

Fine Structural Changes and Autoradiographic Studies of Rat Liver Cells Induced by Aflatoxin B₁ and G₁

Chung Sook Kim(Roe), Dong Sik Kim and Yoo Bock Lee

Department of Pathology, Yonsei University College of Medicine, Seoul, Korea

ABSTRACT

Ultrastructural changes were induced in rat liver by the single administration of aflatoxin B₁ and G₁.

These were assessed at intervals of 1, 6, 18, 24 and 48 hours and 1, 4 and 8 weeks.

It would appear that administration of aflatoxin B₁ induced a marked nucleolar alteration seen from 6 hours after the injection. No significant alterations were noted in animals treated with a small amount of aflatoxin G₁, but injection of a larger amount of aflatoxin G₁ induced nucleolar alterations similar to aflatoxin B₁ treatment. The nucleolar change was characterized by segregation of the granular and fibrillar elements.

All three groups showed cytoplasmic changes, such as dilation of rough endoplasmic reticulum with detachment of ribosomes, hyperplasia of smooth endoplasmic reticulum, increased numbers of lipid droplets and microbodies.

Autoradiographic studies have shown the absorption of both aflatoxin B₁ and G₁ in parenchymal cells, but aflatoxin B₁ was absorbed more intensely than G₁, especially in the nucleus. Autoradiographic findings in electron mic-

roscopy showed a marked difference in the size grains between aflatoxin B₁ and G₁, aflatoxin G₁ being larger than aflatoxin B₁.

INTRODUCTION

The aflatoxins are metabolites of certain strains of *aspergillus flavus* that grow on ground nuts and on some other foodstuffs (Sakai and Uruguchi, 1955; Borcker, 1966; Hartley et al., 1963; Barnes, 1967; Smith and Mckernan, 1962), and have proved to be toxic to several animals (Newberne et al., 1964). Allcroft and Carnaghan (1962), and Asao et al. (1963) have purified the crude aflatoxins B₁, B₂, G₁ and G₂ and found that their purified compounds are closely related to each other in terms of their structures and toxic effects on the cell metabolism of several animals.

It has been demonstrated that administration of aflatoxins to a variety of animals results in an inhibition of protein synthesis due to the effect on the RNA dependent polymerase, which takes place rapidly in a matter of a few hours (Clifford et al., 1967; Rogers and Newberne, 1967). Although B₁ and G₁ have structural similarity with each other, namely the only difference between them is that the dihydrofuran ring of the former is replaced by a

lactone ring in G_1 , it was reported that the LD_{50} of G_1 was three times higher than that of B_1 to 1 day old ducklings (Asao et al., 1963). However, G_1 is also carcinogenic when administered to the rats. (Wogan, 1968 ; Butler et al., 1969). This is also true as regards to a necrotizing does of aflatoxin in the case of a human liver cell culture system (Zuckerman et al., 1967).

The purpose of the present investigations to determine the toxicity of aflatoxin G_1 versus B_1 by comparing the ultrastructural changes in the rat liver and also by comparing the degree of cellular incorporation of B_1 and aflatoxin G_1 in liver cells as determined by autoradiographic techniques.

MATERIALS and METHODS

Male rats weighing about 200—220 gm were used for the experiment. Aflatoxin B_1 and G_1 were dissolved in Dimethylformamide (DMF) and tritium labelling was achieved by the method of Dr. W. Lizinsky(1966).

Animals were sacrificed at time intervals of 1 hour, 6 hours, 18 hours, 24 hours, 7days, 4 weeks and 8 weeks after a single administration of the aflatoxin by subcutaneous injection.

For Light and Electron Microscopy

The tissues were fixed in neutral buffered formalin and sections were prepared by the standard paraffin embedding procedure and stained routinely with Hematoxylin and eosin, Periodic acid Schiff(PAS), Methyl green pyronin and Feulgen reaction were also applied.

For electron microscopy, the animals were killed by blow on the head bled immediately by severing the jugular vein.

Specimens of the median lobe and fixed for 2 hours at 4C' in one percent osmium tetroxide in veronal buffer with pH of 7.4 (Palade 1952). All tissues were dehydrated in graded alcohol and embedded in Epon 812 according to standard procedures (Luft, 1961). Thin sections were cut with a LKB Ultratome then stained with uranyl acetate (Watson, 1958) and lead citrate (Millonig, 1961), and examined with the Hitachi HU 11—E electron microscope.

