable, Cox's regression hazard model was used for the analysis. By multivariate analysis, the presence of distant metastasis was the only factor that influences overall survival (Table 4).

DISCUSSION

In this study, we analyzed the relationships between VEGF expression, MVD, p53, and clinicopathological features in

Table 4. Multivariate analysis of factors that influence the overall survival

Factors	<i>b</i>	SE (<i>b</i>)*	<i>p</i> value	Odd ratio (95% CI)*
T (T3/4 vs T1/2)	0.71	0.48	0.14	2.04 (0.79-5.24)
N (N1 vs N0)	0.214	0.40	0.60	1.24 (0.56-2.73)
M (M1 vs M0)	1.29	0.50	0.01	3.65 (1.36-9.78)
Tumor size (>5 cm vs <5 cm)	0.48	0.37	0.20	1.61 (0.78-3.35)
VEGF (positive vs negative)	0.24	0.37	0.53	1.27 (0.61-2.63)
p53 (positive vs negative)	-0.55	0.38	0.15	0.58 (0.27-1.23)

Cox's proportional hazards regression model is as follows: $h(t) = h_0(t) \exp(\beta_1 \text{ T stage} + \beta_2 \text{ lymph node} + \beta_3 \text{ distant metastasis} + \beta_4 \text{ tumor size} + \beta_5 \text{ VEGF} + \beta_6 \text{ p53} + \beta_7 \text{ microvessel density})$.

*SE: standard error; 95% CI: 95% confidence interval.

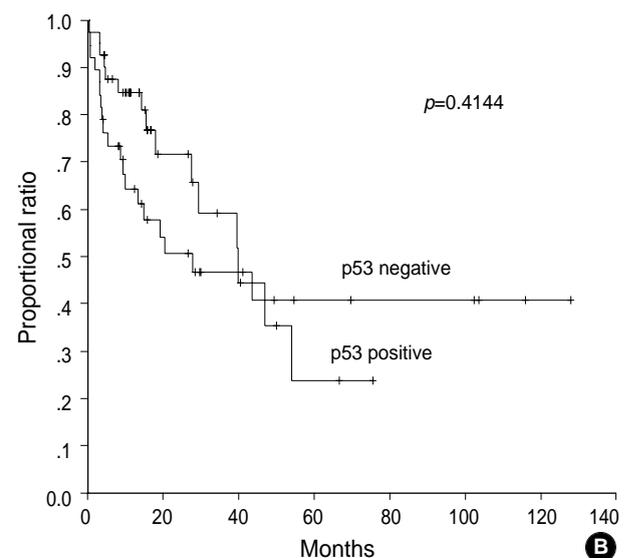
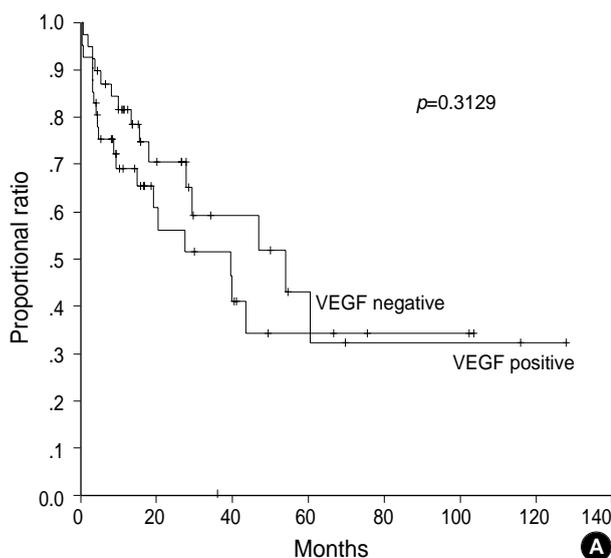


Fig. 3. (A) Overall survival for VEGF-positive and VEGF-negative groups in esophageal carcinomas. (B) Overall survival for p53-positive and p53-negative groups in esophageal carcinomas.

