



Chemicals from *Cimicifuga dahurica* and Their Inhibitory Effects on Pro-inflammatory Cytokine Production by LPS-stimulated Bone Marrow-derived Dendritic Cells

Nguyen Phuong Thao^{1,†}, Young Suk Lee^{2,†}, Bui Thi Thuy Luyen³, Ha Van Oanh³, Irshad Ali⁴, Madeeha Arooj⁴,
Young Sang Koh^{4,*}, Seo Young Yang^{2,*}, and Young Ho Kim^{2,*}

¹Institute of Marine Biochemistry (IMBC), Vietnam Academy of Science and Technology (VAST),
18-Hoang Quoc Viet, Hanoi, Vietnam

²College of Pharmacy, Chungnam National University, Daejeon 34134, Republic of Korea

³Hanoi University of Pharmacy, 13-15 Le Thanh Tong, Hanoi, Vietnam

⁴School of Medicine and Jeju Research Center for Natural Medicine, Jeju National University, Jeju 63243, Republic of Korea

Abstract – Inflammation is a biological response caused by overactivation of the immune system and is controlled by immune cells via a variety of cytokines. The overproduction of pro-inflammatory cytokines enhances abnormal host immunity, resulting in diseases such as rheumatoid arthritis, cardiovascular disease, Alzheimer's disease, and cancer. Inhibiting the production of pro-inflammatory cytokines such as interleukin (IL)-12p40, IL-6, and tumor necrosis factor (TNF)- α might be one way to treat these conditions. Here, we investigated the anti-inflammatory activity of compounds isolated from *Cimicifuga dahurica* (Turcz.) Maxim., which is traditionally used as an antipyretic and analgesic in Korea. In primary cell culture assays, 12 compounds were found to inhibit the production of pro-inflammatory cytokines (IL-12p40, IL-6, and TNF- α) *in vitro* in bone marrow-derived dendritic cells stimulated with LPS.

Keywords – *Cimicifuga dahurica*, Ranunculaceae, Anti-inflammatory, IL-12 p40, IL-6, TNF- α

Introduction

Inflammation is the sum of the complex biological responses caused by an overactivated immune system. Inflammation is closely associated with the pathology of many diseases, such as autoimmune diseases like multiple sclerosis and rheumatoid arthritis as well as degenerative diseases like Alzheimer's disease, cardiovascular disease, and cancer.¹ Consequently, it will be important to develop treatments that can alleviate or cure these diseases.

Dendritic cells (DCs) perform essential roles in innate

and adaptive immunity, such as processing and presenting xenobiotics intruded into the host and acting as a central regulator of the immune responses.² These functions of DCs are controlled by various pro-inflammatory cytokines, such as interleukin (IL)-12p40, IL-6, and tumor necrosis factor (TNF)- α .³

IL-12 has two subtypes, p35 and p40, which have disulfide bonds in different locations. It is produced by antigen-presenting cells, like DCs, monocytes, and macrophages, and induces an inflammatory response to xenobiotics and immune signals.⁴ It might also be a trigger that initiates cellular immunity. IL-6 was originally known as a B-cell differentiation factor and is involved in various biological actions, such as the acute immune response and hematopoiesis.⁵ TNF- α is mainly released by monocytes and macrophages and is a pro-inflammatory cytokine involved in host defenses against xenobiotics and cancer.⁶ TNF- α might also be involved in the pathogenic mechanisms of acute and chronic inflammatory disease.⁷

Pro-inflammatory cytokines play important roles in host immunity, but their overexpression can make a

*Author for correspondence

Young Ho Kim, Ph.D., College of Pharmacy, Chungnam National University, Daejeon 34134, Republic of Korea.
Tel: +82-42-821-5933; E-mail: yhk@cnu.ac.kr

Seo Young Yang Ph.D., College of Pharmacy, Chungnam National University, Daejeon 34134, Republic of Korea.
Tel: +82-42-821-7321; E-mail: syyang@cnu.ac.kr

Young Sang Koh Ph.D., School of Medicine and Jeju Research Center for Natural Medicine, Jeju National University, Jeju 63243, Republic of Korea.
Tel: +82-64-754-3851; E-mail: yskoh7@jejunu.ac.kr

[†]These authors contributed equally to this work.

disease worse, triggering fever and tissue damage and even leading to death. Suppressing the overproduction of these cytokines could be a way to treat diseases related to inflammation.