Autoradiographic Studies by Light Microscopy

The tissue was embedded in paraffin by standard procedures, and cut in 2-4 μ thickness. The sections were deparaffinized by immersing in xylene, and rinsed with distilled water. The slides were then coated with a nuclear emuls-

Experimental schedule

Group	No. of animals	Compounds given	Dose
Group I	21	Dimethylformamide	0.15ml DMF containing H^3 650 μ c.
Group II	21	Aflatoxin B_1	0.87mg AFT- B_1 containing H^3 650 μ c and dissolved in 0.15ml DMF.
Group III	21	Aflatoxin G_1	0.87mg AFT- G_1 containing H^3 650 μ c and dissolved in 0.15 ml DMF.
Group IV	21	Aflatoxin G_1	2.61mg AFT- G_1 containing H^3 650 μ c and dissolved in 0.15ml DMF.

ion, Kodak NTB-2, and stored in a refrigerator at 4°C for 30 days. The sections were developed in Kodak Dektol which was diluted 1:2 with water at 20°C, fixed in Kodac acid fixer, and finally washed in running water. After drying, the slides were stained by hematoxylin and eosin (Leblond, 1965) and examined under conventional light microscope.

Autoradiographic Studies by Electron Microscopy

Preparations were made by the method of Caro and Van Tubergen (1962). This involved direct application of a loop of L-4 nuclear sensitive emulsion to mounted sections. The exposure period was 100 days. They were developed by physical developer, followed by lead citrate staining. Routine observation was with electron microscope.

RESULT

Light Microscopic Findings:

In group 2 and 4, at 6 hours there was slight swelling of periportal parenchymal cells with increased acidophilia. By 18 to 24 hours there was slight necrosis of the periportal area with a neutrophil and mononuclear cell infiltrate. (Fig. 1, 2, 3). By 1 week, there was slight bile duct proliferation.

At these intervals, after stainings with Feulgen, Methyl green-pyronine reactions were much weaker than in the normal state. The results of these studies are shown in Table 1.

Electron Microscopic Findings:

Findings at 1 hour after the injection

In groups 2 and 4, the liver cells of periportal area were studied. The rough endoplasmic reticulum (RER) stacks showed dilation and irregular cisternae. Ribosomes were decreased and scattered through the cytoplasm either singly or as clusters. These changes were more

evident in group 2 than in other groups (Fig.5).

Findings at 6 and 18 hours after the injection

Nucleolar segregation was apparent in many of the nuclei in group 2 and 4. The nucleolus was compact with reduction of the granular components and fibrillar zones, but interchromatin granules were increased in number (Fig. 6, 7, 9).

Group 3 did not show any notable changes in the nucleolus at this time. The mitochondria showed swelling and exhibited irregular profiles and infolding of both inner and outer membranes, accompanied by increased dense granules and in some instances, membranous material was observed. The golgi membrane and intracisternal spaces showed dilation and increased dense particles (Fig. 10).

In group 4, there was more prominent irregular vesiculation of smooth endoplasmic reticulum (Fig. 8, 9).

The findings at 24 hours after injection

There were still a few segregations in the nucleolus in group 2 and 4. The rough endoplasmic reticulum stacks showed slight dilatation and detached ribosomes, and alteration of mitochondria could be observed in all groups (Fig. 7, 9, 10).

The findings at 4 and 8 weeks after injection

Recovery to normal conditions was almost complete except for slight changes in the cytoplasm, such as the irregular size of mitochondria and increased number of microbodies in group 4.

The ultrastructural changes in each group are summarized on Table 2.

Autoradiographic Findings:

Light microscopic findings

Group 2 (aflatoxin B₁) showed labelling in

Table 1. Light microscopic findings in the rat liver cells injection after aflatoxin B₁ and G₁

