



Estrogenic Activity of Sanguiin H-6 through Activation of Estrogen Receptor α Coactivator-binding Site

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Abstract – A popular approach for the study of estrogen receptor α inhibition is to investigate the protein-protein interaction between the estrogen receptor (ER) and the coactivator surface. In our study, we investigated phytochemicals from *Rubus coreanus* that were able to disrupt ER α and coactivator interaction with an ER α antagonist. The E-screen assay and molecular docking analysis were performed to evaluate the effects of the estrogenic activity of *R. coreanus* extract and its constituents on the MCF-7 human breast cancer cell line. At 100 μ g/mL, *R. coreanus* extract significantly stimulated cell proliferation ($574.57 \pm 8.56\%$). Sanguiin H6, which was isolated from *R. coreanus*, demonstrated the strongest affinity for the ER α coactivator-binding site in molecular docking analysis, with a binding energy of -250.149. The initial results of the study indicated that sanguiin H6 contributed to the estrogenic activity of *R. coreanus* through the activation of the ER α coactivator-binding site.

Keywords – *Rubus coreanus*, Sanguiin H-6, Estrogen receptor α , Coactivator-binding site, MCF-7 human breast cancer cells.

Introduction

In the last decade, cancer was the leading cause of death in Korea; one in four deaths resulted from the disease. Among Korean women, breast cancer was the second common cancer: cancer statistics from 2013 reported the diagnosis of 17,231 new cases, the occurrence of 2,231 deaths, and 146,416 established cases.¹ Breast cancer was also predicted to be the most common cancer in Korean women in 2017, with 21,544 estimated new cases and 2,519 estimated deaths.²

Owing to the development of science and technology, several new therapies for the treatment of breast cancer have evolved, such as oncoplastic surgery,³ bone-directed

treatments,⁴ and targeted therapy drugs.⁵ In addition to these novel remedial treatments, typical therapies such as tumor excision, adjuvant chemotherapy, or endocrine therapy have also been constantly improved. Estrogen antagonists play an important role in the hormonal manipulation used for treatment of breast cancer.⁶ Tamoxifen has been the most effective drug for breast cancer with estrogen-related tumors for over 30 years.⁷ However, tamoxifen interacts with estrogen receptors other than in the breast cancer cells, which leads to side effects in other tissues.⁸ Screening for new estrogenic antagonists is considered the optimal solution to enhance the efficacy of treatment.

Chemical constituents from herbal plants that exert anticancer effects are expected to provide a potential source of estrogenic antagonists for breast cancer treatment. Numerous published studies have described the anticancer effects of compounds isolated from medicinal plants, such as curcumin from turmeric, thymoquinone from black cumin, 6-shogaol from ginger, diallyl sulfide from garlic, piperine from black pepper, capsaicin from red chili

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pepper, and eugenol from clove.⁹ In spite of these compounds, few researchers have compared the mechanism of the anticancer effect exerted via estrogenic activities with other mechanisms.

Rubus coreanus Miquel, also known as Korean black raspberry or bokbunja, belongs to the *Idaeobatus* subgenus of the *Rubus* genus. *R. coreanus* is a native plant of East Asian countries, including China, Japan, and Korea.¹⁰ Bokbunja is used not only as Korean traditional beverage, but also a medicinal plant for the treatment of asthma, involuntary urination, and impotence.¹¹ The main chemical constituents of *R. coreanus* are anthocyanins, polyphenols, triterpenoid glycosides, and tannins.¹² These compounds are reported to possess anti-inflammatory¹³ and anti-cancer activity.¹⁴

In our study, MCF-7 human breast cancer cells were used in the E-screen assay as an *in vitro* model to determine the stimulatory effects of *R. coreanus* on cell proliferation. The ER α estrogenic activities of four compounds isolated from *R. coreanus* (3,4-dihydroxybenzoic acid, cyanidin 3-*O*-glucoside, nigaichigoside F1, and sanguiin H6) were evaluated by a molecular docking assay of the ER α ligand-binding domain. These 4 components were selected as a representative chemical component of anthocyanins (sanguiin H6), polyphenols (3,4-dihydroxybenzoic acid), triterpenoid glycosides (nigaichigoside F1), and tannins (cyanidin 3-*O*-glucoside) to find estrogenic chemical structures from various components *R. coreanus*.¹⁰⁻¹⁴ The results indicated that among the tested compounds, sanguiin H6 showed the strongest effect on the estrogenic activity of *R. coreanus* through the activation of the coactivator-binding site of ER α .

