

An immunohistochemical study on the pancreatic islets cells of the Mongolian gerbils, *Meriones unguiculatus*

Sae-kwang Ku, Hyeung-sik Lee^{*1}, Ki-dae Park² and Jae-hyun Lee²

Pharmacology & Toxicology Lab., Central Research Laboratories, Dong-Wha Pharm. Ind. Co., Anyang 430-017, Korea

¹Department of Biology, Faculty of Natural Sciences, Kyungsan University, Kyungsan 712-240, Korea

²Department of Histology, College of Veterinary Medicine, Kyungpook National University, Taegu 702-701, Korea

In order to study the regional distribution and relative frequency of the immunoreactive endocrine cells in the pancreatic islets of the Mongolian gerbil, pancreatic sections of *Meriones unguiculatus* were immunostained using an immunohistochemical (PAP) method with four types of specific antisera against insulin, glucagon, somatostatin and human pancreatic polypeptide (PP). The pancreatic islets were subdivided into three portions (central region, mantle zone and peripheral region) according to their composition of immunoreactive cells. Spherical to spindle shaped insulin, glucagon, somatostatin and PP-immunoreactive cells were observed in this study. Insulin-immunoreactive cells were present in the central regions with high frequency, and a few of these cells were also demonstrated in the mantle zones. Glucagon-immunoreactive cells were mainly restricted to the mantle zones. However, rare examples were found in the peripheral regions. As for the glucagon-immunoreactive cells, somatostatin-immunoreactive cells were detected in the mantle zones and peripheral regions with moderate and rare frequencies, respectively. PP-immunoreactive cells were found in the mantle zones and peripheral regions with rare and moderate frequencies, respectively. In the mantle and the peripheral regions, cytoplasmic process of glucagon-, somatostatin- and PP-immunoreactive cells were intermingled. In conclusion, the regional distribution of endocrine cells in the pancreatic islets of Mongolian gerbil was found to be similar to that of other mammals, especially other rodents, except for the topographical different distribution of somatostatin which differs that of other rodents.

Key words: Mongolian gerbil, pancreatic islets, endocrine cell, immunohistochemistry

Introduction

It is generally known that the pancreas of vertebrates is subdivided into exocrine and endocrine portions. Digestive enzymes are released in the exocrine and regulatory hormones such as insulin, glucagon, somatostatin and pancreatic polypeptide (PP) are produced in the endocrine and released into blood circulation. The appearance, regional distribution and relative frequency of these regulatory hormones secreted by endocrine cells in the pancreas are well recognized by their histochemistry using [17], the immunofluorescence method [23] and immunohistochemistry [32]. In addition to the above regulatory hormones, peptide YY-, neuropeptide YY- [1], chromogranin family- [14, 27] and motilin- [35] immunoreactive cells have also been demonstrated in the vertebrate pancreas. The pancreas has been treated as a valuable organ for endocrine study and the endocrine pancreas has been extensively studied in association with diabetes [15]. In addition, investigations of gastroenteropancreatic (GEP) endocrine cells are considered to be an important part of phylogenetic study [6].

The Mongolian gerbil, *Meriones unguiculatus*, is a rodent of the family Cricetidae, although it has been included alternatively among the Muridae. The animal is an active, nearly odorless, usually nonaggressive rodent distinguished by its monogamous mating behavior, water and temperature conservation mechanisms, spontaneous epileptiform seizures, relative freedom from spontaneous disease, and several other unique attributes of interest in research [11].