L.M. findings		Swelling	Vesiculation	Necrosis	Pyronin stain positive	Feulgen reaction positive	PAS stain positive	Bile duct proliferation
Group								
(Control) I	1 hour	—	—	—	‡‡	‡‡	‡‡	—
	6 "	—	—	—	‡‡	‡‡	‡‡	—
	18 "	—	—	—	‡‡	‡‡	‡‡	—
	24 "	—	—	—	‡‡	‡‡	‡‡	—
	1 week	—	—	—	‡‡	‡‡	‡‡	—
	4 "	—	—	—	‡‡	‡‡	‡‡	—
	8 "	—	—	—	‡‡	‡‡	‡‡	—
Group II (Aflatoxin B ₁) 0.87mg	1 hour	—	—	—	‡‡	‡‡	‡‡	—
	6 "	+	+	—	‡	‡‡	‡‡	—
	18 "	‡‡	+	+	‡	‡‡	‡‡	—
	24 "	‡	‡	‡	‡‡	‡‡	‡‡	—
	1 week	‡	‡	‡	‡‡	‡‡	‡‡	‡
	4 "	—	‡	—	‡‡	‡‡	‡‡	‡
	8 "	—	—	—	‡‡	‡‡	‡‡	‡
Group III Aflatoxin G ₁ 0.87mg	1 hour	—	—	—	‡‡	‡‡	‡‡	—
	6 "	—	—	—	‡‡	‡‡	‡‡	—
	18 "	‡	‡	—	‡‡	‡‡	‡‡	—
	24 "	‡	‡	‡	‡‡	‡‡	‡‡	—
	1 week	‡	‡	—	‡‡	‡‡	‡‡	—
	4 "	‡	—	—	‡‡	‡‡	‡‡	‡
	8 "	—	—	—	‡‡	‡‡	‡‡	—
Group IV Aflatoxin G ₁ 2.6mg	1 hour	—	—	—	‡‡	‡‡	‡‡	—
	6 "	‡	—	‡‡	‡	‡‡	‡‡	—
	18 "	‡	‡	‡‡	‡	‡‡	‡‡	—
	24 "	‡	‡‡	‡	‡‡	‡‡	‡‡	—
	1 week	‡	‡‡	‡	‡‡	‡‡	‡‡	‡
	4 "	—	‡‡	‡	‡‡	‡‡	‡‡	‡
	8 "	—	‡‡	—	‡‡	‡‡	‡‡	—

+ : Slight ‡‡ : Moderate ‡‡‡ : Severe

parenchymal cells first in the cytoplasm and later heavily in the nucleus (Fig. 12).

Group 3 (aflatoxin G₁ small dose) showed diffuse labelling in parenchymal cells and did not show localized labelling in the nucleus, but there was heavy labelling in mesenchymal cells (sinusoid, kupffer cells and endothelial cell of portal area) (Fig. 13).

Group 4 (aflatoxin G₁ high dose) showed labelling of similar degree as in Group 2 but it showed localized labelling in mesenchymal

cells (Fig. 14).

Electron microscopic findings

Labelling in the nucleus was noted at 6 hours in group 2 and group 4. There is a difference of the size of grain between group 2, 3 and 4 (aflatoxin B₁ and G₁) (Fig. 15, 16).

DISCUSSION

Since curd aflatoxin was purified into several

Table 2. Ultrastructure findings in the liver cells of the rat injection after aflatoxin B₁ and G₁

Group	E.M. finding	Nucleus		Cytoplasm							
		Membrane irregularity	Nuclear "cap"	RER	Ribosome	SER	Mitochondria		Lyso-	Micro-	Lipid
				dilation	detached	hyperplasia	swelling	irregularity	some increase	body increase	increase
Group I Control	1 hour	-	-	-	-	-	-	-	-	-	-
	6 "	-	-	-	-	-	-	-	-	-	-
	18 "	-	-	-	-	-	-	-	-	-	-
	24 "	-	-	-	-	-	-	-	-	-	-
	1 week	-	-	-	-	-	-	-	-	-	-
	4 "	-	-	-	-	-	-	-	-	-	-
	8 "	-	-	-	-	-	-	-	-	-	-
Group II Aflatoxin B ₁ 0.87mg	1 hour	+	-	+	++	+	+	-	+	-	-
	6 "	++	##	##	##	++	++	+	+	+	-
	18 "	++	++	##	##	++	+	+	+	+	++
	24 "	+	++	++	++	+	+	+	+	+	++
	1 week	+	+	+	-	+	-	-	-	-	+
	4 "	-	-	+	-	-	-	-	-	-	+
	8 "	-	-	-	-	-	-	-	-	-	+
Group III Aflatoxin G ₁ 0.87mg	1 hour	-	-	+	+	+	+	-	-	-	-
	6 "	-	-	+	+	+	++	++	-	-	-
	18 "	-	-	+	+	+	++	++	+	-	-
	24 "	-	-	+	+	-	+	+	+	+	+
	1 week	-	-	-	-	-	-	-	+	-	+
	4 "	-	-	-	-	-	-	-	-	-	+
	8 "	-	-	-	-	-	-	-	-	-	-
Group IV Aflatoxin G ₁ 2.61mg	1 hour	-	-	+	+	++	++	+	-	-	-
	6 "	-	++	+	++	##	++	##	-	-	-
	18 "	-	++	++	##	++	++	++	+	+	+
	24 "	-	+	+	+	+	+	++	+	+	+
	1 week	-	-	+	-	+	+	+	+	+	+
	4 "	-	-	-	-	-	+	+	-	+	-
	3 "	-	-	-	-	-	-	-	-	-	-