Experimental

Extraction and isolation – The dried fruits of *Rubus coreanus* Miquel (Korean black raspberry) were purchased from Kyoungdong Herbal Market in Seoul, in May 2016. The plant material was identified by one of the authors, Ki Hyun Kim, and a voucher specimen (BBJ-2016) was deposited at the School of Pharmacy Herbarium, Sungkyunkwan University (Suwon, Korea). The dried fruits of *R. coreanus* (500 g) were percolated three times with 80% methanol (2 L, for 3 days each time) at room temperature and filtered. The filtrate was evaporated under reduced pressure to produce a crude extract (104.5 g). Sanguiin H-6 was isolated from *R. coreanus* by using Sephadex LH-20 column chromatography and preparative HPLC. Briefly, 50 mL of aqueous red raspberry extract was loaded onto a Sephadex LH-20 column and polyphenols such as antho-

cyanins were eluted with 500 mL methanol:water (30:70, v/v). The fraction containing sanguiin H6 was eluted with 500 mL acetone:water (70:30, v/v). The sanguiin H6-rich fraction was further purified by preparative HPLC.

Cell culture – The estrogen receptor-positive MCF-7 human breast adenocarcinoma cell line was purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were grown in RPMI 1640 medium (Catalog number: MT10041CV; Corning, Manassas, VA, USA) supplemented with 100 units/mL penicillin, 100 μ g/mL streptomycin (Catalog number: 15140122; Gibco, Carlsbad, CA, USA), and 10% fetal bovine serum (Catalog number: FP-0500-A; Atlas Biologicals, Fort Collins, CO, USA) and incubated at 37 °C in a humidified atmosphere with 5% CO₂.

E-Screen assay – The E-screen assay for estrogenic effects on cell growth and specific protein synthesis was performed in the MCF-7 human breast adenocarcinoma cell line because of its estrogen sensitivity. The cells were incubated in 24-well culture plates at concentration of 20,000 cells/well in RPMI 1640 medium supplemented with 100 units/mL penicillin, 100 μ g/mL streptomycin, and 10% fetal bovine serum for 24 hours. The cells were treated with several concentrations of test materials (5 – 100 μ g/mL for *R. coreanus* extract and 25 – 100 μ M for sanguiin H-6) in phenol red-free RPMI (Catalog number: 11835030; Gibco, Carlsbad, CA, USA) supplemented with 5% charcoal-dextran-stripped human serum (Catalog number: IPLA-SER-CS; Innovative Research, Novi, USA) for 144 hours. For the antagonistic test, the pure estrogen receptor antagonist ICI 182 780 (Catalog number: 1047; Tocris Bioscience, Bristol, UK) was added with the test materials. Estrogenic activity from serum was minimized by the use of charcoal-dextran-stripped human serum. Ez-Cytox reagent (Catalog number: EZ-3000; Daeil Lab Service, Seoul, South Korea) was added to each well for 1 h and the cell viability was then calculated from the measurement of the optical density at 450 nm using a microplate reader (PowerWave XS; Bio-Tek Instruments, Winooski, VT, USA).^{15,16}

Molecular docking assay – The estrogenic effects of the compounds isolated from *R. coreanus* on ER α were evaluated by molecular docking assay of the ER α ligand-binding domain. Initially, the molecular model was constructed from the X-ray structure of ER α complexed with estradiol and a synthesized stable peptide inhibitor in the coactivator-binding groove. The atomic coordinates were obtained from the Protein Data Bank (PDB Code: 5dxb) and manipulated using Discovery Studio molecular modeling package. Chain B and all water molecules of

