

Fatty Acid Patterns in Gastric Mucosa of Stomach Cancer Patients

Jiyoung Ahn¹, In Suh Park², Kyong Sik Lee³, Soo Yeon Kim¹, Eun Jung Chung⁴, Jiyoung Kim¹, Dae Jung Kim², Sun Yoon¹, and Yang Cha Lee-Kim¹

Departments of ¹Food and Nutrition, ²Institute of Gastroenterology, ³Surgery, Yonsei University, Seoul;

⁴Department of General Education, Kangnam University, Kyunggido, Korea.

$\omega 6$ and $\omega 3$ fatty acids are important cellular components and known to be involved in disease processes. However, few studies have focused on mucosa fatty acid in human gastric cancer. The purpose of this study was to investigate how fatty acid patterns of mucosa are altered in gastric cancer. Fatty acids were analyzed by gas chromatography and their relative compositions (%) were determined and evaluated both in mucosa total-fatty acids and in phospholipid-fatty acids in paired cancerous and non-cancerous gastric cancer tissues (n=18).

The level of arachidonic acid (20:4 $\omega 6$, AA) appeared significantly higher both in phospholipid-fatty acids ($p < 0.05$) and in total-fatty acids ($p < 0.001$) in cancerous mucosa compared to non-cancerous mucosa. The $\omega 6/\omega 3$ fatty acid ratio of phospholipid-fatty acids was also significantly higher in cancerous mucosa. The higher level of AA in cancerous tissue can be partially explained by the higher ratio of 20:4 $\omega 6/20:3 \omega 6$ (desaturation index) and the lower ratio of 22:4 $\omega 6/20:4 \omega 6$ (elongation index). The change in the relative composition of arachidonic acid may influence the production of prostaglandins and related metabolites, which regulate cell differentiation and proliferation. The findings of this study with respect to fatty acid changes, especially in terms of arachidonic acid metabolism, may be of relevance in the understanding of the roles of specific fatty acids and possibly of eicosanoids in gastric cancer.

Key Words: Gastric cancer, fatty acid pattern, $\omega 6/\omega 3$ fatty acid ratio, arachidonic acid

INTRODUCTION

Phospholipid-fatty acids (PL-FAs) are important structural and functional components of the cell and have profound effects on membrane fluidity

and cell metabolism.¹ Changes in lipid composition also alter the membrane transduction pathways of external signals and may consequently modulate the response of tumor cells to growth factors, thereby modifying the evolution of cancer.^{2,3} Membrane FA composition of tumor cells results from the type of FAs available to the tumor cell, together with the specific metabolic properties of the tumor tissue, which includes alterations to enzyme pathways involved in FA uptake, activation, acylation, desaturation/elongation and oxidation.⁴

Dietary FAs can influence mucosa FAs.^{5,6} In experimental animal models of mammary carcinogenesis, high dietary $\omega 6$ polyunsaturated FAs (PUFAs) were shown to stimulate mammary tumor growth, development and metastases, whereas a long chain $\omega 3$ PUFA enriched diet inhibited tumor growth.⁷ Therefore, dietary long chain $\omega 3$ PUFAs seem to oppose the stimulation of tumor growth induced by $\omega 6$ FAs in the rat.⁸ However, it is not known how dietary FAs influence the system. The wide range of biochemical conditions among cancer cells⁹ makes it difficult to understand FA metabolism.

Despite the substantial amount of data on the role of dietary fats in cancer, few studies have focused on mucosal FA in human gastric cancer. In this research, the different FA patterns in human gastric cancer mucosa were investigated. The relative compositions (%) of FAs were analyzed both in the total mucosa and in the phospholipids in paired non-cancerous and cancerous gastric cancer tissues.

Received July 20, 2000

Accepted January 17, 2001

* This work was supported by the Brain Korea 21 Project in 2001.

MATERIALS AND METHODS

Subjects

Nine patients diagnosed with gastric cancer at the Yonsei Medical Center in Seoul, Korea, participated in this study. Gastric tissue, which otherwise would have been discarded, was obtained after surgical resection of the stomach (gastrectomy). Cancerous and non-cancerous (at least 10 cm away from the malignant site) gastric tissue samples weighing 2.0~3.0 g of full thickness were dissected from the tissues obtained by surgery. The tissue samples were frozen immediately in liquid nitrogen and stored at -70°C until use. Patients characteristics are shown in Table 1.