+ : Slight. ++ : Moderate. ## : Severe.
 RER (rough endoplasmic reticulum).
 SER (smooth endoplasmic reticulum).

Table 3. Percentage of nucleus in the liver cells of the rats at different dosages of H³-aflatoxin B₁ and G₁

Group	1 hour	6 hours	18 hours	24 hours	1 week	4 weeks	8 weeks
2	32.5%	38.5%	21.1%	13.6%	8.9%	6.7%	3.4%
3	14.1%	16.3%	10.1%	9.9%	6.0%	2.6%	2.0%
4	26.1%	31.5%	18.1%	16.1%	8.7%	6.9%	3.8%

Percentage of nucleus is expressed as number of labelled per 1,000 cells.

compounds, the one which was most extensively tested in terms of toxicity to the animal is aflatoxin B₁.

Although B₁ and G₁ have structural similarity, It was reported by Asao et al, (1963) that LD 50 of aflatoxin B₁ was three times higher than that of aflatoxin G₁ to 1 day old duckling.

It is also true as regards to the necrotizing dose of aflatoxin B₁ in the case of a human liver cell culture system reported by zuckerman (1967). Clifford and Rees (1967) and wogan (1966) compared the ability of interactions of B₁ and G₁ with DNA, and found that the action is similar but qualitatively may be different, because of a spectral shift indicating that the ability of interactions of aflatoxins B₁ and G₁ with DNA molecules is different in vitro.

These results coincide with the difference in toxicity of aflatoxins B₁ and G₁ in vitro experiments.

A consideration of the structure of aflatoxins B₁ and G₁, with their five rings and carbonyl groups capable of hydrogen binding, suggested that this might be competitive to DNA hydrogen binding itself. This would be expected if such as interaction of aflatoxin B₁ and G₁ occurred with DNA in vitro. B₁ is more competitive than G₁ as indicated by the evidence of spectral shift.

Our experiment shows that group 2 (aflatoxin B₁) revealed a marked nucleolar alteration after 6 hours up to 1 week.

Also group 4 (aflatoxin G₁ high dose) showed nucleolar segregation from 6 hours until 24 hours.

Ultrastructural lesions in liver cell nucleoli of rats treated with aflatoxin B₁ were described by Bernhard et al. (1965), who injected the compound into rats after partial hepatectomy.

These lesions consisted of segregation of the granular and fibrillar components with the formation of so-called nucleolar "caps."

Lafarge et al. (1966) described similar effects of the toxin in partially hepatectomized rats, and studied simultaneously the development of the nucleolar lesion and nuclear RNA synthesis.

Nucleolar ultrastructural changes were also observed by Svoboda et al. (1966), who performed electron microscopy on livers of rats and monkeys treated with aflatoxin B₁.

The various agents are known to cause nucleolar segregation. Actinomycin D produced this nucleolar change in the hepatocytes, as observed by Smuckler and Benditt (1955). Svoboda and Soga (1966) and Harris et al. (1968) reported anthromycin and pyrrolizidine alkaloids to affect nucleoli of hepatocytes.

The mechanism by which aflatoxin acts is unknown. However Freidman and Wogan (1967) demonstrated that large doses of the aflatoxin produce an inhibition of RNA polymerase activity.

Clifford et al. (1967) also reported marked RNA polymerase inhibition in livers of rats with twice the LD 50 dose of aflatoxin B₁. Clifford et al. (1967), using similar preparations, found that aflatoxin G₁ also inhibited RNA synthesis, but to a lesser extent than aflatoxin B₁. In conclusion, nucleolar segregation is the ultrastructural reflection of nucleolar dysfunction. That is, alteration in the synthesis of ribosomal precursors by various agents which bind DNA and RNA synthesis and protein synthesis may also be partly responsible for the ultrastructural changes observed.