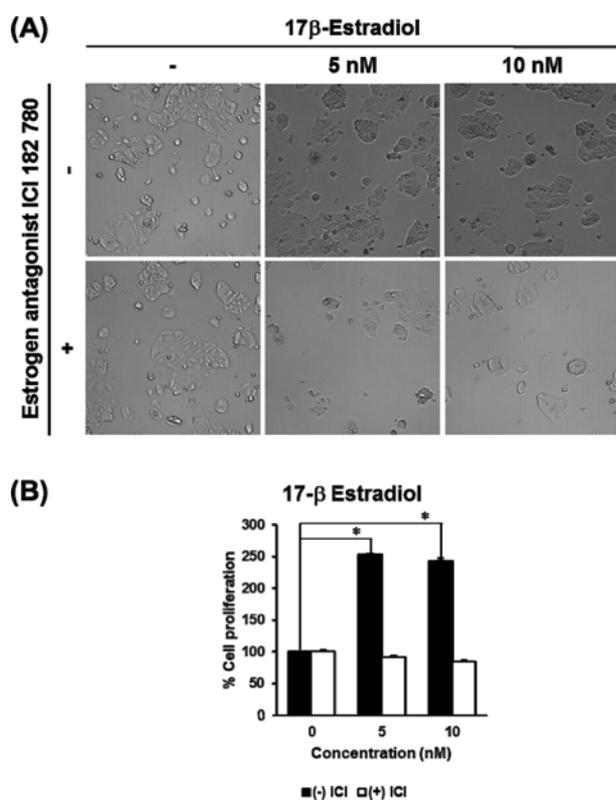


Fig. 1. The estrogenic effect of 17β-estradiol on the proliferation of MCF-7 human breast cancer cells. (A) The microscopic pictures from E-screen assay results of 17β-estradiol. (B) The comparative graph illustrates the percentage increase in cell proliferation compared with the untreated group. In this E-screen assay, 17β-estradiol was used as a positive control. MCF-7 cells were incubated in 24-well plates and treated with tested sample in phenol red-free RPMI medium supplemented with 5% charcoal-dextran-stripped human for 144 hours. For the antagonistic test, the pure estrogen receptor antagonist ICI 182 780 was added with the test materials. Ez-Cytox reagent was added to each well for 1 hour and the cell viability was then calculated from the measurement of the optical density at 450 nm using a microplate reader. P values of less than 0.001 were considered statistically significant.

chain A were deleted. To identify the docking structures of *R. coreanus* compounds that fitted into the coactivator-binding site of ERα, the docking simulation was implemented by CDocker program. All atomic charges of ligands were assigned using the Momany-Rone partial charge and CHARMM force fields.¹⁷⁻¹⁹

Statistical analysis – All data in our study were presented as mean ± standard deviation (SD). Statistical significance was determined by the one-way analysis of variance (ANOVA). P values of less than 0.001 were considered statistically significant.

Result and Discussion

The estrogenic activity of *R. coreanus* extract was

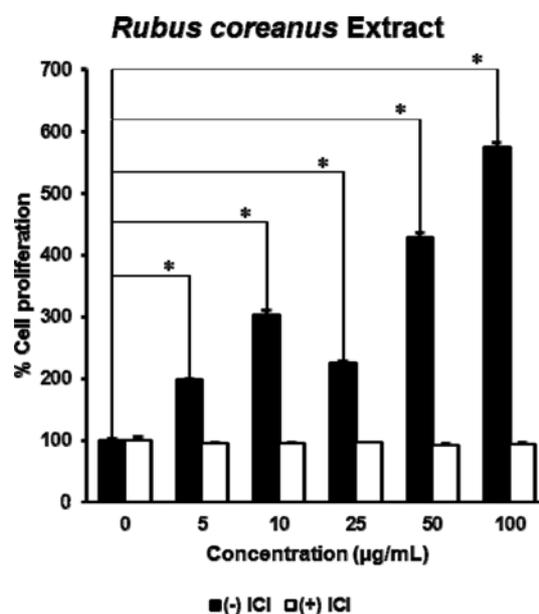


Fig. 2. The estrogenic effect of *Rubus coreanus* extract on the proliferation of MCF-7 cells. The extract of *R. coreanus* significantly stimulated cell growth in a dose-dependent manner, especially at the concentration of 100 μg/mL. P values of less than 0.001 were considered statistically significant.

evaluated through the measurement of the proliferation of MCF-7 human breast adenocarcinoma cells in response to estrogen, using the E-Screen assay. In Fig. 1, the estrogen-responsive cell growth was determined by treatment with the positive control, 17β-estradiol, at 5 and 10 nM. Whereas, pre-treatment with ICI 182 780, a steroidal estrogen antagonist, completely prevented 17β-estradiol-induced the proliferation of MCF-7 cells. The extract of *R. coreanus* was found to significantly stimulate cell growth in a dose-dependent manner (Fig. 2). As shown in Fig. 2, the proliferation of MCF-7 cells increased markedly after treatment with 100 μg/mL *R. coreanus* extract (574.57 ± 8.56%). Additionally, the proliferative effect of *R. coreanus* extract was completely prevented, in the same way as that of 17β-estradiol, by the presence of ICI 182 780.