This work was performed after obtaining approval from the Human Investigation Committee of the Yonsei University College of Medicine.

Fatty acid analysis

Preparations of the homogenates of stomach mucosal (cancerous and non-cancerous) tissues for fatty acid analyses were conducted as follows. The gastric mucosa was scraped with a scalpel, and approximately 50 mg (wet weight) of the scraped mucosa was used for analysis. Mucosal lipids were extracted using the method of Folch et al.¹⁰ Phospholipids from the mucosa were separated by thin layer chromatography (TLC). The phospholipid portion in the silica gel was scraped off immediately after the TLC procedure and methylated, using the method of Lepage, Roy.¹¹ Lipid extract was obtained using the Folch method, and the fatty acid methyl esters were

quantified by gas liquid chromatography (Hewlett Packard 6890A GC). For the gas chromatographic separation, we used a bonded fused-silica capillary column (OmegawaxTM 250). The GLC oven temperature was held at 180°C for 5 min and increased to 210°C in 2°C/min increments. The temperature of the injection and detector ports was maintained at 280°C. Helium was used as a carrier gas, at a flow rate 0.8 ml/min and with a split ratio of 10:1. Methyl esters of the various fatty acids were identified by comparison with GLC reference standards (#GLC-87A), OmegawaxTM test mix (#4-8476), and PUFA- 2 (#4-7015, Supelco, Bellefonte, PA, USA). The peaks of the standard FA fractions were quantified using a Hewlett Packard 3365A series III Chemstation integrator. All GC analyses were performed in duplicate.

Statistical analysis

Results were analyzed using the SAS software package and the significance of the difference between the mean values of paired cancerous and non-cancerous gastric mucosa was tested using the paired t-test. All values were expressed as mean \pm SEM.

RESULTS

Fatty acid patterns in tissue total-fatty acids (Total-FAs)

The composition of the total-fatty acid (total-FA) is shown in Table 2. The relative compo-

Table 1. Characterization of Patients with Gastric Cancer

| Patient No. | Sex | Age | Height (cm) | Weight (kg) | BMI (Wt/Ht ²) | Pathology |
|----------------|-----|---------------|-----------------|---------------|---------------------------|---|
| 1 | M | 66 | 150 | 55 | 24.4 | Adenocarcinoma, poorly differentiated |
| 2 | M | 36 | 168 | 53 | 18.8 | Adenocarcinoma, poorly differentiated |
| 3 | M | 57 | 163 | 44 | 16.6 | Adenocarcinoma, poorly differentiated |
| 4 | M | 42 | 168 | 55 | 19.5 | Adenocarcinoma, poorly differentiated |
| 5 | M | 57 | 167 | 48 | 17.2 | Adenocarcinoma, poorly differentiated |
| 6 | M | 62 | 167 | 68 | 24.4 | Adenocarcinoma, moderately differentiated |
| 7 | M | 59 | 164 | 55 | 20.4 | Adenocarcinoma, lymphoepithelioma-like carcinoma type |
| 8 | F | 68 | 162 | 61 | 23.2 | Adenocarcinoma, poorly differentiated |
| 9 | F | 68 | 153 | 58 | 24.8 | Adenocarcinoma, well differentiated |
| Mean \pm SEM | | 57 \pm 1.25 | 1.64 \pm 0.01 | 55 \pm 0.77 | 20.9 \pm 0.4 | |