Through this experiment, autoradiographic studies have shown the incorporation of both aflatoxin B₁ and G₁ on parenchymal cells, but aflatoxin B₁ was incorporated more intensely than G₁, especially in the nucleus. Autoradiog-

raphic findings with electron microscopy showed a marked difference in the size of grains between aflatoxin B₁ and G₁, namely G₁ being larger than B₁.

However, it is not certain that this difference in size of grains is related to the storage time of aflatoxin in liver cells.

In summary, the data obtained by the present experiments indicate that both aflatoxin B₁ and G₁ exert a toxic effect upon rat liver cells. However, the degree of toxic effect of aflatoxin B₁ is much stronger than that of G₁ as judged by nucleolar segregation, and capacity for nuclear incorporation.

REFERENCES

- Allcroft, R. and Carnaghan, R.B.A.: *Ground nut toxicity-Aspergillus flavus toxin (aflatoxin) in animal products*. *Vet. Record*, 74:863, 1962.
- Asao, T., Büchi, G., Abdel-kader, M.N., Chang, S. B., Wick, E.L. and Wogan, G.N.: *The structure of aflatoxin B and G*. *J. Am. Chem. Soc.*, 85:1706, 1963.
- Barnes, J.M. and Butler, W.H.: *Carcinogenic activity of aflatoxin to rats*. *Nature*, 202:1016, 1964.
- Barnes, J.M.: *Toxic fungi with special reference to aflatoxin*. *Trop. Sci.*, 9:64, 1967.
- Bernhard, W., Frayssinet, C., Lafarge, C. and Le-Breton, E.: *Lesions nucleolaires precoces provoquées par l'aflatoxine dans les cellules hépatiques du rat*. *Compt. Rend. Acad. Sci.*, 261:1785, 1965.
- Borker, E.: *Mycotoxins in feeds and foods*. *Advan. Appl. Microbiol.*, 8:315, 1966.
- Bowgeois, C.H., Shank, R.C., Grossman, R.A., Johnson, D.C. and Wooding, W.L.: *Acute aflatoxin B₁ toxicity in the Macaque and its similarities to Reye's Syndrome*. *Lab. Invest.*, 24:206, 1971.
- Büchi, G.: *The total synthesis of racemic aflatoxin B₁*. *Am. Chem. Soc.*, 89:6745, 1967.
- Butler, W.H., Greenblatt, M. and Lizinsky, W.: *Carcinogenesis in Rats by aflatoxins B₁, G₁ and B₂*. *Cancer Res.*, 29:2206, 1969.
- Butler, W.H. and Barnes, J.M.: *Toxic effects of ground nut meal containing aflatoxin to rats and guinea pigs*. *British. J. Cancer*, 17:699, 1963.
- Butler, W.H. and Barnes, J.M.: *Toxic effects of ground nut meal containing aflatoxin to rats and guinea pigs*. *British. J. Cancer*, 17:699, 1963.
- Butler, W.H.: *Early hepatic parenchymal changes induced in the rat by aflatoxin B₁*. *Am. J. Path.* 49:113, 1966.
- Carnaghan, R.B.A.: *Acute toxicity of aflatoxin*. *Brit. J. Cancer*, 21:811, 1967.
- Carnaghan, R.B.A.: *Toxicity and fluorescence properties of aflatoxins*. *Nature*, 200:1101, 1963.
- Caro, L.G. and Van Tubergen, R.P.: *Method for high resolution autoradiography*. *J. Cell Bio.*, 15:173, 1962.
- Chang, S.B., Abdel-kader, M.N., Wick, E.L. and Wogan, G.N.: *Aflatoxin B₂, chemical identity and biological activity*. *Science*, 142:1191, 1963.
- Clifford, J.I. and Rees, K.R.: *Aflatoxin; A site of action in the rat liver cell*. *Nature*, 206:312, 1966.
- Clifford, J., Rees, K.R. and Stevens, M.E.M.: *The effect of the aflatoxin B₁ and G₁ on protein and nucleic synthesis in rat liver*. *Biochem. J.*, 103:258, 1969.
- Cuthbertson, W.F.J., Laurson A.C. and Prat, D.A. H.: *Effect of ground nut meal containing aflatoxin on cynomolgus monkey*. *Brit J. Cancer*, 17:691, 1967.
- Dalton, A.J.: *Organized in benign and malignant cell*. *Lab. Invest.*, 8:510, 1959.
- Davidson, C.S.: *Plants and fungi as etiologic agents of cirrhosis*. *New Eng. J. Med.* 268:1072, 1963.
- Davison, C.S. and Alpert, M.E.: *Mycotoxins; Possible cause of primary carcinoma of the liver*. *Amer. J. Med.* 46:425, 1969.
- Dickens, F. and Jones, H.E.H.: *The carcinogenic action of aflatoxin after its subcutaneous injection in the rat*. *Brit. J. Cancer*, 17:691,