Cimicifuga dahurica (Turcz.) Maxim. belongs to the Ranunculaceae and grows widely in Korea, China, and Japan where it is traditionally used as an antipyretic and analgesic.⁸ This medicinal plant produces cycloartane-type triterpenoids, indolinone alkaloids, phenolics, and their glycosides. Plant preparations are believed to have anti-cancer, anti-inflammatory, and antioxidative effects.⁹⁻¹² Previously, we reported that components from *C. dahurica* inhibit soluble epoxide hydrolase, which is one of the causes of cardiovascular and renal disease.¹³⁻¹⁵ As part of our ongoing investigations of the anti-inflammatory activity of Korean medicinal plants, we found that the methanol extract of *C. dahurica* and dichloromethane and water fractions showed significant *in vitro* anti-inflammatory activity. Here, we studied its immunoregulatory effect by measuring excreted pro-inflammatory cytokines in the culture supernatant of bone marrow-derived dendritic cells (BMDCs) stimulated by lipopolysaccharide (LPS).

Experimental

Extraction and isolation – The fresh roots of *C. dahurica* (2.5 kg) were cut into pieces and extracted with 95% aqueous EtOH (3 × 5.0 L) under ultrasonic agitation at 90 Hz and 40°C. The ethanol solution was concentrated under vacuum and filtered through a Buchner funnel to give a brown extract (65.3 g), which was suspended in distilled H₂O and successively partitioned with *n*-hexane and CH₂Cl₂ to afford *n*-hexane (9.6 g, A), CH₂Cl₂ (15.2 g, B) fractions, and a H₂O layer (W).

The CH₂Cl₂ fraction was partitioned by column chromatography over silica gel and eluted with *n*-hexane-EtOAc (95:5, 80:20, 40:60, v/v) to produce seven fractions (Fractions, B-1 to B-7). Fraction B-3 (0.8 g) was chromatographed by Sephadex LH-20 CC eluted with MeOH-H₂O (95:5, 70:30, 50:50, v/v) to give three sub-fractions (Fractions, B-3.1 to B-3.3) and further purified by YMC RP-C₁₈ CC using acetone-H₂O (3:2) as the eluent to afford 24-*epi*-acerinol (**1**, 6.8 mg), 23-*O*-methylcimicerogenin B (**2**, 3.5 mg), and 25-triepoxy-12 β -acetoxy-3 β ,26-dihydroxy-9,19-cyclolanost-7-ene (**3**, 10.2 mg). Next, fraction B-4 (1.1 g) was chromatographed over a silica gel CC eluted with *n*-hexane-EtOAc (2:1) to obtain 7,8-didehydrocimigenol (**4**, 13.5 mg), 24-*epi*-24-*O*-acetyl-7,8-didehydroshengmanol (**5**, 4.0 mg), and 25-anhydrocimigenol (**6**, 3.1 mg). Similarly, fraction B-5 (1.3

g) was separated by YMC RP-C₁₈ CC using acetone-H₂O (2.5:1) and Sephadex LH-20 using CH₂Cl₂-MeOH (20:80) as eluent to give 25-*O*-acetyl-7,8-didehydrocimigenol (**7**, 5.6 mg). Next, fraction B-7 (1.1 g) was chromatographed on a column of Sephadex LH-20 and eluted with H₂O in MeOH, successively, to give three sub-fractions (Fractions, B-7.1 to B-7.3). Subfraction B-7.2 (0.5 g) was separated by CC over silica gel using a mixture solvent of CH₂Cl₂-MeOH-H₂O (8:1:0.08) as eluents and further purified by YMC RP-C₁₈ CC, eluted with MeOH-H₂O (4:1), to give ferulic acid methyl ester (832.6 mg).