Table 2. Relative Fatty Acid Composition of Total-fatty Acids in Gastric Mucosa

| | Relative Composition(%) | | | Cancer/Non-cancer |
|--------------------------------------|-------------------------|--------------|---------|-------------------|
| | Non-cancer* | Cancer* | p value | |
| 12:0 | 0.36 ± 0.06 | 0.70 ± 0.06 | 0.001 | 1.94 |
| 14:0 | 1.27 ± 0.25 | 0.96 ± 0.14 | | 0.76 |
| 16:0 | 21.21 ± 0.67 | 20.34 ± 0.70 | | 0.96 |
| 18:0 | 8.01 ± 1.02 | 11.08 ± 0.87 | 0.04 | 1.38 |
| 20:0 | 0.22 ± 0.01 | 0.30 ± 0.03 | | 1.36 |
| 22:0 | 0.26 ± 0.04 | 0.41 ± 0.04 | 0.02 | 1.58 |
| 24:0 | 0.62 ± 0.27 | 0.40 ± 0.52 | | 0.65 |
| SFA | 31.55 ± 1.03 | 33.96 ± 0.50 | 0.05 | 1.08 |
| 16:1 ω 7 | 5.82 ± 0.34 | 3.05 ± 0.51 | 0.0004 | 0.52 |
| 18:1 ω 7 | 2.81 ± 0.27 | 2.59 ± 0.24 | | 0.92 |
| 18:1 ω 9 | 29.23 ± 1.14 | 20.79 ± 1.76 | 0.001 | 0.71 |
| 20:1 | 0.52 ± 0.06 | 0.33 ± 0.07 | 0.05 | 0.63 |
| 22:1 | 0.15 ± 0.04 | 0.15 ± 0.03 | | 1.00 |
| 24:1 | 0.24 ± 0.07 | 0.66 ± 0.09 | 0.002 | 2.75 |
| MUFA | 38.31 ± 1.54 | 27.56 ± 2.30 | 0.001 | 0.72 |
| 18:2 ω 6 | 12.14 ± 0.27 | 12.88 ± 0.85 | | 1.06 |
| 18:3 ω 6 | 0.12 ± 0.03 | 0.08 ± 0.01 | | 0.67 |
| 20:3 ω 6 | 0.71 ± 0.17 | 1.30 ± 0.14 | 0.02 | 1.83 |
| 20:4 ω 6 | 3.87 ± 0.37 | 7.69 ± 0.72 | 0.0005 | 1.99 |
| 22:4 ω 6 | 0.37 ± 0.06 | 0.66 ± 0.08 | 0.01 | 1.78 |
| 22:5 ω 6 | 0.14 ± 0.02 | 0.13 ± 0.01 | | 0.93 |
| $\Sigma \omega$ 6 | 19.30 ± 2.19 | 22.71 ± 1.54 | | 1.18 |
| 18:3 ω 3 | 0.29 ± 0.04 | 0.23 ± 0.03 | | 0.79 |
| 20:3 ω 3 | 0.08 ± 0.03 | 0.05 ± 0.01 | | 0.63 |
| 20:5 ω 3 | 0.77 ± 0.09 | 0.95 ± 0.13 | | 1.23 |
| 22:5 ω 3 | 0.73 ± 0.07 | 1.04 ± 0.08 | 0.01 | 1.42 |
| 22:6 ω 3 | 2.91 ± 0.22 | 3.43 ± 0.37 | 0.003 | 1.80 |
| $\Sigma \omega$ 3 | 4.78 ± 1.43 | 5.29 ± 0.49 | | 1.10 |
| PUFA | 23.48 ± 0.90 | 28.41 ± 1.78 | 0.002 | 1.35 |
| Others | 6.06 ± 1.20 | 10.48 ± 1.26 | | |
| TOTAL | 100.00 | 100.00 | | |
| $\Sigma \omega$ 6/ $\Sigma \omega$ 3 | 4.70 ± 0.10 | 4.20 ± 0.38 | | |

*Values are mean ± SEM.

The statistical significances between non-cancerous tissues were calculated using Student's t-test for paired data.

sitions of saturated fatty acids (SFA) ($p < 0.05$) and PUFA ($p < 0.05$) were significantly higher, and the monounsaturated fatty acid (MUFA, $p < 0.001$) levels were lower in cancerous mucosa than in the paired non-cancerous mucosa. In particular, 16:1, 18:1 and 20:1 were significantly lower in cancerous mucosa than in non-cancerous mucosa. The higher PUFA compositions ($p < 0.01$) in the total-FAs of cancerous tissue were mostly due to the longer chain PUFAs (20:3 ω 6, 20:4 ω 6, 22:4 ω 6, 22:5 ω 3, and 22:6 ω 3). While the ratio of polyunsaturated fatty acids/saturated fatty acids (P/S)

ratios was significantly higher in the cancerous mucosa, the ratio of ω 6/ ω 3 was similar in the two paired groups.