Until now, the regional distribution and relative frequency of four major immunoreactive cells, insulin-, glucagon-, somatostatin and PP-, have been reported in the pancreas of the hamster [3], sand rat (*Psammomys obesus*) [8], C57BL/6 mouse [10], herbivorous Japanese field vole (*Microtus montebelli*) [22], guinea pig [25], vole (*Microtus arvalis*) [28], obese ob+/ob+ mouse [30], preobese and obese yellow Avy/- mice [33] and wood mouse (*Apodemus speciosus*) [36]. In addition, angiotensin II-immunoreac-

*Corresponding author

Phone: +82-53-819-1436; Fax: +82-53-819-1558

E-mail: endohist@kyungsan.ac.kr

tive cells were found in the pancreas of the mouse [20] and the appearance of calcitonin gene-related peptide- and cholecystokinin-immunoreactive cells have been reported in the rat pancreas [7, 29]. With the increasing demand for diabetic animal models in many fields, the regional distribution and relative frequency of pancreatic endocrine cells, especially, insulin- and glucagon-producing cells in laboratory animals is of interest [9, 10, 33]. It has been accepted that insulin-immunoreactive cells are located in the central regions and that the other immunoreactive cells such as glucagon-, somatostatin- and PP-immunoreactive cells are located in the peripheral or mantle zones. But, many researchers have suggested that species-dependent hormone producing cell distributions in the pancreas of different species might due to feeding habits, and this is now generally accepted [34]. In addition, it has also been reported that different regional distributions and relative frequencies of endocrine cells in the pancreatic islets were demonstrated in different portion of the pancreas, which included the pancreas of single animals [36] and strain-dependent characteristic distributions of these immunoreactive cells was also detected in connection with attempts to increase the production of genetically mutated laboratory animals, and to increase the breeding rate of laboratory animals having specific diseases or unique characteristics, especially in the rat and mouse [9, 10, 30, 33, 36].

Although many studies have concerned the regional distribution and relative frequency of different endocrine cells in the pancreas of various vertebrates including several species and strains of rodents, there have been no reports on immunohistochemical studies into the endocrine cells of the pancreatic islets of the Mongolian gerbil, in spite of

their biological, physiological and anatomical differences from the other rodents. The purpose of the present study was to clarify the regional distribution and relative frequency of endocrine cells in the pancreatic islets of the Mongolian gerbil, *Meriones unguiculatus* using an immunohistochemical method (PAP method) and four types of specific antisera against insulin, glucagon, somatostatin and PP.

Materials and Methods

Five adult (40~50 g of body weight) Mongolian gerbils, *Meriones unguiculatus*, were acquired from the Asan Institute for Life Science (Seoul, Korea) and were used in this study without sexual distinction. After food restriction for 24 hours, the animals were anesthetized with ethyl ether and then phlebotomized. Samples from the pancreas were fixed in Bouin's solution, and after paraffin embedding, 3-4 μ m serial sections were prepared. Representative sections of each tissue were stained with hematoxylin and eosin for light microscopic examination of the normal gastrointestinal architecture.

Each representative section was deparaffinized, rehydrated and immunostained by the peroxidase anti-peroxidase (PAP) method [31]. Nonspecific reactions were blocked with normal goat serum prior to incubation with the specific antisera (Table 1). After rinsing in phosphate buffered saline (PBS; 0.01 M, pH 7.4), the sections were incubated in secondary antiserum. They were then washed in PBS buffer and finally the PAP complex was prepared. The peroxidase reaction was carried out in a solution of 3,3'-diaminobenzidine tetrahydrochloride, containing

Table 1. Antisera used in this study

Antisera raised*	Code	Source	Dilution
Insulin	PUO290395	BioGenex Lab., San Ramon.	1 : 20
Glucagon	PUO390598	BioGenex Lab., San Ramon.	1 : 20
Somatostatin	PUO421295	BioGenex Lab., San Ramon.	1 : 20
PP ¹⁾	PUO660495	BioGenex Lab., San Ramon.	1 : 20

*All antisera were raised in rabbits except for insulin, which was raised in rabbits.

¹⁾PP: human pancreatic polypeptide

Table 2. Regional distributions and relative frequencies of the endocrine cells in the pancreatic islets of the Mongolian gerbil, *Meriones unguiculatus*

Immunoreactive cells	Pancreatic islets		
	Central region	Mantle zone	Peripheral region
Insulin	+++	+	-
Glucagon	-	+++	±
Somatostatin	-	++	±
PP ¹⁾	-	±	++

* Relative frequencies; +++: numerous, ++: moderate, +: a few, ±: rare, -: not detected

¹⁾ PP: human pancreatic polypeptide.