The H₂O fraction was separated using a Diaion HP-20 column and was eluted with a gradient solvent mixture of MeOH-H₂O (25:75, 50:50, 65:35, 75:25, to pure MeOH, stepwise) to yield four fractions (W-1 to W-4), based on TLC analysis. When the same steps were repeated as above, 7,8-didehydro-25-anhydrocimigenol-3-*O*- β -D-xylopyranoside (**9**, 18.2 mg) was also obtained by purifying subfraction W-2 on YMC RP-C₁₈ silica gel and followed by passage over a Sephadex LH-20 column using mixtures of MeOH-acetone-H₂O (8:1:1). Next, fraction W-3 (1.7 g) was subjected to YMC RP-C₁₈ silica gel CC and was eluted with a solvent mixture of H₂O-MeOH (5:1, 3:1, 1:1, and 100% MeOH) to afford four smaller subfractions (W-3.1 to W-3.4). Further purification of subfraction W-3.1 (0.42 g) via silica gel column eluted with CH₂Cl₂-MeOH (7:1), yielded a compound mixture (18.2 mg) that was further purified by preparative TLC (acetone-H₂O, 1:2) to yield 24-*epi*-25-*O*-acetyl-7,8-didehydrocimigenol 3-*O*- β -D-xylopyranoside (**10**, 9.5 mg). In a similar process to that described above, subfraction W-3.3 (0.07 g) was subjected to YMC RP-C₁₈ silica gel CC (MeOH-H₂O, 1:1) and then was further purified by silica gel CC using to yield cimircaside C (**11**, 10.2 mg). Finally, the fractionation of subfraction W-4 (1.3 g) was separated via silica gel CC and eluted repeatedly with CHCl₃-EtOAc (8:1, 4:1, and 2:1) to yield five smaller subfractions (W-4.1 to W-4.5), which were separated by passage over a Sephadex LH-20 column and then applied to repeated silica gel CC (LiChroprep RP-C₁₈ column) and a silica gel CC eluted successively with CH₂Cl₂-EtOAc-H₂O (5:1:0.1) and CHCl₃-acetone (1:1) to yield (26*R*)-cimicifugoside (**12**, 9.8 mg to yield).

Cell cultures – Bone marrow-derived dendritic cells were grown from wild-type C57BL/6 mice (Orient Bio Inc., Seoul, Korea). All animal procedures were approved by and performed according to the guidelines of the Institutional Animal Care and Use Committee of Jeju National University (#2010-0028). Briefly, the mouse

tibia and femur were obtained by flushing with Dulbecco's modified Eagle medium to yield bone marrow cells. The cells were cultured in Rosell Park Memorial Institute (RPMI) 1640 medium containing 10% heat-inactivated fetal bovine serum (FBS; Gibco, Grand Island, NY, USA), 50 μ M β -mercaptoethanol, and 2 mM glutamine supplemented with 3% J558L hybridoma cell culture supernatant containing granulocyte-macrophage colony-stimulating factor (GM-CSF). The culture medium was replaced with fresh medium every other day. At day six of culture, non-adherent cells and loosely adherent dendritic cell (DC) aggregates were harvested, washed, and resuspended in RPMI 1640 supplemented with 5% FBS.

Cytokine production measurements – The BMDCs were incubated in 48-well plates in 0.5 mL containing 1×10^5 cells per well, and then treated with the extract, fractions and isolated compounds (**1** - **12**) at various concentrations for 1 h before stimulation with 10 ng/mL LPS from *Salmonella minnesota* (Alexis, Farnham, UK, USA). Supernatants were harvested 18 h after stimulation. Concentrations of murine IL-12 p40, IL-6, and TNF- α , in the culture supernatant were determined by ELISA (BD PharMingen, San Diego, CA, USA) according to the manufacturer's instructions. The data are presented as means \pm S.D. of at least three independent experiments performed in triplicate. The inhibitory effects are represented as giving 50% inhibition (IC_{50}) relative to the vehicle (DMSO) control. SB203580, an inhibitor of p38 MAP kinase, was used as a positive control.

Result and discussion

The methanol extract and dichloromethane and water fractions were subjected to cell treatment to evaluate the level of cytokines in LPS-stimulated BMDCs. The methanol extract of *C. dahurica* inhibited IL-12p40, IL-6, and TNF- α production (IC_{50} = 2.45, 1.21, and 22.0 μ g/mL, respectively; Table 1). Since the methanol extract

significantly reduced inflammation, it was partitioned to obtain a dichloromethane-soluble portion and an aqueous phase. As shown in Table 1, the dichloromethane-soluble fraction strongly inhibited LPS-stimulated IL-12p40, IL-6, and TNF- α production (IC_{50} = 6.82, 0.07, and 11.62 μ g/mL, respectively), while the aqueous layer strongly suppressed the production of IL-12p40 (IC_{50} = 2.43 μ g/mL) and TNF- α (IC_{50} = 2.98 μ g/mL) (Table 1).