Fatty acid patterns in tissue phospholipid-fatty acids (PL-FAs)

In terms of phospholipid fatty acids, 3 series fatty acids showed significantly ($p < 0.05$) lower levels in cancerous mucosa than in non-cancerous tissue, although no significant differences were found in relative compositions of SFA, MUFA,

and PUFA. Among the $\omega 6$ fatty acids, arachidonic acid, the major precursor of the series II eicosanoids, was significantly higher ($p < 0.01$) in the cancerous mucosa (Table 3, Fig. 1). The same result was obtained in total-FAs as in phospholipid fatty acids.

The $\omega 6/\omega 3$ FA ratio of PL-FAs was also significantly higher in cancerous mucosa ($p < 0.01$) than in non-cancerous mucosa. This difference may be explained by higher levels of 20:4 $\omega 6$ and 22:5 $\omega 6$ and lower levels of 20:5 $\omega 3$ (EPA) and 22:6 $\omega 3$ (DHA) in the cancerous mucosa.

To examine to what extent desaturation mechanisms may account for the changes of fatty acid metabolism in cancerous mucosa, desaturation and elongation indices of PL-FAs were calculated (Table 4, Fig. 2). Though there was no apparent trend in desaturation or elongation indices in total-FAs, the delta-6, 5, and 4-desaturation indices calculated from the 18:3/18:2 $\omega 6$, 20:4/20:3 $\omega 6$, and 22:5/22:4 $\omega 6$ ratios, respectively, of the PL-FAs were all significantly higher and the elongation indices were all significantly lower in cancerous mucosa than in non-cancerous mucosa.

Table 3. Relative Fatty acid Composition of Phospholipid-fatty Acids in Gastric Mucosa

| | Relative composition (%) | | p value | Cancer/Non-cancer |
|-----------------------------------|--------------------------|------------------|---------|-------------------|
| | Non-cancer* | Cancer* | | |
| 12:0 | 0.85 \pm 0.14 | 0.35 \pm 0.09 | 0.009 | 0.41 |
| 14:0 | 1.34 \pm 0.16 | 1.10 \pm 0.17 | | 0.82 |
| 16:0 | 29.05 \pm 1.88 | 31.42 \pm 1.42 | | 1.08 |
| 18:0 | 16.60 \pm 0.83 | 20.57 \pm 0.94 | 0.006 | 1.24 |
| 20:0 | 0.60 \pm 0.05 | 0.59 \pm 0.04 | | 0.98 |
| 22:0 | 0.85 \pm 0.13 | 1.13 \pm 0.25 | | 1.33 |
| 24:0 | 0.65 \pm 0.30 | 0.38 \pm 0.07 | | 0.58 |
| SFA | 49.72 \pm 2.48 | 55.54 \pm 1.88 | | 1.12 |
| 16:1 $\omega 7$ | 0.84 \pm 0.22 | 0.48 \pm 0.16 | | 0.57 |
| 18:1 $\omega 7$ | 1.02 \pm 0.17 | 0.97 \pm 0.29 | | 0.95 |
| 18:1 $\omega 9$ | 3.10 \pm 0.86 | 2.27 \pm 0.58 | | 0.73 |
| 20:1 | 0.39 \pm 0.05 | 0.72 \pm 0.16 | 0.002 | 1.85 |
| 22:1 | 1.23 \pm 0.23 | 0.24 \pm 0.05 | | 0.20 |
| 24:1 | 0.46 \pm 0.15 | 0.38 \pm 0.12 | | 0.83 |
| MUFA | 6.97 \pm 1.16 | 5.06 \pm 1.03 | | 0.71 |
| 18:2 $\omega 6$ | 1.35 \pm 0.25 | 1.48 \pm 0.28 | | 1.10 |
| 18:3 $\omega 6$ | 0.18 \pm 0.02 | 0.45 \pm 0.14 | | 2.50 |
| 20:3 $\omega 6$ | 0.51 \pm 0.08 | 0.32 \pm 0.08 | | 0.63 |
| 20:4 $\omega 6$ | 0.62 \pm 0.07 | 1.23 \pm 0.08 | 0.02 | 1.89 |
| 22:4 $\omega 6$ | 0.43 \pm 0.09 | 0.50 \pm 0.18 | | 1.16 |
| 22:5 $\omega 6$ | 9.38 \pm 1.31 | 11.73 \pm 1.00 | | 1.25 |
| $\Sigma \omega 6$ | 12.60 \pm 1.41 | 15.52 \pm 0.98 | | 1.23 |
| 18:3 $\omega 3$ | 0.18 \pm 0.02 | 0.34 \pm 0.08 | | 1.89 |
| 20:3 $\omega 3$ | 0.63 \pm 0.11 | 0.31 \pm 0.07 | 0.04 | 0.49 |
| 20:5 $\omega 3$ | 2.66 \pm 0.99 | 0.60 \pm 0.05 | | 0.23 |
| 22:5 $\omega 3$ | 0.40 \pm 0.06 | 0.52 \pm 0.15 | | 1.30 |
| 22:6 $\omega 3$ | 1.49 \pm 0.34 | 1.01 \pm 0.09 | | 0.68 |
| $\Sigma \omega 3$ | 5.32 \pm 0.41 | 2.78 \pm 0.10 | 0.04 | 0.43 |
| PUFA | 17.91 \pm 0.76 | 18.26 \pm 1.06 | | 1.02 |
| Others | 25.42 \pm 1.20 | 21.14 \pm 1.26 | | |
| TOTAL | 100.00 | 100.00 | | |
| $\Sigma \omega 6/\Sigma \omega 3$ | 2.45 \pm 0.73 | 5.57 \pm 0.96 | 0.003 | 2.27 |