The E-screen assay showed that *R. coreanus* extract exerted estrogenic activity that induced MCF-7 breast cancer cell proliferation. Thus, the active compounds from *R. coreanus* are expected to be regulators of ERα transcription activity. A molecular docking assay of the ERα ligand-binding domain was performed to analyze the estrogenic effect of *R. coreanus* compounds on ERα. Based on a literature search, four representative isolated compounds (3,4-dihydroxybenzoic acid, cyanidin 3-O-glucoside, nigaichigoside F1, and sanguin H6) were tested. However, the 4 compounds we selected have low

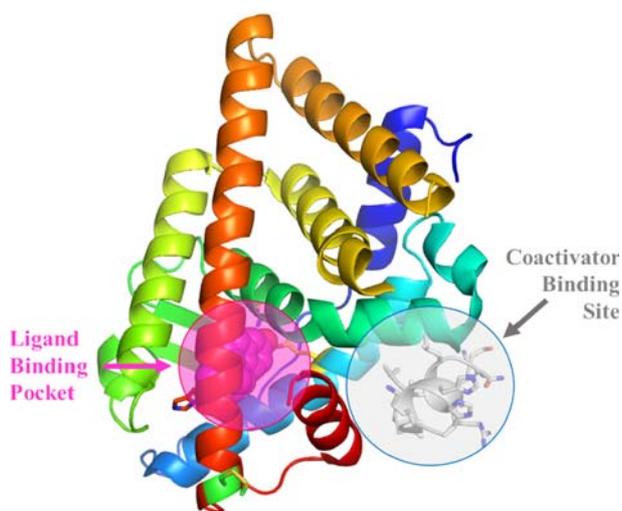


Fig. 3. Structure of ligand-binding domain of estrogen receptor α with the ligand-binding pocket and coactivator-binding site. Not only the binding between estrogen and its ligand binding pocket, the protein-protein interactions between coactivator proteins and their binding sites also results in cell proliferation. Competitive blockade of the coactivator binding site could effectively halt cell proliferation for breast cancer therapeutics.

binding energy or high molecular weight to ER α . We then examined the possible action of the components on ER α by the ER α coactivator. The binding energies of these compounds with the ER α coactivator pocket were calculated. The values for cyanidin-3-O-glucoside, 3,4-dihydroxybenzoic acid, nigaichigoside F1, and sanguin H6 were -39.0127, -29.9156, -6.6683, and -250.149, respectively. The key interaction features between *R. coreanus* compounds and the coactivator pocket are the hydrophobic interactions with Val, Ile, Leu, and Met (Fig. 4), which were general protein-protein interaction related residues. Sanguin H6 showed the strongest affinity for the ER α coactivator-binding site and was selected for the evaluation of the effect on the proliferation of MCF-7 human breast cancer cells.

The E-screen assay was conducted with a dose-response study to determine the effects of *R. coreanus* compounds on MCF-7 cell proliferation (Fig. 5). The results indicated that sanguin H6 and cyanidin-3-O-glucoside had an effect on the proliferation of MCF-7 cells only at 100 μ M, the highest test concentration ($127.41 \pm 0.26\%$). These compounds also had the stronger affinity with the ER α coactivator-binding site than the other compounds. Therefore, we could not identify an active ingredient that explains the effects of *R. coreanus* extract. Further research on other ingredients or synergies action of components is needed.