All of the compounds (**1** - **12**, Fig. 1) isolated from the dichloromethane fraction and water layer of the *C. dahurica* extract were tested for inhibitory effects on the production of the pro-inflammatory cytokines IL-12p40, IL-6, and TNF- α . The results of 3-(4,5-dimethyl-2,5-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assays showed that compounds **1** - **12** did not exhibit significant cytotoxicity at the concentrations tested (data not shown). The positive control, SB203580, an inhibitor of p38 MAP kinase, inhibited IL-12, IL-6, and TNF- α production with IC_{50} values of 5.00 ± 0.15 , 3.50 ± 0.25 , and 7.20 ± 0.21 μ M, respectively (Table 2). Of the tested compounds, **1** - **8** and **10** - **12** inhibited IL-12p40 production more strongly than SB203580, with IC_{50} values of 3.0 μ M or less (Table 2). The variability in inhibition of the inflammatory response by **1** - **8** and **10** - **12** may be explained by the different levels of secreted inflammatory factors following LPS stimulation. Compound **6** (a cycloartane-type triterpenoid) strongly inhibited LPS-stimulated IL-12p40 production (IC_{50} = 1.92 μ M), at a level comparable to that of SB203580 (IC_{50} = 5.00 μ M), while the inhibitory effect of compound **9** on IL-12p40 production was moderate (IC_{50} = 16.35 μ M). When the sugar unit at C-3 of the aglycone in compound **9** was linked to a xylose chain, the effect was weaker than that of compound **6**, suggesting that a hydroxyl group at position C-3 of aglycone is an important functional element.

Regarding IL-6, the methanol extract, dichloromethane-soluble fraction, and water layer strongly inhibited LPS-stimulated IL-6 production (IC_{50} s = 1.21, 0.07, and 24.00

Table 1. Anti-inflammatory effects of extract and fractions on LPS-stimulated bone marrow-derived dendritic cells

Extract and fractions		IC_{50} (μ g/mL) ^a		
		IL-12 p40	IL-6	TNF- α
Extract	Methanol	2.45 ± 0.12	1.21 ± 0.03	22.00 ± 1.21
Fractions	Dichloromethane	6.82 ± 0.31	0.07 ± 0.00	11.62 ± 0.32
	Water	2.43 ± 0.34	24.00 ± 1.02	2.98 ± 0.15
SB203580 ^b		5.00 ± 0.15	3.50 ± 0.25	7.20 ± 0.21

^a The inhibitory effects are represented as giving 50% inhibition (IC_{50}) relative to the vehicle control. These data represent the average values of three repeated experiments (mean \pm SD). IC_{50} values for selected extracts are given in column IL-12 p40, IL-6 and TNF- α . Values < 25 μ M are considered to be active.

^b SB203580 was used as a positive control.