*Values are mean \pm SEM.

The statistical significances between non-cancerous tissues were calculated using Student's t-test for paired data.

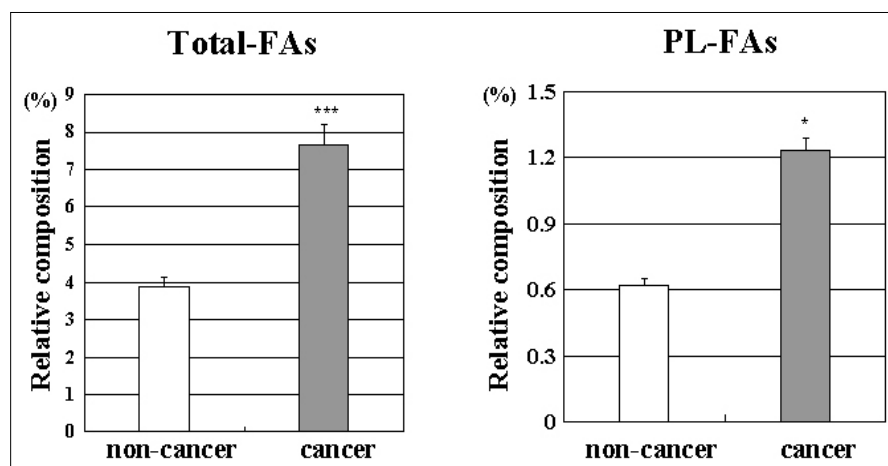


Fig. 1. Relative composition of arachidonic acid in cancerous and non-cancerous gastric mucosa. * $p < 0.05$, *** $p < 0.001$. Total-FAs: total fatty acids, PL-FAs: phospholipid fatty acids. The statistical significances between non-cancerous and cancerous tissues were calculated using Student's t-test for paired data.

Table 4. Desaturation & Elongation Index of Total-fatty Acids and Phospholipid-fatty Acids