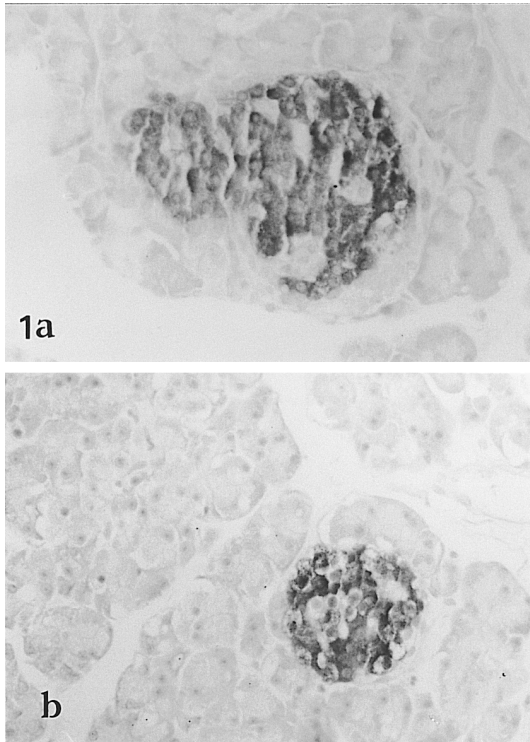


Fig. 1. Insulin-immunoreactive cells in the pancreatic islets of Mongolian gerbils. Note that most of the immunoreactive cells were located in the central regions of pancreatic islets regardless of size. a, b: $\times 240$. PAP method.

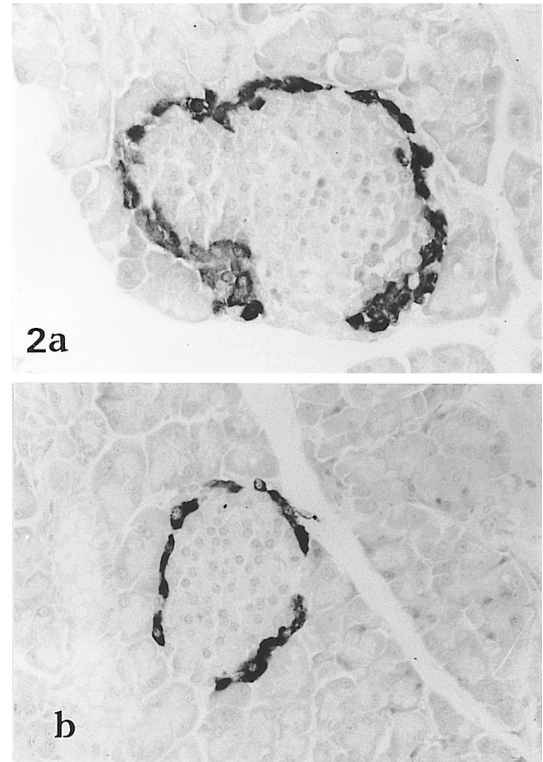


Fig. 2. Glucagon-immunoreactive cells in the pancreatic islets of Mongolian gerbils. Note that most of the immunoreactive cells were located in the mantle zones of pancreatic islets regardless of size. a, b: $\times 240$. PAP method.

0.01% H_2O_2 in Tris-HCl buffer (0.05M, pH 7.6). After immunostaining, the sections were lightly counterstained with Mayer's hematoxylin and immunoreactive cells were observed under a light microscope.

The specificity of each immunohistochemical reaction was determined as recommended by Sternberger [31], including the replacement of specific antiserum by the same antiserum, which had been preincubated with its corresponding antigen. The relative frequency of occurrence of each type of immunoreactive cell was allocated to one of five categories according to its frequency, as observed by light microscopy.