The binding between estrogen and the ligand-binding

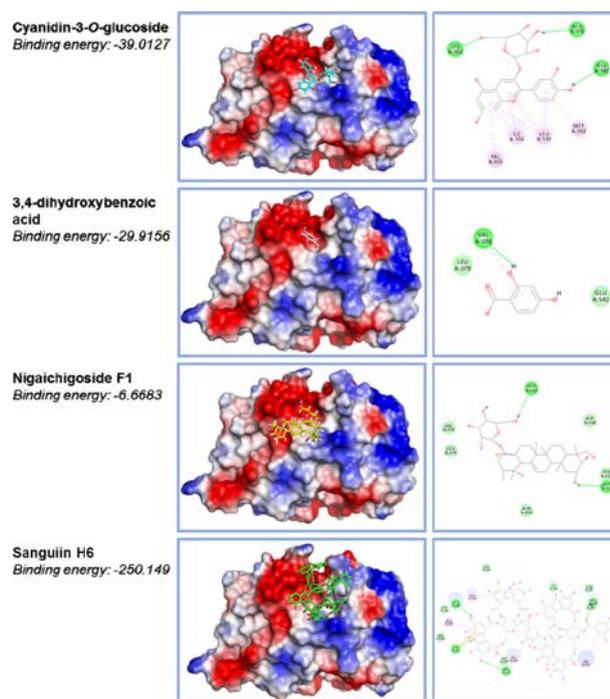


Fig. 4. The docking structures of *R. coreanus* compounds with coactivator-binding site of estrogen receptor α . Molecular docking assay was constructed from the X-ray structure of ER α complexed with estradiol and a synthesized stable peptide inhibitor in the coactivator-binding groove. The atomic coordinates were obtained from the Protein Data Bank and manipulated using Discovery Studio molecular modeling package. The docking simulation was implemented by CDocker program. All atomic charges of ligands were assigned using the Momany-Rone partial charge and CHARMM force fields. Among four compounds isolated from *R. coreanus*, Sanguin H6 showed the strongest affinity for the ER α coactivator-binding site.

domain (LBD) of the estrogen receptor (ER) initiates a biological cascade that ultimately results in cell proliferation. This binding induces a conformational change in the nuclear receptor (NR) to allow the binding of coactivator proteins, which thereby enables the further recruitment of the necessary proteins for gene transcription.²⁰ The structure of the ER α ligand-binding domain with the ligand-binding pocket and coactivator-binding site is shown in Fig. 3.

Breast cancer therapeutics were initially developed with a focus on antagonists that directly block the binding of estrogen to ER-LBD. However, many recent studies have targeted the regulation of ER function by blocking the protein-protein interactions between coactivator proteins and their binding sites.²¹ The coactivators interact with NR-LBD through a two-turn amphipathic α -helical motif that contains the conserved motif, LXXLL (where L is leucine and X represents any amino acid), which is known as nuclear receptor-interaction box (NR-box).²²

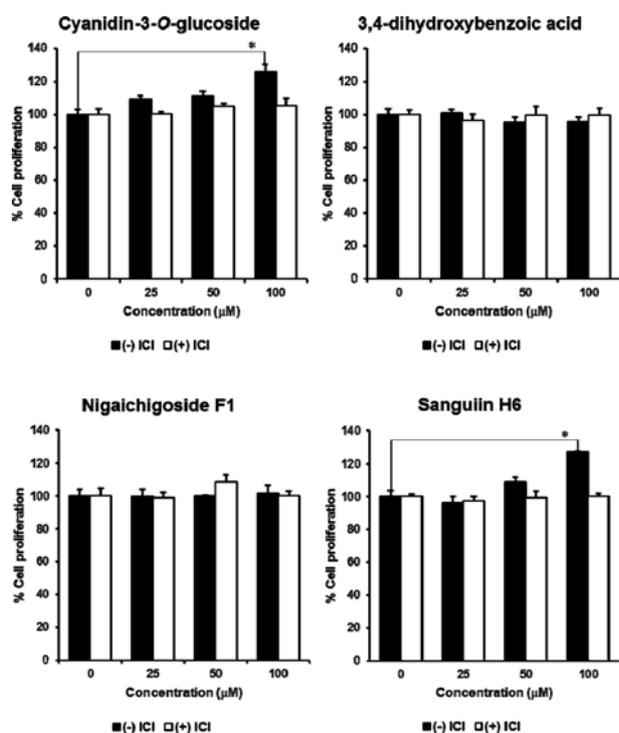


Fig. 5. The estrogenic effects of *R. coreanus* compounds on the proliferation of MCF-7 cells. Sanguin H6 and cyanidin-3-O-glucoside slightly induced the proliferation of MCF-7 cells at the concentration of 100 μ M. P values of less than 0.001 were considered statistically significant.

Competitive blockade of this binding site prevents recruitment of the transcription apparatus and could effectively halt cell proliferation.