| | Total-fatty acids | | | | Phospholipid-fatty acids | | | |
|---------------------------------|-------------------|------------------|---------|---------------------|--------------------------|-------------------|---------|---------------------|
| | Non-cancer* | Cancer* | p value | Cancer / Non-cancer | Non-cancer* | Cancer* | p value | Cancer / Non-cancer |
| 18:3 ω 6/18:2 ω 6 | 0.01 \pm 0.001 | 0.01 \pm 0.001 | | 1.00 | 0.17 \pm 0.04 | 0.38 \pm 0.10 | | 2.24 |
| 20:3 ω 6/18:3 ω 6 | 16.67 \pm 1.47 | 20.51 \pm 3.45 | 0.05 | 1.23 | 3.05 \pm 0.62 | 0.92 \pm 0.16 | 0.009 | 0.30 |
| 20:4 ω 6/20:3 ω 6 | 20.95 \pm 4.92 | 16.19 \pm 0.64 | | 1.20 | 1.87 \pm 0.41 | 5.79 \pm 1.64 | 0.04 | 3.10 |
| 22:4 ω 6/20:4 ω 6 | 0.1 \pm 0.06 | 0.08 \pm 0.01 | | 0.80 | 0.86 \pm 0.33 | 0.44 \pm 0.12 | | 0.51 |
| 22:5 ω 6/22:4 ω 6 | 0.459 \pm 0.02 | 0.21 \pm 0.02 | 0.03 | 0.46 | 30.62 \pm 6.74 | 51.82 \pm 19.15 | | 1.69 |

*Values are mean \pm SEM.

The statistical significances between non-cancerous tissues were calculated using Student's t-test for paired data.

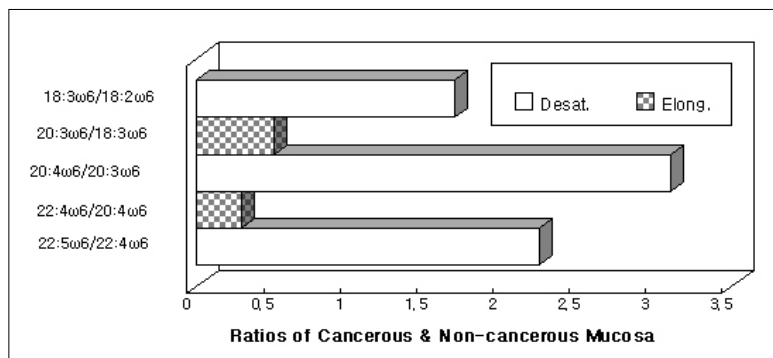


Fig. 2. Comparison between Desaturation and Elongation Index of PL-FAs in Cancerous and Non-cancerous Gastric Mucosa. Desat.: Desaturation, Elong.: Elongation PL-FAs: Phospholipid-fatty acids

The ratios of cancer/non-cancer values of all desaturation indices of ω 6 fatty acids were remarkably higher (range 1.69~3.10) in PL-FAs than the elongation indices. There is a possibility that once desaturation from 20:3 ω 6 to 20:4 ω 6 increases, the subsequent elongation from 20:4 ω 6 to 22:4 ω 6 decreases, resulting in a larger amount of arachidonic acid (20:4 ω 6 AA).

DISCUSSION

The present study investigated the relative compositions (%) of FAs in both total-FAs and phospholipid (PL)-FAs in cancerous and non-cancerous gastric mucosa. The distribution pattern of fatty acids in tissues was found to broadly parallel the plasma profile¹², which may be the result of a combination of many different factors,

including biosynthesis, oxidation, esterification, turn-over rate, rate of conversion to eicosanoids, antioxidant systems and diet. This study ruled out the possibility of bias caused by different dietary FA compositions by comparing cancerous and non-cancerous mucosa in the same subjects.

In the total-FAs of cancerous mucosa, the relative compositions of SFAs were higher ($p < 0.05$) and those of MUFAs (e.g. 16:1 ω 7, 18:1 ω 9) were lower ($p < 0.001$) than those of non-cancerous mucosa. The fact that there was higher level of 18:0 and lower level of 16:1 ω 7 and 18:1 ω 9 in cancerous mucosa may be attributed to palmitic acid (16:0) being more converted to stearic acid (18:0) than to palmitoleic acid (16:1 ω 7) and stearic acid (18:0) being less converted to oleic acid (18:1 ω 9).