Results

In this study, all four kinds of the immunoreactive endocrine cells were detected using antisera against insulin, glucagon, somatostatin and PP in the pancreatic islets, which were distinguished as three distinct layers, a central region, a mantle zone and a peripheral region with their composition of immunoreactive cells. Different regional distributions and relative frequencies of these immunoreactive cells were observed in the different pancreatic regions, and these differences are shown in Table 2. Spherical to spindle or occasionally oval to round-shaped immu-

noreactive cells were observed in the pancreatic islets of the Mongolian gerbil.

Insulin-immunoreactive cells

Spherical to spindle shaped insulin-immunoreactive cells were located in the central pancreatic islet region with numerous frequency and rarely round to oval shaped cells of variable size were also observed. In addition, a few frequented cells were also observed in the mantle zone intermingled with other immunoreactive cells, especially glucagon- and somatostatin-immunoreactive cells. However, no insulin-immunoreactive cells were found in the peripheral regions, which predominantly contained PP-immunoreactive cells (Fig. 1a, b).

Glucagon-immunoreactive cells

Spherical to spindle shaped glucagon-immunoreactive cells were located in the mantle and peripheral regions of the pancreatic islets with numerous and rare frequencies, respectively, regardless of size. Occasionally, rare round to ovally shaped cells were also observed in these regions. In the mantle and peripheral regions, the cytoplasmic processes of glucagon-immunoreactive cells were intermingled with other immunoreactive cells especially

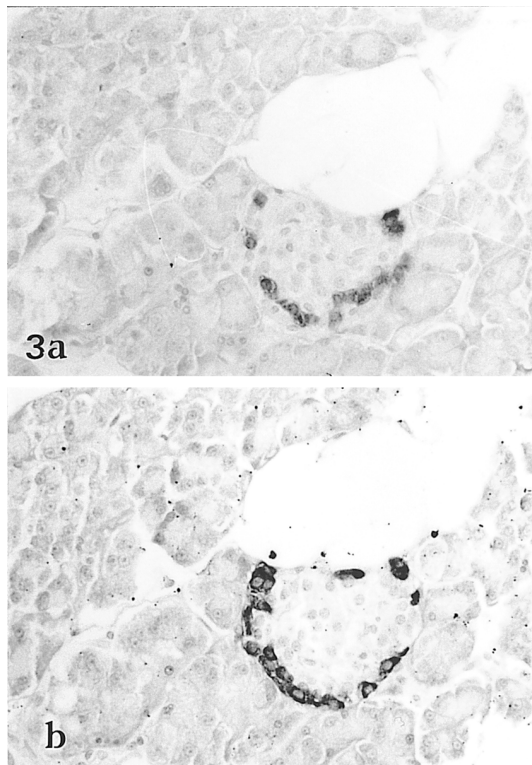


Fig. 3. Somatostatin (a)- and human pancreatic polypeptide (b)-immunoreactive cells in the pancreatic islets of Mongolian gerbils. Note that most of the somatostatin-immunoreactive cells were located in the mantle zones of pancreatic islets regardless of size while human pancreatic polypeptide-immunoreactive cells were observed in the outermost peripheral regions of pancreatic islets. a, b: $\times 240$. PAP method.

somatostatin- and PP-immunoreactive cells. However, no glucagon-immunoreactive cells were observed in the central regions, where numerous insulin-immunoreactive cells were found (Fig. 2a, b).

Somatostatin-immunoreactive cells

Spherical to spindle shaped somatostatin-immunoreactive cells were found in the mantle and peripheral regions of the pancreatic islets with moderate and rare frequencies, respectively, regardless of size. Occasionally, rare round to ovally shaped cells were also observed in these regions. In the mantle and peripheral regions, the cytoplasmic processes of these immunoreactive cells were intermingled with other immunoreactive cells, especially glucagon- and PP-immunoreactive cells. However, no somatostatin-immunoreactive cells were observed in the central regions where numerous insulin-immunoreactive cells were found (Fig. 3a).