E-screen assay was known as a common cell proliferation assay which allows investigating the total estrogenic activity via the increase in cell number of estrogen receptor-positive cells like MCF-7 human breast adenocarcinoma cell.¹⁶ Thus, E-screen assay was performed first to evaluate the estrogenic activity of *R. coreanus* extract. As the results, the MCF-7 cell proliferation was significantly stimulated while treated *R. coreanus* extract at the concentration of 100 μ g/mL. Many previous studies mentioned the pharmacological effects of *R. coreanus* as anti-cancer,²³ anti-metastatic,²⁴ and protective effect against oxidative stress.²⁵ However, to the best of our knowledge, this is one initial study about the estrogenic activity of *R. coreanus*.

Molecular docking assay was conducted to measure the binding affinity of a phytochemical from *R. coreanus* for the estrogen receptor. In this study, we focused on exploring the interaction between *R. coreanus* compounds and the coactivator binding site of estrogen receptor. Among these compounds, sanguin H6 had the most

negative interaction energy thus favorable to binding. The estrogenic activity of sanguin H6 was also investigated by E-screen assay, the result showed that sanguin H6 slightly stimulate the proliferation of MCF-7 cells at the highest test concentration of 100 μ M.

Sanguin H6 was considered as the potential anti-cancer candidate that suppressing the cell proliferation of several cancer cell lines such as A549,²⁶ MDA-MB-231, and even MCF-7.²³ There seems to be a contradiction in this present study results with the previous published data because we found that sanguin H6 slightly stimulate the proliferation of MCF-7 cells via E-screen assay. However, it should be noted that for E-screen assay, the proliferation of human estrogen-sensitive cells was initially inhibited by human serum-borne molecules, and subsequently the estrogen-like activity of natural compounds, for example, sanguin H6 in this case, stimulate the cell proliferation by eliminating this inhibitory effect.¹⁶ In comparison with the previous study of Park *et al.*, the cell viability assay was conducted by adding only sanguin H6 to check its cytotoxic effect on MCF-7 cells.²³ In addition, many references pointed out that phytoestrogens can have both of positive and negative effects on the proliferation of breast cancer cells depend on a lot of factors such as using amount, exposure time, or the capacity of absorption and metabolism each cell lines.²⁷ For instance, genistein is a well-known estrogen antagonist which enhanced the proliferation of MCF-7 cells by stimulating the expression of pS2 gene.²⁸ In spite of this, genistein also worked as an anti-cancer agent while involved in the regulation of several signalling pathways like NF- κ B, Akt, protein-tyrosine kinase (PTK), matrix metalloproteinases (MMPs), and Bax/Bcl-2.²⁹

In conclusion, the E-screen assay and molecular docking analysis were conducted to evaluate the effects of the estrogenic activity of *R. coreanus* extract and its constituents on the MCF-7 human breast cancer cell line. The extract of *R. coreanus* was found to significantly stimulate cell proliferation at 100 μ g/mL ($574.57 \pm 8.56\%$). Sanguin H6 showed the strongest affinity for the ER α coactivator-binding site with a binding energy of -250.149. However, only a small effect was seen on the viability of stimulated breast cancer cells at 100 μ M ($127.41 \pm 0.26\%$). Future studies will be conducted to confirm the molecular mechanism for the estrogenic activities of *R. coreanus*.