- 1963.
- Dutton, M.F. and Heathcote, J.G.: *The structure, biochemical properties and origin of the aflatoxin B_{2a} and G_{2a}*. *Chem. Ind. (London)*, 418, 1966.
- Emmelot, P. and Benedetti, E.L.: *Changes in the fine structure of rat liver cells brought by Dimethylnitrosamine*. *J. Cell Biol.*, 7:393, 1960.
- Friedman, M.A. and Wogan, G.N.: *Effects of aflatoxin B₁ on enzyme induction and nuclear RNA metabolism in the rat*. *Fed. Proc.*, 25:662, 1966.
- Harris, C., Grady, H. and Svoboda, D.: *Segregation of nucleolus produced anthromysin*. *Cancer Res.* 28:81, 1968.
- Hartley, R.D., Nesbitt, B.F. and O'Kelly, J.: *Toxic metabolites of Aspergillus flavus*. *Nature*, 198: 1956, 1963.
- Hesseltine, C.W., Shotwel, O.L., Ellis, J.J. and Stubblefield, R.D.: *Aflatoxin formation by Aspergillus flavus*. *Bact. Res.*, 30:795, 1966.
- Holzapfal, C.W., Steyn, P.S. and Purchase, I.F.H.: *Isolation and structure of aflatoxin M₁ and M₂*. *Tetrahedron Letters*, 25:2799, 1966.
- Jezequel, A.M and Bernhard, W.: *Modification ultra structurales du pancreas exocrine de rat sous l'effet de l' actinomycin D*. *J. Micro.*, 3:279, 1965.
- Kraybill, H.F. and Shimkin, M.B.: *Carcinogenesis related to foods contaminated by processing and fungal metabolites*. *Advan. Cancer Res.*, 8:191, 1964.
- Kulik, M.M. and Holaday, C.E.: *Aflatoxin metabolic product of several fungi*. *Mycopathol. Mycol. Appl.*, 30:137, 1967.
- Lancastar, M.C.: *Comparative aspects of aflatoxin induced hepatic tumors*. *Cancer Res.*, 28:2288, 1968.
- Lafarge, C., Frayssinet, C. and De Recondo, A.M.: *Inhibition preferentielle des synthesis de RNA nuclealaire provoquee par l' aflatoxin don less cellules hepatiques due rat*. *Compt. Rend. Acad. Sci.*, 263:1101, 1966.
- Lapis, K. and Bernhard, W.: *The effect of Mitomycin C on the nucleolar fine structure of KB cells in cell culture*. *Cancer Res.*, 25:628, 1965.
- Laqueur, G.L., Mickelsen, O., Whiting, M.G. and Kurland, L.T.: *Carcinogenic properties of nuts from cycas circinalis*. *J. Nat. Cancer. Inst.*, 31:915, 1963.
- Leblond, C.P.: *The use of Rodioautography in investigating protein synthesis*. *Academic press.*, 321:328, 1965.
- Lew, J., Kwon, S.P., Koho, C.M. and Chung, Y.: *Study on mycotoxin in Korean Fremented foodstuffs*. *J. Yonsei*, 7:191, 1969.
- Lizinsky, W. and Butler W.H.: *Purification and toxicity of aflatoxin G*. *Proc. Soc. Exp. Med.*, 123:151, 1966.
- Luft, J.H.: *Improvements in epoxy resin embedding method*. *J. Biophys. Cytol.*, 11:736, 1961.
- Madhaven, T.V., Tulpule P.G. and Goplan C.: *Aflatoxin Induced Hepatic fibrosis in rhasus monkeys*. *Arch. Path.*, 79:466, 1965.
- Magee, P.M. and Barnes, J.M.: *Induction of kidney tumors in the rat with. DMN*. *J Path. Bact.* 84:19, 1962.
- Millonig, G.A.: *Modified procedure for lead staining of thin sections*. *J. Biophys. Biochem. Cytol.*, 11:736, 1961.
- Nesbitt, B.F., O'Kelly, J., Sargeant, K. and Sheridan, A.: *Aspergillus flavus and turkey X-disease. Toxic metabolites of Aspergillus flavus*. *Nature*, 195:1062, 1962.
- Newberne, P.M., Carlton, W.W. and Wogan, G.N.: *Hepatomas in rats and hepatorenal injury in duckling fed peanut meal or Aspergillus flavus extract*. *Path. Vet.*, 1:105, 1964.
- Newberne, P.M. and Butler, W.H.: *Acute and chronic effects of aflatoxin on the liver of domestic and laboratory animals. A Review*. *Cancer Res.*, 29:236, 1969.
- Oettl's, A.G.: *Cancer in Africa especially in regions south of the Sahara*. *J. Nat. Cancer Inst.*, 33:333, 1964.
- Palade, G.E.: *A study of fixation for electron microscopy*. *J. Exp. Med.*, 95:285, 1952.