PP-immunoreactive cells

Spherical to spindle shaped somatostatin-immunoreactive cells were observed in the mantle and peripheral

regions of the pancreatic islets with rare and moderate frequencies, respectively, regardless of their size. Occasionally, rare round to ovally shaped cells were also found in these regions. In the mantle and peripheral regions, the cytoplasmic processes of PP-immunoreactive cells were intermingled with other immunoreactive cells, especially glucagon- and somatostatin-immunoreactive cells. However, no PP-immunoreactive cells were demonstrated in the central regions, where numerous insulin-immunoreactive cells were found (Fig. 3a).

Discussion

Unlike other rodents, Mongolian gerbils of both sexes have a distinct midventral abdominal pad composed of large sebaceous glands under the control of gonadal hormones [4], and Mongolian gerbil has unique feeding habits [11]. In addition, the male gerbils have higher packed red-cell volumes (PCV), hemoglobin levels, total leukocyte counts, and circulating lymphocyte counts than the females, and some erythrocytes of both sexes show a prominent polychromasia and basophilic stippling [11]. In spite of their biological, physiological and anatomical differences from other rodents, no immunohistochemical studies are available on the pancreatic endocrine cells in the pancreatic islets of the Mongolian gerbil. In the present study, the four major types of endocrine cells, insulin-, glucagon-, somatostatin- and PP-immunoreactive cells that are generally found in the mammalian pancreas, were detected in the pancreatic islets.

Insulin is synthesized in the B cells of the pancreatic islets and regulates the serum glucose levels [13]. In mammals, the regional distribution and relative frequency of insulin-immunoreactive cells in the pancreas have been reported in the hamster [3], three-toed sloth (*Bradypus variegatus*) [5], C57BL/6 mouse [10], opossum [18], Australian brush-tailed possum [19], voles [28], various laboratory animals [34] and wood mouse [36]. From these reports, it is known that insulin-immunoreactive cells are situated in the central regions of the mammalian pancreas and that other cells, such as, glucagon-, somatostatin- and PP-immunoreactive cells, surrounded them. However, somewhat contradicting the finding of other researchers, Reddy *et al.* [26] reported that these-immunoreactive cells are observed in the majority of islets where they occur peripherally as groups of cells, and within the pancreatic islets of several marsupial species. In the present study, most of the insulin-immunoreactive cells were restricted to the central regions of islets in the Mongolian gerbil, which is similar to previous reports on rodents [3, 10, 28, 33, 34, 36].

Glucagon is synthesized in the A cells of the pancreas and regulates blood glucose levels [13]. Morphologically similar cells are also present in the digestive tract of the

dog. In the present study, glucagon-immunoreactive cells were found in the mantle and the peripheral regions of pancreatic islets. Although glucagon-immunoreactive cells have been found in the mantle and peripheral regions of mammalian pancreatic islets [3, 5, 10, 18, 19, 28, 33, 34, 36] including the present study, species-dependent variations have been reported in the equine pancreas, in which A-cells, demonstrated by anti-glucagon, were found in the center of pancreatic islets [12]. In addition, it has also been reported that under specific disease conditions, such as, those in the obese (diabetic condition) mouse, glucagon-immunoreactive cells are intermingled with insulin-immunoreactive cells in the central regions of the pancreatic islets. In contrast, normal non-obese littermates showed a peripheral localization of these immunoreactive cells [30].

Somatostatin, which consists of 14 amino acids, was isolated initially from the hypothalamus of sheep. It was found to be present in straight and cyclic forms [2]. This substance inhibits the secretion of gastrin, cholecystokinin, secretin, glucagon, insulin, motilin and gastric acid [16] and the absorption of amino acids, glucose and fatty acids in the gastrointestinal tract [2]. To date, somatostatin-immunoreactive cells have been found in the outermost regions of mammalian pancreatic islets [3, 5, 10, 18, 19, 28, 33, 34, 36]. However, in the present study, most of these immunoreactive cells were found in the mantle zones, mixed with glucagon-immunoreactive cells; PP-immunoreactive cells were found to occupy the outermost regions of pancreatic islets in this study. These topographically different distributional patterns in mammalian species [3, 5, 10, 18, 19, 28, 33, 34, 36] are considered as species-dependent variations, and intra-species topographical variations are considered as reflections of unique diseases and suggests that under specific disease conditions, such as obesity (diabetic condition) mouse, somatostatin-immunoreactive cells show different distributional patterns [30].