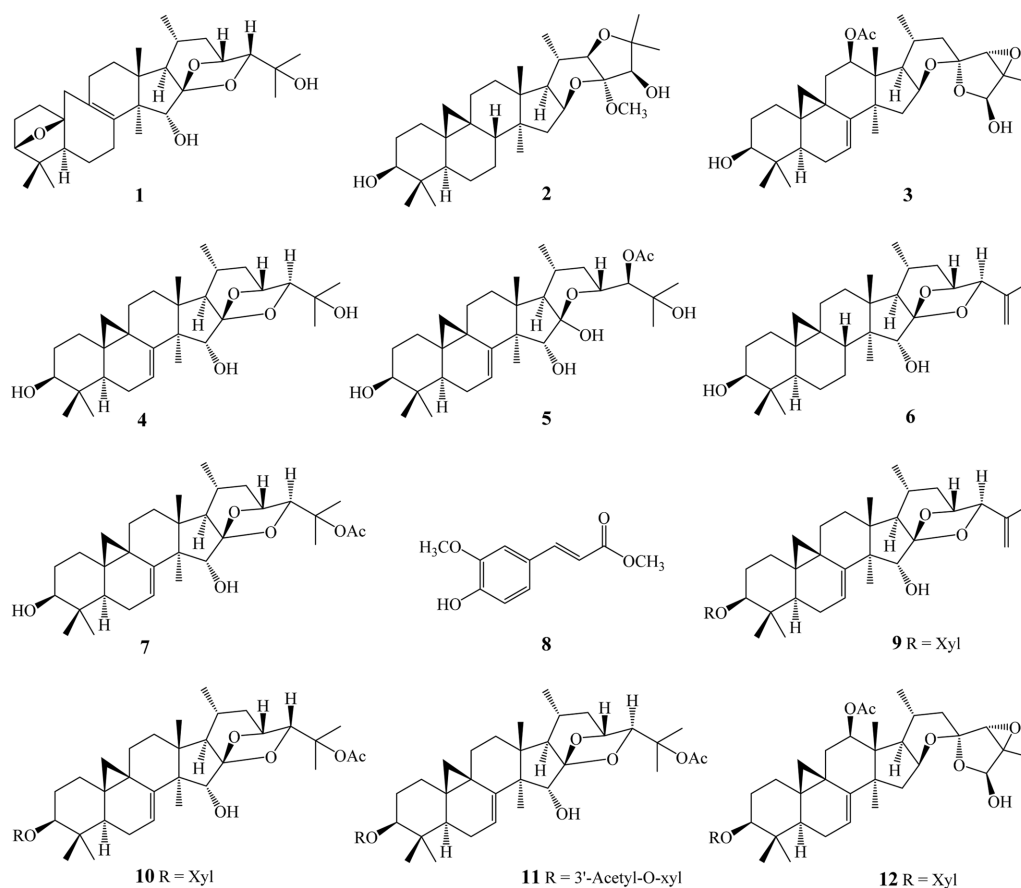


Fig. 1. Structures of compounds **1** - **12** isolated from *C. dahurica*.

Table 2. Anti-inflammatory effects of compounds on LPS-stimulated bone marrow-derived dendritic cells

Compounds	IC ₅₀ (μM) ^a		
	IL-12 p40	IL-6	TNF-α
1	2.07 ± 0.12	12.20 ± 0.56	8.17 ± 0.32
2	1.11 ± 0.09	8.10 ± 0.45	2.71 ± 0.11
3	3.90 ± 0.21	0.45 ± 0.02	13.41 ± 0.08
4	1.91 ± 0.13	0.02 ± 0.00	3.11 ± 0.16
5	1.27 ± 0.05	6.90 ± 0.45	3.37 ± 0.16
6	1.92 ± 0.08	21.77 ± 1.45	4.60 ± 0.33
7	2.62 ± 0.16	0.41 ± 0.33	4.16 ± 0.35
8	3.42 ± 0.21	4.41 ± 0.24	4.77 ± 0.35
9	16.35 ± 1.06	0.01 ± 0.00	12.20 ± 0.98
10	1.71 ± 0.07	0.04 ± 0.00	17.39 ± 1.02
11	1.05 ± 0.08	1.32 ± 0.09	17.30 ± 1.03
12	2.40 ± 0.16	4.81 ± 0.34	4.78 ± 0.35
SB203580 ^b	5.00 ± 0.15	3.50 ± 0.25	7.20 ± 0.21

^a The inhibitory effects are represented as giving 50% inhibition (IC₅₀) relative to the vehicle control. These data represent the average values of three repeated experiments (mean ± SD). IC₅₀ values for selected extracts are given in column IL-12 p40, IL-6 and TNF-α. Values < 25 μM are considered to be active.

^b SB203580 was used as a positive control.

μg/mL, respectively). Of the tested compounds, **3**, **4**, **7**, and **9** - **11** exerted greater inhibitory effects than SB203580 on IL-6 production with IC₅₀ values of 0.45, 0.023, 0.41, 0.01, 0.04, and 1.32 μM, respectively (Table 2). This variability in the inhibition of the inflammatory response by **3**, **4**, **7**, and **9** - **11** may be explained by the secretion of different levels of inflammatory factors upon LPS stimulation.

Overexpression of the pro-inflammatory cytokines TNF-α and IL-6 is associated with the development of autoimmune, inflammatory, and immunopathological diseases. Therefore, blocking TNF-α, IL-6, and their respective signaling pathways might be effective in the treatment of inflammatory diseases. Our results indicated that the water layer inhibited LPS-stimulated TNF-α production (IC₅₀ = 2.98 μg/mL). Notably, compounds **2**, **4** - **8**, and **12** strongly inhibited TNF-α production (IC₅₀s = 2.71, 3.11, 3.37, 4.60, 4.16, 4.77, and 4.78 μM, respectively; Table 2). Nevertheless, the water layer was more active than these compounds (**2**, **4** - **8**, and **12**), suggesting the presence of other active derivatives that acted individually or synergistically with the isolated cycloartane-type deri-

vatives.