- Reich, E.: *Biochemistry of actinomycin D. Cancer Res.*, 23:1428, 1963.
- Reynolds, R.C., Montgomery, P.O.B. and Hoghes, B.: *Nuclear 'caps' produced by Actinomycin D. Cancer Res.*, 24:1269, 1964.
- Rey, R.D.R., Morgan, G. and Baral, J.: *Encephalopathy and fatty degeneration of the viscera; a disease entity in child food. Lancet* 2:749, 1963.
- Robertson, J.A., Pons, W.A. Jr. and Goldblatt, L. A.: *Preparation of aflatoxins and determination of their ultraviolet and fluorescent characteristics. Agr. Food Chem.*, 15:798, 1967.
- Rogers, A.E. and Newberne, P.M.: *The effects of Aflatoxin B₁ and Dimethylsulfoxide on thymidine H³ uptake and mitosis in rat liver. Cancer Res.*, 27:855, 1967.
- Sakai, F. and Uraguchi, K.: *Studies by long terms feeding by toxic substance from yellow rice. Nisshin Igaku.*, 42:609, 1955.
- Seakins, A. and Robinson, D.S.: *Changes associated with the production of fatty livers by phosphorus and by ethanol in the rat. Biochem. J.* 92:308, 1964.
- Shroeder, H.W. and Vellett M.J.: *Production of aflatoxin by Aspergillus wentii. Wehmer. Card. J. Microbiol.*, 15:895, 1969.
- Smuckler, E.A. and Benditt, E.P.: *The early effects of Actinomycin D on rat liver changes in ribosomes and polysomes. Lab. Invest.*, 14:1966, 1965.
- Smuckler, E.A., Iseri, O.A. and Benditt, E.P.: *An intracellular defect in protein synthesis induced carbon tetrachloride. J. Exp. Med.*, 116:55, 1962.
- Smith, R. and McKernan, W.: *Hepatotoxic action of chromatographically separated fractions of Aspergillus flavus extracts. Nature*, 195:1301, 1962.
- Sporn, M.B., Dingman, C.W., Phelps, H.L. and Wogan, G.N.: *Aflatoxin B₁. Binding to DNA in vitro and alteration of RNA metabolism in vivo. Science*, 151:1539, 1966.
- Svoboda, D., Grady, H.J. and Higginson, J.: *Aflatoxin B₁ injury in rat and monkey liver, Amer. J. Path.* 49:1023, 1966.
- Svoboda, D. and Soga, J.: *Early effects of pyrrolizidine alkaloids on the fine structure of rat liver cells. Am. J. Path.*, 48:347, 1968.
- Theron, J.J.: *Active liver injury in ducklings as result of aflatoxin poisoning. Lab. Invest.*, 14:1586, 1965.
- Trotter, N.L.: *A fine structure study of lipid in mouse liver regeneration after hepatectomy. J. Cell. Biol.*, 21:233, 1964.
- Tulpule, P.G., Madhavan, T.V. and Gopalan, C.: *Effect of feeding aflatoxin to young monkeys. Lancet*, 1:962, 1964.
- Watson, M.L.: *Staining of tissue sections for electron microscopy with heavy metals. J. Biophys. Biochem. Cytol.*, 4:475, 1958.
- Wogan, G.N.: *Biochemical Responses to aflatoxin. Cancer Res.*, 28:2282, 1968.
- Wogan, G.N.: *Chemical nature and biological effects of the aflatoxin. Bacteriol. Rev.*, 30:460, 1966.
- Wogan, G.N. and Newberne, P.M.: *Dose-Response characteristics of aflatoxin B₁ carcinogenesis in the rat. Cancer Res.*, 27:2379, 1967.
- Wood, R.L.: *The fine structure of hepatic cells in chronic ethionin poisoning and during recovery. Amer. J. path.* 46:307, 1965.
- Zuckerman, A.J., Ttsiquaye, K.N. and Futon, F.: *Tissue culture of human embryo liver cells and the cytotoxicity of aflatoxin B₁. Brit. J. Exp. Path.*, 48:20, 1967.