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References

- (1) Oh, C. M.; Won, Y. J.; Jung, K. W.; Kong, H. J.; Cho, H.; Lee, J. K.; Lee, D. H.; Lee, K. H. *Cancer Res. Treat.* **2016**, *48*, 436-450.
- (2) Jung, K. W.; Won, Y. J.; Oh, C. M.; Kong, H. J.; Lee, D. H.; Lee, K. H. *Cancer Res. Treat.* **2017**, *49*, 306-312.
- (3) Carmichael, A. R.; Mokbel, K. *Arch. Plast. Surg.* **2016**, *43*, 222-223.
- (4) Irelli, A.; Cocciolone, V.; Cannita, K.; Zugaro, L.; Di Staso, M.; Lanfiuti Baldi, P. L.; Paradisi, S.; Sidoni, T.; Ricevuto, E.; Ficorella, C. *Bone* **2016**, *87*, 169-175.
- (5) de Pedro, M.; Baeza, S.; Escudero, M. T.; Dierssen-Sotos, T.; Gómez-Acebo, I.; Pollán, M.; Llorca, J. *Breast Cancer Res. Treat.* **2015**, *149*, 525-536.
- (6) Esteva, F. J.; Hortobagyi, G. N. *Sci. Am.* **2008**, *298*, 58-65.
- (7) Jameera Begam, A.; Jubie, S.; Nanjan, M. J. *Bioorg. Chem.* **2017**, *71*, 257-274.
- (8) Howell, S. J.; Johnston, S. R. D.; Howell, A. *Best Pract. Res. Clin. Endocrinol. Metab.* **2004**, *18*, 47-66.
- (9) Zheng, J.; Zhou, Y.; Li, Y.; Xu, D. P.; Li, S.; Li, H. B. *Nutrients* **2016**, *8*, 495.
- (10) Lee, J.; Dossett, M.; Finn, C. E. *Molecules* **2014**, *19*, 10524-10533.
- (11) Heo, J. Donguibogam; Yeogang: Korea, **1994**, pp 946-947.
- (12) Li, J.; Du, L. F.; He, Y.; Yang, L.; Li, Y. Y.; Wang, Y. F.; Chai, X.; Zhu, Y.; Gao, X. M. *Chem. Biodivers.* **2015**, *12*, 1809-1847.
- (13) Ju, H. K.; Cho, E. J.; Jang, M. H.; Lee, Y. Y.; Hong, S. S.; Park, J. H.; Kwon, S. W. *J. Pharm. Biomed. Anal.* **2009**, *49*, 820-827.
- (14) Choung, M. G.; Lim, J. D. *Korean J. Med. Crop Sci.* **2012**, *20*, 259-269.
- (15) Körner, W.; Hanf, V.; Schuller, W.; Kempter, C.; Metzger, J.; Haagenmaier, H. *Sci. Total Environ.* **1999**, *225*, 33-48.
- (16) Soto, A. M.; Sonnenschein, C.; Chung, K. L.; Fernandez, M. F.; Olea, N.; Serrano, F. O. *Environ. Health Perspect.* **1995**, *103*, 113-122.
- (17) Lee, S.; Barron, M. G. *PloS One* **2017**, *12*, 1-14.
- (18) Ng, H. W.; Zhang, W.; Shu, M.; Luo, H.; Ge, W.; Perkins, R.; Tong, W.; Hong, H. *BMC Bioinformatics.* **2014**, *15*, 1-15.
- (19) Pang, X.; Fu, W.; Wang, J.; Kang, D.; Xu, L.; Zhao, Y.; Liu, A. L.; Du, G. H. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 1-11.
- (20) Jordan, V. C. *J. Med. Chem.* **2003**, *46*, 883-908.
- (21) McDonnell, D. P.; Chang, C. Y.; Norris, J. D. *J. Steroid Biochem. Mol. Biol.* **2000**, *74*, 327-335.
- (22) Sun, A.; Moore, T. W.; Gunther, J. R.; Kim, M. S.; Rhoden, E.; Du, Y.; Fu, H.; Snyder, J. P.; Katzenellenbogen, J. A. *Chem. Med. Chem.* **2011**, *6*, 654-666.
- (23) Park, E. J.; Lee, D.; Baek, S. E.; Kim, K. H.; Kang, K. S.; Jang, T. S.; Lee, H. L.; Song, J. H.; Yoo, J. E. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 4389-4392.
- (24) Park, E. H.; Park, J. Y.; Yoo, H. S.; Yoo, J. E.; Lee, H. L. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 3291-3294.
- (25) Choi, M. H.; Shim, S. M.; Kim, G. H. *J. Food Sci. Technol.* **2016**, *53*, 1214-1221.
- (26) Ko, H.; Jeon, H.; Lee, D.; Choi, H. K.; Kang, K. S.; Choi, K. C. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 5508-5513.
- (27) Helferich, W. G.; Andrade, J. E.; Hoagland, M. S. *Inflammopharmacology* **2008**, *16*, 219-226.
- (28) Hsieh, C. Y.; Santell, R. C.; Haslam, S. Z.; Helferich, W. G. *Cancer Res.* **1998**, *58*, 3833-3838.
- (29) Lee, J. Y.; Kim, H. S.; Song, Y. S. *J. Tradit. Complement. Med.* **2012**, *2*, 96-104.

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