In terms of PL-FAs, the ω 6/ ω 3 FA ratio was significantly higher ($p < 0.01$) in cancerous mucosa than in non-cancerous mucosa. This high ratio of ω 6/ ω 3 FA was mainly due to a high level of AA and low levels of ω 3 fatty acids, such as 20:5 ω 3 (EPA) and 22:6 ω 3 (DHA). In a previous study⁷ the results showed that high ω 6 FA diets stimulate mammary tumor growth and development, as well as metastases, while long chain ω 3 FAs or 18:3 ω 3 enrichment of the diet inhibits tumor growth. Erickson KL, Hubbard,¹³ using a model of mammary tumor metastases, showed that mice fed ω 6 PUFA-rich diets showed increased in survival of metastatic cells. Other similar research has shown that dietary EPA and DHA caused increases in the low levels of EPA and decreases in the high content of AA in colon cancer mucosa and inhibited mucosa cancer cell growth¹⁴. According to Huang YC, et al.¹⁵, plasma phospholipid ω 6/ ω 3 FA ratio may be used as a nutritional compliance marker for chronic epithelial cell hyperproliferation. Among the saturated fatty acids of phospholipids, stearic acid (18:0), which was postulated to stimulate delta-5-desaturase, was found to be significantly higher in cancerous tissue.¹⁶

In this study, the fatty acid patterns in total fatty acids and in phospholipid-fatty acids were different. Membrane phospholipids are important reservoirs of the essential fatty acids for the body, and these may play important roles in the cell, for example they have a role in the maintenance of

membrane structure and in the regulation of signal transduction.¹⁶ Therefore, fatty acids in phospholipids may be more finely regulated in the normal physiological state, and any deviation from the normal pattern may be related to abnormal or disease states. The ratio of ω 6/ ω 3 may provide a good example for explaining the different patterns between total-FAs and PL-FAs. Although, no change was found in the ω 6/ ω 3 ratio of total mucosal FAs, a significant change was found in the total mucosal lipid subfraction, i.e. the phospholipid. The high level of ω 6 FA in cancerous tissue-PUFAs was mainly due to high AA levels, which is present in both total-FA and PL-FA. Arachidonic acid is usually incorporated into tissue PLs at carbon 2 and has to be released by the action of phospholipase A.

The AA content of the tissue is, therefore, related to the production of eicosanoids and related metabolites regulating cell differentiation and proliferation.^{17,18} It has been reported that AA can directly or synergistically activate some species of protein kinase C^{19,20} which affect membranes and Ca^{2+} availability, and activate cell differentiation and proliferation.²¹ The higher levels of AA, especially in the PL of cancerous tissue, can be partially explained by the higher ratio of 20:4 ω 6/20:3 ω 6 (desaturation) and the lower ratio of 22:4 ω 6/20:4 ω 6 (elongation) as shown in Figure 2. It is interesting to note that changes in the specific activity of desaturase will alter the regulation of all fatty acids, because of its key role in fatty acid metabolism, and that this is particularly the case in chronic diseases, such as cancer, and cardiovascular disease.²² The present study with gastric cancer showed that the ω 6-desaturation indices of PL-FAs were all higher in cancerous mucosa than in non-cancerous mucosa. There is clear evidence²³ of changes in membrane fluidity in diseases, which may be either a cause or an effect of alterations of the membrane phospholipid metabolism or membrane bound enzyme activity. It can be speculated that the existence of a compensatory mechanism, which is evident in the higher levels of phospholipid-fatty acid desaturase indices in cancerous mucosa, increases the availability of PUFA for membrane fluidity regulation.

In conclusion, the findings of this study on fatty

acid changes, especially on arachidonic acid metabolism, may be of relevance to the understanding of the roles of specific fatty acids and possibly of eicosanoids in gastric cancer. Further studies are needed to differentiate the roles of fatty acids in different phospholipid fractions, such as phosphatidyl ethanolamine (PE) and phosphatidyl inositol (PI) and to clarify the roles of fatty acids, eicosanoids, and enzymes involved in the process of cancer development.