PP is a peptide hormone, which contains 36 amino acids, and is synthesized by F cells in the pancreatic islets [13]. The specific function of this peptide is not clear, however, it has been postulated that it is related to food intake inhibition [13] and Polak et al [24] reported that it promoted the secretion of gastric acid and stimulated the glycogenesis of liver in avian species. It has been reported that PP-immunoreactive cells are conspicuously distributed in the peripheral regions of the pancreatic islets in mammalian species [3, 10, 18, 19, 28, 33, 34, 36]. In addition, the colocalization of these immunoreactive cells with serotonin-immunoreactive cells has been demonstrated in the pancreatic islets of the opossum [18] and cattle [21] though da Mota et al. [5] reported that PP-immunoreactive cells were not found in the pancreas of the three-toed sloth. In the present study, which is in agreement with previous studies [3, 10, 18, 19, 28, 33, 34, 36], PP-immunoreactive

cells were detected in the outermost regions of the pancreatic islets, although rarely cells were intermingled with other immunoreactive cells in the mantle zone, where glucagon-immunoreactive cells predominated, followed by somatostatin-immunoreactive cells.

In conclusion, the regional distribution of endocrine cells in the pancreatic islets of Mongolian gerbil was found to be similar to that of other mammals, especially rodents, except for the topographically different distribution of somatostatin compared to that of other rodents. Cell core in the pancreatic islets of Mongolian gerbils were composed of centrally located insulin-immunoreactive cells and glucagon- and somatostatin-immunoreactive cells located in the mantle zone, PP-immunoreactive cells surrounded immunoreactive cells located in the mantle zone at the peripheral regions of pancreatic islets.

References

1. **Alli-Rachedi, A., Varndell, I. M., Adrian, T. E., Gapp, D. A., van Noorden, S., Bloom, S. R. and Polak, J. M.** Peptide YY (PYY) immunoreactivity is co-stored with glucagon-related immunoreactants in endocrine cells of the gut and pancreas. *Histochemistry* 1984, **80**(5), 487-491.
2. **Brazeau, P., Vale, W., Burgurs, R., Ling, N., Butcher, M., Rivier, J. and Guillermin, R.** Hypothalamic polypeptide that inhibits the secretion of immunoreactive pituitary growth hormone. *Science* 1973, **179**(68), 77-79.
3. **Camihort, G., Del Zotto, H., Gomez Dumm, C. L. and Gagliardino, J. J.** Quantitative ultrastructural changes induced by sucrose administration in the pancreatic B cells of normal hamsters. *Biocell* 2000, **24**(1), 31-37.
4. **Cheal, M.** Life span environmental influences on species typical behavior of *Meriones unguiculatus*. *Basic Life Sci.* 1987, **42**, 145-159.
5. **da Mota, D. L., Yamada, J., Gerge, L. L. and Pinheiro, P. B.** An immunohistochemical study on the pancreatic endocrine cells of the three-toed sloth, *Bradypus variegatus*. *Arch. Histol. Cytol.* 1992, **55**(2), 203-209.
6. **D'Este, L., Buffa, R., Pelagi, M., Siccardi, A. G. and Renda, T.** Immunohistochemical localization of chromogranin A and B in the endocrine cells of the alimentary tract of the green frog, *Rana esculanta*. *Cell Tissue Res.* 1994, **277**(2), 341-349.
7. **Ding, W. G., Guo, L. D., Kitasato, H., Fujimura, M. and Kimura, H.** Phylogenic study of calcitonin gene-related peptide-immunoreactive structures in the pancreas. *Histochem. Cell Biol.* 1998, **109**(2), 103-109.
8. **Donev, S., Petkov, P., Marquie, G., Duhault, J. and Jablenska, R.** Immunohistochemical investigations of the endocrine pancreas in normoglycemic sand rat (*Psammomys obesus*). *Acta. Diabetol. Lat.* 1989, **26**(4), 309-313.
9. **Fu, Q., Honda, M., Ohgawara, H., Igarashi, N., Toyada, C., Omori, Y. and Kobayashi, M.** Morphological analysis of pancreatic endocrine cells in newborn animals delivered by experimental diabetic rats. *Diabetes Res. Clin. Pract.*