The IC₅₀ values of the seven compounds (**2** - **5**, **7**, **8**, and **12**) that inhibited the pro-inflammatory cytokines IL-12p40, IL-6, and TNF- α in LPS-stimulated BMDCs were much higher. The most promising activity was found with cycloartane-type triterpenoid compounds, which appear to have specific anti-inflammatory activity. It is difficult to recognize clear structure–activity relationships within the series of compounds discovered. However, there was a clear, albeit weak, correlation with their lipophilicity, with higher activity with increasing lipophilicity.

In conclusion, this work is the first to report the inhibitory effect of extracts of *C. dahurica* on pro-inflammatory cytokine production by LPS-stimulated BMDCs *in vitro*. The results support their ethnomedical use in the treatment of inflammation. This report suggests why these medicinal plants possess anti-inflammatory properties and could provide a scientific basis for their use as supplementary herbal products for treating inflammation.

Acknowledgments

This work was supported by research fund of Chungnam National University.

References

- (1) Schwab, J. M.; Serhan, C. N. *Curr. Opin. Pharmacol.* **2006**, *6*, 414-420.
- (2) Efron, P. A.; Tsujimoto, H.; Bahjat, F. R.; Ungaro, R.; Debernardis, J.; Tannahill, C.; Baker, H. V.; Edwards, C. K.; Moldawer, L. L. *J. Endotoxin Res.* **2005**, *11*, 145-160.
- (3) Ueno, H.; Klechevsky, E.; Morita, R.; Asford, C.; Cao, T.; Matsui, T.; Di Pucchio, T.; Connolly, J.; Fay, J. W.; Pascual, V.; Palucka, A. K.; Banchereau, J. *Immunol. Rev.* **2007**, *219*, 118-142.
- (4) Gubler, U.; Chua, A. O.; Schoenhaut, D. S.; Dwyer, C. M.; McComas, W.; Motyka, R.; Nabavi, N.; Wolitzky, A. G.; Quinn, P. M.; Familletti, P. C. *Proc. Natl. Acad. Sci. U. S. A.* **1991**, *88*, 4143-4147.
- (5) Kishimoto, T. *Int. Immunol.* **2010**, *22*, 347-352.
- (6) Gahring, L. C.; Carlson, N. G.; Kulmar, R. A.; Rogers, S. W. *Neuroimmunomodulation* **1996**, *3*, 289-303.
- (7) Beutler, B.; Cerami, A. *Nature* **1986**, *320*, 584-588.
- (8) Chinese Pharmacopoeia Commission. Pharmacopoeia of the People's Republic of China, vol. 1.; China Medical Science and Technology Press, china, **2010**, pp 68-69.
- (9) Qin, R.; Zhao, Y.; Zhao, Y.; Zhou, W.; Lv, C.; Lu, J. *Fitoterapia* **2016**, *115*, 52-56.
- (10) Zhang, L. L.; Si, J. Y.; Zhang, L. J.; Xiao-Wei, H.; Lin, L.; Li, R. Y.; Chen, D.; Cao, L. *Chin. J. Integr. Med.* **2016**, 1-9.
- (11) Tian, Z.; Si, J.; Chang, Q.; Zhou, L.; Chen, S.; Xiao, P.; Wu, E. *BMC Cancer* **2007**, *7*, 237.
- (12) Lv, C.; Yang, F.; Qin, R.; Qi, Z.; Zhou, W.; Lu, J. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 3305-3309.
- (13) Thao, N. P.; Kim, J. H.; Thuy Luyen, B. T.; Dat, N. T.; Kim, Y. H. *Int. J. Biol. Macromol.* **2017**, *98*, 526-534.
- (14) Thao, N. P.; Luyen, B. T.; Lee, J. S.; Kim, J. H.; Kim, Y. H. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 1874-1879.
- (15) Thao, N. P.; Luyen, B. T. T.; Lee, J. S.; Kim, J. H.; Dat, N. T.; Kim, Y. H. *J. Nat. Prod.* **2017**, *80*, 1867-1875.

Received March 21, 2018

Revised April 30, 2018

Accepted May 1, 2018