Y. C. Lee-Kim, Ph.D.
 Department of Food and Nutrition,
 College of Human Ecology, Yonsei University,
 C.P.O. Box 8044, Seoul 120-749, Korea.
 Tel: 82-2-361-3118, Fax: 82-2-312-5229,
 E-mail: ycleekim@yonsei.ac.kr

REFERENCES

1. Rivers IP, Frankel TL. Essential fatty acid deficiency. *Br Med Bull* 1981;37:59-64.
2. Spector AA, Burns CP. Biological and therapeutic potential of membrane lipid modification in tumors. *Cancer Res* 1987;47:45-9.
3. Das UN, Madhavi N, Sravan Kumar G, Padma M, Sangeetha P. Can tumour cell drug resistance be reversed by essential fatty acids and their metabolites? *Prostaglandins Leukot Essent Fatty Acids* 1998;58:39-54.
4. Spector AA. FA metabolism in tumors. *Prog Biochem Pharmacol* 1975;10:42-75.
5. Bougnoux P. n-3 polyunsaturated fatty acids and cancer. *Curr Opin Clin Nutr Metab Care* 1999;2:121-6.
6. Woutersen RA, Appel MJ, van Garderen-Hoetmer A, Wijnands MV. Dietary fat and carcinogenesis. *Mutat Res* 1999;443:111-27.
7. Cave WT. Dietary n-3 polyunsaturated fatty acid effects on animal tumorigenesis. *FASEB J* 1991;5:2160-6.
8. Lands WEM. Biochemistry and physiology of 3 fatty acids. *FASEB J* 1992;6:2536-9.
9. Rudden RW. *Cancer biology*. 3rd ed. Oxford University Press: 1995.
10. Folch J, Lees M, Sloane-Stanley GH. A sample method for the isolation and purification of total lipids from animal tissues. *J Biol Chem* 1957;249:7-509.
11. Lepage, Roy CC. Direct transesterification of all classes of lipids in the one-step reaction. *J Lipid Res* 1986;27:114-20.
12. Anti M, Armelao F, Marra G, Percesepe A, Bartoli GM, Palozza P, et al. Effect of different doses of fish oil on rectal cell proliferation in patients with sporadic colonic adenomas. *Gastroenterology* 1995;107:1707-8.
13. Erickson KL, Hubbard NE. Dietary fat and tumor metastasis. *Nutr Rev* 1990;48:6-14.
14. Anti M, Marra G, Armelao F, Bartoli GM, Ficarella R, Percesepe A, et al. Effect of 3 fatty acids on rectal mucosal cell proliferation in subjects at risk for colon cancer. *Gastroenterology* 1992;103:883-91.
15. Huang YC, Jessup JM, Forse RA, Flicker S, Pleskow D, Anastopoulos HT, et al. N-3 fatty acids decrease colonic epithelial cell proliferation in high-risk bowel mucosa. *Lipids* 1996;31 Suppl:313-7.
16. Berdanier CD. Fatty acids and membrane function. In: Chow, CK, editor. *Fatty acids in foods and their health implications*. 2nd ed. New York: Marcel Dekker, Inc.; 2000.
17. Bockman R, Bellin A, Hickok N. Disordered prostaglandin production and cell differentiation/ proliferation in cancer. In: Thaler-Dao H, Crastes de Paulet A, Paoletti R, editors. *Eicosanoid and cancer*. New York: Raven Press; 1996. p.169-72.
18. Dantew B, Spagnuolo PJ. Tumor cell-endothelial cell interactions: evidence for roles for lipoxygenase products of arachidonic acid in metastasis. *Prostaglandins Leukot Essent Fatty Acids* 1997;56:295-300.
19. Shearnan MS, Shinomura T, Oda T, Nishizuka Y. Protein kinase C subspecies in adult rat hippocampal synaptosomes. Activation by diacylglycerol and arachidonic acid. *FEBS Lett* 1991;279:261-4.
20. Azzi A, Boscoboinik D, Hensey C. The protein kinase C family. *Eur J Biochem* 1992;208:547-57.
21. Nishizuka Y. The role of protein kinase C in cell surface signal transduction and tumor promotion. *Nature* 1984;308:693-8.
22. Bhathena SJ. Dietary fatty acids and fatty acid metabolism in diabetes. In: Chow CK, editor. *Fatty acid in foods and their health implications*. 2nd ed. New York: Marcel Dekker, Inc.; 2000.
23. Michalak J, Kadziolka A, Pruszkowska R, Ledwozyw A, Madejczyk A. Compensatory mechanisms in erythrocyte lipids in patients with atherosclerosis. *Lipids* 1988;23:476-80.