Legends of Figures

- Fig. 1.** Rat liver, 6 hours after the injection DMF (Control group). Hematoxylin and eosin stain. $\times 430$.
- Fig. 2.** Rat liver, 6 hours after the injection aflatoxin B_1 . Necrosis of contiguous cells is evident along with cell debris. Hematoxylin and eosin stain. $\times 430$.
- Fig. 3.** Rat liver, 24 hours after the injection aflatoxin G_1 . Numerous vesicles seen in hepatocytes. $\times 430$.
- Fig. 4.** Hepatic parenchymal cell, rat killed 6 hours after the injection DMF (control group).
The stacks of rough endoplasmic reticulum (rer) with ribosome (ri) and mitochondria (mi) showing normal arrangement. $\times 16,000$.
- Fig. 5.** Part of nucleus (Nu) and cytoplasm of rat liver cells, 6 hours after the injection aflatoxin B_1 .
Irregular shape of nucleus (Nu) and mitochondria (mi), increased number of lysosome (ly) and slightly dilation of rough endoplasmic reticulum stacks with detached ribosome. $\times 16,000$.
- Fig. 6.** Part of Nucleus (Nu), 6 hours after the injection aflatoxin B_1 .
The nucleolus fragmentation and segregation of granular and fibrillar components. $\times 20,000$.
- Fig. 7.** Part of Nucleus (Nu) and Cytoplasm of rat liver cells, 24 hours after the injection of aflatoxin G_1 (group 3).
The fibrillar components (f) and the granular components (g) show complete segregation, and there is marked proliferation of smooth endoplasmic reticulum. $\times 30,000$.
- Fig. 8.** Part of Cytoplasm of rat liver cells, 18 hours after the injection aflatoxin B_1 .
The most consistent change is dilation of the cisterns with detached ribosome (arrow), and increased number of lipid droplets. $\times 30,000$.
- Fig. 9.** Part of Nucleus (Nu) and Cytoplasm of rat liver cells 6 hours after the injection aflatoxin G_1 (group 4).
The nucleolar "caps" formed with dissociation of fibrillar (f) and granular components (g). There is marked proliferation of smooth endoplasmic reticulum. $\times 8,900$.
- Fig. 10.** Part of Cytoplasm of rat liver cells, 18 hours after the injection aflatoxin G_1 .
Showing irregular shape of mitochondria (mi) and contained membranous material and vesicular formation of smooth endoplasmic reticulum. $\times 18,000$.
- Fig. 11.** Radioautograph of rat liver cells at the control group. Paraffin section. H-E stain. $\times 1,800$.
- Fig. 12.** Radioautograph of rat liver cells at the 6 hours after given aflatoxin B_1 . Paraffin section. H-E stain.
Silver grains are heavily located over nucleus. $\times 1,800$.
- Fig. 13.** Radioautograph of rat 6 hours given aflatoxin G_1 . Paraffin section. H-E stain.
There is radioactivity in the nucleus and cytoplasm. $\times 1,800$.
- Fig. 14.** Radioautograph of rat liver cells at the 6 hours given aflatoxin G_1 (group 4). Paraffin section. H-E stain.
Silver grains are located over nucleus $\times 1,200$.
- Fig. 15.** Electron microscope radioautograph of rat liver cell 6 hours after administration of aflatoxin B_1 .
Numerous silver grains seen on the nucleus area. $\times 30,000$.
- Fig. 16.** Electron microscope radioautograph of liver rat cell 6 hours after administration of aflatoxin G_1 .
Few large silver grains located nucleus and the cytoplasm $\times 24,000$.