esophageal carcinomas. VEGF expression was noted in more than half of the patients and these data are consistent with the findings of the previous reports (24-28). However, the association between VEGF expression and clinicopathological findings is controversial. Some reports showed that lymphatic and/or venous invasion, tumor stage, or histological grade was correlated with VEGF expression (24-27). However, recently Shih et al. (28) reported that VEGF expression was not correlated with any of the clinical features, such as, tumor size, lymph node metastasis, histological grade, and lymphatic or venous invasion. We could not find any correlation between VEGF expression and the clinico-pathological findings, either. In this study, the median number of lymph node metastasis in the VEGF-positive group was slightly higher than that of the VEGF-negative group. It has been reported by Shih et al. that the average number of metastatic lymph nodes at surgery was 5.6 in the patients with VEGF-positive tumors and 3.0 in those with VEGF-negative tumors, and was significantly higher in those with VEGF-positive tumors.

To investigate the association between VEGF expression and tumor angiogenesis, we examined MVD immunohistochemically using anti-CD34 antibody. In the present study, the MVD in esophageal carcinoma tissues had a wide range, and the MVD of VEGF-positive carcinomas tended to be higher than that of VEGF-negative carcinomas. These results suggest that VEGF may be one of the key angiogenic factors, and promotes tumor angiogenesis in esophageal carcinoma tissues, in the same way as previously described in other carcinomas. However, we could not find a significant correlation between VEGF expression and MVD, as compared with previous reports (25-27). Several studies also reported that there was no correlation between VEGF expression and MVD (24, 28).

Factors that regulate VEGF expression in tumor and non-tumor cells have been investigated. Hypoxia has been known to be one of the most important mediators inducing the increase of VEGF. Mutations of the *ras* and *p53* genes have been shown to up-regulate the VEGF expression (22, 23). Some reports showed that mutations in this gene might be connected with angiogenesis by regulating VEGF expression in human cancers (29, 30). Although in this study the *p53* expression was noted in 51.9% of esophageal carcinomas, there was no significant correlation between VEGF expression and accumulation of *p53* protein in esophageal carcinomas. Recent study by Shimada et al. also reported that *p53* and VEGF were highly expressed in esophageal carcinoma tissues by immunohistochemical analysis, however, by multivariate analysis both molecular and biological markers were not associated with poor survival. Rather, cyclin D1, E-cadherin, and epidermal growth factor receptor were revealed to have prognostic relevance for survival and recurrence (31). These results suggest that numerous cytokines and growth factors, such as epidermal growth factor, platelet-derived

growth factor, transforming growth factor β , and insulin-like growth factors produced by tumor and normal cells may affect the VEGF expression.

The prognostic factors influencing disease-free and overall survival in esophageal carcinomas are based on both the histological type and the tumor stage. In the present study, by univariate analysis, tumor stage, depth of invasion, lymph node metastasis, distant metastasis, and tumor size were correlated with poor overall survival. However, the distant metastasis was the only prognostic factor affecting disease-free and overall survival in this cohort of esophageal carcinoma patients by multivariate analysis. This finding is inconsistent with other studies demonstrating that tumor stage and lymph node metastasis are the important prognostic factors. This can be explained by the facts that our study was a retrospective one and many patients underwent palliative esophageal surgery, so that the definitive lymph node dissection and accurate pathological staging were not possible in these patients. However, even if we had selected patients under curative surgery, we could not have found any specific prognostic findings. These conflicting results should be verified by prospective studies with a large number of patients. Previous studies have demonstrated that the angiogenesis is associated with the prognosis of patients with several malignancies. We followed the patients to determine whether a higher vessel count and VEGF expression could predict the risk for recurrence. We found that the prognosis of patients was not associated with the microvessel density or VEGF positivity. These findings suggest that the angiogenesis is not simply controlled by the presence of VEGF but may be mediated by other angiogenic factors. Since the angiogenic process is complex, additional studies concerning other angiogenic regulators are warranted.