- 1996, **31**(1-3), 57-62.
10. **Gomez Dumm, C. L., Console, G. M., Lunna, G. C., Dardenne, M. and Goya, R. G.** Quantitative immunohistochemical changes in the endocrine pancreas of nonobese diabetic (NOD) mice. *Pancreas* 1995, **11**(4), 396-401.
11. **Harkness, J. E. and Wagner, J. E.** The biology and medicine of rabbits and rodents, pp. 50-57 4th ed. Williams & Wilkins, Baltimore, 1995.
12. **Helmstaedter, V., Feurle, G. E. and Forssmann, W. G.** Insulin-, glucagon- and somatostatin-immunoreactive cells in the equine pancreas. *Cell Tissue Res.* 1976, **172**(4), 447-454.
13. **Hsu, W. H. and Crump, M. H.** The endocrine pancreas. In: McDonald, L. E. and Pineda, M. H. (ed), *Veterinary endocrinology and reproduction*, pp 186-201, Lea & Febiger, Philadelphia, 1989.
14. **Ito, H., Hashimoto, Y., Kitagawa, H., Kon, Y. and Kudo, N.** Distribution of chromogranin containing cells in the porcine gastroenteropancreatic endocrine system. *Jpn. J. Vet. Sci.* 1987, **50**, 395-404.
15. **Jansson, L. and Sandler, S.** The influence of cyclosporin A on the vascular permeability of the pancreatic islets and on diabetes induced by multiple low dose of streptozotocin in the mouse. *Virchows Archiv A Pathol. Anat. Histopathol.* 1988, **412**(3), 225-230.
16. **Kitamura, N., Yamada, J., Calingasan, N. Y. and Yamashita, T.** Immunocytochemical distribution of endocrine cells in the gastrointestinal tract of the horse. *Equine Vet. J.* 1984, **16**(2), 103-107.
17. **Kobayashi, K. and Ali, S. S.** Cell types of the endocrine pancreas in the shark, *Scylliorhinus stellaris* as revealed by correlative light and electron microscopy. *Cell Tissue Res.* 1981, **215**(3), 475-490.
18. **Krause, W. J., Cutts, J. H. 3rd, Cutts, J. H. and Yamada, J.** Immunohistochemical study of the developing endocrine pancreas of the opossum (*Didelphis virginiana*). *Acta. Anat. (Basel).* 1989, **135**(1), 84-96.
19. **Leigh, C. M. and Edwin, N. A.** Light-microscopic immunocytochemical study of the endocrine pancreas in the Australian brush-tailed possum (*Trichosurus vulpecula*). *Eur. J. Histochem.* 1992, **36**(2), 237-241.
20. **Leung, P. S., Chan, H. C. and Wong, P. Y.** Immunohistochemical localization of angiotensin II in the mouse pancreas. *Histochem. J.* 1998, **30**(1), 21-25.
21. **Nakajima, S., Kitamura, N., Yamada, J., Yamashita, T. and Watanabe, T.** Immunohistochemical study on the endocrine pancreas of cattle with special reference to coexistence of serotonin and glucagon or bovine pancreatic polypeptide. *Acta. Anat. (Basel).* **131**(3), 235-240.
22. **Ohara, N., Kitamura, N., Yamada, J. and Yamashita, T.** Immunohistochemical study of gastroenteropancreatic endocrine cells of the herbivorous Japanese field vole, *Microtus montebelli*. *Res. Vet. Sci.* 1986, **41**(1), 21-27.
23. **Orci, L.** Macro- and micro-domains in the endocrine pancreas. *Diabetes* 1982, **31**(8 pt 1), 538-564.
24. **Polak, J. M., Adrian, T. E., Bryant, M. G., Bloom, S. R., Heitz, P. H. and Pearse, A. G. E.** Pancreatic polypeptide in the insulomas, gastrinomas and glucagonomas. *Lancet* 1979, **55**, 328-330.