In conclusion, this study has demonstrated that VEGF is highly expressed in human esophageal carcinomas. Since the VEGF expression is not correlated with the *p53* expression, MVD or clinicopathological findings, further studies with other angiogenic molecules are needed to determine the role in esophageal carcinomas.

ACKNOWLEDGMENTS

We thank Dr Moran Ki for her help with the statistical analysis.

REFERENCES

1. Folkman J. *Seminars in medicine of the Beth Israel Hospital, Boston. Clinical applications of research on angiogenesis. N Engl J Med* 1995; 333: 1757-63.
2. Folkman J. *New perspectives in clinical oncology from angiogenesis research. Eur J Cancer* 1996; 14: 2534-9.

3. Dickinson AJ, Fox SB, Persad RA, Hollyer J, Sibley GN, Harris AL. *Quantification of angiogenesis as an independent predictor of prognosis in invasive bladder carcinomas. Br J Urol 1994; 74: 762-6.*
4. Fox SB. *Tumor angiogenesis and prognosis. Histopathology 1997; 30: 294-301.*
5. Weidner N, Semple JP, Welch WR, Folkman J. *Tumor angiogenesis and metastasis-correlation in invasive breast carcinoma. N Engl J Med 1991; 324: 1-8.*
6. Nicosia RF, T'chao R, Leighton J. *Interaction between newly formed endothelial channels and carcinoma cells in plasma clot culture. Clin Exp Metastas 1986; 4: 91-104.*
7. Hamada J, Cavanaugh PG, Lotan O, Nicoloson G. *Separable growth and migration factors for large-cell lymphoma cells secreted by microvascular endothelial cells derived from target organs for metastasis. Br J Cancer 1992; 66: 349-54.*
8. Rak J, Filmus J, Kerbel RS. *Reciprocal paracrine interactions between tumor cells and endothelial cells; the angiogenesis progression hypothesis. Eur J Cancer 1996; 32: 2438-50.*
9. Gasparini G, Weidner N, Maluta S, Pozza F, Boracchii P, Mezzetti M, Testolin A, Bevilacqua P. *Intratumoral microvessel density and p53 protein; correlation with metastasis in head and neck squamous cell carcinoma. Int J Cancer 1993; 55: 739-44.*
10. Ellis LM, Fidler IJ. *Angiogenesis and breast cancer metastasis. Lancet 1995; 346: 388-90.*
11. Maeda K, Chung YS, Takatsuka S, Ogawa Y, Swada T, Yamashita Y, Onoda N, Kato Y, Nitta A, Arimoto Y, Kondo Y, Sowa M. *Tumor angiogenesis as a predictor of recurrence in gastric carcinoma. J Clin Oncol 1995; 13: 477-81.*
12. Wiggins DL, Granai CO, Steinhoff MM, Calabresi P. *Tumor angiogenesis as a prognostic factor in cervical carcinoma. Gynecol Oncol 1995; 56: 353-6.*
13. Fontanini G, Lucchi M, Vignati S, Mussi A, Ciardiello F, De Laurentiis M, De Placido S, Basolo F, Angeletti CA, Bevilacqua G. *Angiogenesis as a prognostic indicator of survival in non-small cell lung carcinoma: a prospective study. J Natl Cancer Inst 1997; 89: 881-6.*
14. Acenero MJ, Gonzalez JF, Gallego MG, Ballesteros PA. *Vascular enumeration as a significant prognosticator for invasive breast carcinoma. J Clin Oncol 1998; 16: 1684-9.*
15. Ferrara N, Hezel WJ. *Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. Biochem Biophys Res Commun 1989; 161: 851-8.*
16. Gospodarowicz D, Abraham JA, Schilling J. *Isolation and characterization of a vascular endothelial cell mitogen produced by pituitary-derived follicular stellate cells. Proc Natl Acad Sci USA 1989; 86: 7311-5.*
17. Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N. *Vascular endothelial growth factor is a secreted angiogenic mitogen. Science 1989; 246: 1306-9.*
18. Ferrara N, Houck K, Jakeman L, Leung DW. *Molecular and biological properties of the vascular endothelial growth factor family of proteins. Endocr Rev 1992; 13: 18-32.*
19. Toi M, Inada K, Suzuki H, Tominaga T. *Tumor angiogenesis in breast cancer; its importance as prognostic indicator and the association with vascular endothelial growth factor expression. Breast Cancer Res Treat 1995; 36: 193-204.*
20. Maeda K, Chung YS, Ogawa Y, Takatsuka S, Kang SM, Ogawa M, Swada T, Sowa M. *Prognostic value of vascular endothelial growth factor expression in gastric carcinoma. Cancer 1996; 77: 858-63.*
21. Tanigawa N, Amaya H, Matsumura M, Shimonatsuya T. *Correlation between expression of vascular endothelial growth factor and tumor vascularity, and patient outcome in human gastric carcinoma. J Clin Oncol 1997; 15: 826-32.*
22. Kieser A, Weich HA, Brandner G, Marme D, Kolch W. *Mutant p53 potentiates protein kinase C induction of vascular endothelial growth factor expression in primary esophageal squamous cell carcinoma. Oncogene 1994; 9: 963-9.*
23. Mukhopadhyay D, Tsolkas I, Sukhatme VP. *Wild type p53 and v-Src exert opposing influences on human vascular endothelial growth factor gene expression. Cancer Res 1995; 55: 6161-5.*
24. Uchida S, Shimada Y, Watanabe G, Tanaka H, Shibagaki I, Miyahara T, Ishigami S, Imamura M. *In oesophageal squamous cell carcinoma vascular endothelial growth factor is associated with p53 mutation, advanced stage and poor prognosis. Br J Cancer 1998; 77: 1704-9.*
25. Koide N, Nishio A, Kono T, Yazawa K, Igarashi J, Watanabe H, Nimura Y, Hanazaki K, Adachi W, Amano J. *Histochemical study of vascular endothelial growth factor in squamous cell carcinoma of the esophagus. Hepato-Gastroenterol 1999; 46: 952-8.*
26. Kitadai Y, Haruma K, Tokutomi T, Tanaka S, Sumii K, Carvalho M, Kuwabara M, Yoshida K, Hirai T, Kajuyama G, Tahara E. *Significance of vessel count and vascular endothelial growth factor in human esophageal carcinomas. Clin Cancer Res 1998; 4: 2195-200.*
27. Inoue K, Ozeki Y, Sukanuma T, Sugiura Y, Tanaka S. *Vascular endothelial growth factor expression in primary esophageal squamous cell carcinoma. Cancer 1997; 79: 206-13.*
28. Shih CH, Ozawa S, Ando N, Ueda M, Kitajima M. *Vascular endothelial growth factor expression predicts outcome and lymph node metastasis in squamous cell carcinoma of esophagus. Clin Cancer Res 2000; 6: 1161-8.*
29. Riedel F, Gotte K, Schwalb J, Schafer C, Hormann K. *Vascular endothelial growth factor expression correlates with p53 mutation and angiogenesis in squamous cell carcinoma of the head and neck. Acta Otolaryngol 2000; 120: 105-11.*
30. Strohmeyer D, Rossing C, Bauerfeind A, Kaufmann O, Schlechte H, Bartsch G, Loening S. *Vascular endothelial growth factor and its correlation with angiogenesis and p53 expression in prostate cancer. Prostate 2000; 45: 216-24.*
31. Shimada Y, Imamura M, Watanabe G, Uchida S, Harada H, Maki-no T, Kano M. *Prognostic factors of esophageal squamous cell carcinoma from the perspective of molecular biology. Br J Cancer 1999; 80: 1281-8.*