25. **Reddy, S. N., Bibby, N. J. and Elliott, R. B.** Cellular distribution of insulin, glucagon, pancreatic polypeptide hormone and somatostatin in the fetal and adult pancreas of the guinea pig: a comparative immunohistochemical study. *Eur. J. Cell Biol.* 1985, **38**(2), 301-305.
26. **Reddy, S., Bibby, N. J., Fisher, S. L. and Elliott, R. B.** Immunolocalization of insulin, glucagon, pancreatic polypeptide and somatostatin in the pancreatic islets of the possum, *Trichosurus vulpecula*. *Gen. Comp. Endocrinol.* 1986, **64**(1), 157-162.
27. **Rindi, G., Buffa, R., Sessa, F., Tortora, O. and Solcia, E.** Chromogranin A, B and C immunoreactivities of mammalian endocrine cells: Distribution from costored hormones/prohormones and relationship with argyrophil component of secretory granules. *Histochemistry* 1986, **85**(1), 19-28.
28. **Sasaki, M., Arai, T., Usui, T. and Oki, Y.** Immunohistochemical, ultrastructural, and hormonal studies on the endocrine pancreas of voles (*Microtus arvalis*) with monosodium aspartate-induced diabetes. *Vet. Pathol.* 1991, **28**(6), 497-505.
29. **Shimizu, K., Kato, Y., Shiratori, K., Ding, Y., Song, Y., Furlantto, R., Chang, T. M., Watanabe, S., Hayashi, N., Kobayashi, M. and Chey, W. Y.** Evidence for existence of CCK-producing cells in rat pancreatic islets. *Endocrinology* 1998, **139**(1), 389-396.
30. **Starich, G. H., Zafirova, M., Jabelenska, R., Petkov, P. and Lardinois, C. K.** A morphological and immunohistochemical investigation of endocrine pancreas from obese ob+/ob+ mice. *Acta. Histochem.* 1991, **90**(1), 93-101.
31. **Sternberger, L. A.** The unlabeled antibody peroxidase-antiperoxidase (PAP) method. In: Sternberger, L. A. (ed), *Immunocytochemistry*, pp. 104-169, John Wiley & Sons, New York, 1979.
32. **Sternberger, L. A., Hardy, P. H., Cuculis, J. J. and Meyer, H. G.** The unlabeled antibody enzyme method of immunocytochemistry: Preparation and properties of soluble antigen-antibody complex (horseradish peroxidase-antihorseradish peroxidase) and use in identification of spirochetes. *J. Histochem. Cytochem.* 1970, **18**(5), 315-333.
33. **Warbritton, A., Gill, A. M., Yen, T. T., Bucci, T. and Wolff, G. L.** Pancreatic islet cells in preobese yellow Avy/mice: relation to adult hyperinsulinemia and obesity. *Proc. Soc. Exp. Biol. Med.* 1994, **206**(2), 145-151.
34. **Wieczorek, G., Pospischil, A. and Perentes, E. A.** Comparative immunohistochemical study of pancreatic islets in laboratory animals (rats, dogs, minipigs, nonhuman primates). *Exp. Toxicol. Pathol.* 1998, **50**(3), 151-172.
35. **Yamada, J., Campos, V. J. M., Kitamura, N., Pacheco, A. C., Yamashita, T. and Yanaihara, N.** An immunohistochemical study of endocrine cells in the pancreas of *Caiman latirostris* (Alligatorinae), with special reference to pancreatic motilin cells. *Biomed. Res.* 1986, **7**, 199-208.
36. **Yukawa, M., Takeuchi, T., Watanabe, T. and Kitamura, S.** Proportions of various endocrine cells in the pancreatic islets of wood mice (*Apodemus speciosus*). *Anat. Histol. Embryol.* 1999, **28**(1), 13-16.