

# Decreased Chitotriosidase Activity and Levels in Familial Mediterranean Fever

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Different studies have demonstrated changes in chitotriosidase (ChT) activity and concentrations in multiple diseases. However, changes in ChT activity and concentrations have not been concurrently evaluated in patients with Familial Mediterranean Fever (FMF). In this study, we analyzed the changes in serum ChT activity and concentrations in patients with FMF. The study included a total of 80 patients with FMF and 80 healthy controls. ChT enzyme activity and concentrations were measured and then compared between the groups. ChT activity was measured by using fluorometric ELISA and ChT concentrations were measured by using colorimetric ELISA methods. The median ChT activity was 10.00 (6.00–15.00) nmol/mL/hr in the patients and 14.00 (6.25–20.75) nmol/mL/hr in the controls. There was a statistically significant difference in the ChT activity between the controls and patients ( $P = 0.027$ ). The median ChT concentrations were 65.40 (46.20–84.92) pg/mL and 125.00 (75.72–143.95) pg/mL in the patients and controls, respectively ( $P < 0.001$ ), which were expressed as median percentiles (25th–75th). Additionally, we found no correlation between C-reactive protein and ChT activity ( $P = 0.978$ ,  $r = 0.003$ ) and concentrations ( $P = 0.446$ ,  $r = -0.87$ ). Serum ChT enzyme activity and concentrations may not be considered as a biomarker in FMF patients taking colchicine. New studies are needed to evaluate the changes of enzyme activity and concentration in colchicine-negative patients.

**Keywords:** Chitotriosidase Activity; Familial Mediterranean Fever; Chitotriosidase Concentration

## INTRODUCTION

Familial Mediterranean fever (FMF) is an autosomal recessive autoinflammatory disease. Recurrent fever, abdominal attacks, pleural attacks, prodromes, and pericarditis are common clinical manifestations of FMF. Mutations in the Mediterranean fever (MEFV) gene encoding of a protein called pyrin appear to cause the disease in many cases. Mutations have been widely found in exons 1, 2, 3, 5, 9, and 10 of the MEFV gene. The five most frequent mutations are E148Q, M680I, M694V, M694I, and V726A (1–3). Whereas FMF is widely found among Mediterranean and Middle Eastern populations, sporadic cases have been reported in the Far East (4).

Abnormal innate immune system activation, which is responsible for activating inflammasome, is the main player in the pathogenesis of autoinflammatory diseases (5). The proposed molecular mechanism in the pathogenesis of FMF is increased inflammasome activation due to restricted pyrin expression (6). Inflammasomes activate caspase 1 and trigger the release of pro-inflammatory cytokines, such as interleukin-1 $\beta$  (IL-1 $\beta$ ) and IL-18. These cytokines stimulate neutrophils and macrophages

and induce an inflammatory response (7).

Two different phenotypes (types 1 and 2) might be seen in patients with FMF. Type 1 is characterized by short recurrent episodes of inflammation. The episodes may last from 24 to 72 hours and have variable frequency. The frequency of attacks may vary from once per week to once every 5–10 years. Approximately half of the affected individuals report experiencing a prodrome of discomfort and/or psychological uneasiness up to 24 hours before the actual attack (1,2). Colchicine is used in the treatment of FMF. It reduces the frequency of attacks and prevents amyloidosis (8).

Chitotriosidase (ChT) is a member of the chitinase family capable of hydrolyzing chitin. This enzyme is mainly secreted from activated and differentiated macrophages, therefore, ChT is a marker of macrophage activation (9). Human ChT is associated with Gaucher's disease, lysosomal storage disorders, sarcoidosis, Crimean Congo hemorrhagic fever, and beta-thalassemia (10–13). It has also been indicated that ChT can be up-regulated during the immunological response in other acute or chronic inflammatory conditions (14). Therefore, we wanted to investigate whether differences in ChT concentrations or activity could

be seen in patients with FMF.

The purpose of this study was to investigate the changes in the activity and concentration of ChT in FMF patients compared to healthy controls. To the best of our knowledge, this is the first study to evaluate both ChT activity and concentrations in FMF patients.

## MATERIALS AND METHODS

### Patients and controls

The study groups were comprised of 80 FMF patients (37 males and 43 females; 18-68 years old [mean age:  $34.86 \pm 9.5$ ]) and 80 controls (38 males and 42 females; 23-50 years old [mean age:  $35.43 \pm 6.44$ ]). FMF diagnoses were made according to the Tel-Hashomer criteria (15). Twenty patients were in an acute attack period of FMF. The diagnosis of FMF attacks was confirmed by the presence of fever, clinical findings of serositis/arthritis, skin rash, and elevated C-reactive protein (CRP  $> 5$  mg/L) concentrations. Seventy-two patients had been receiving stable doses of colchicine (1.5 mg/day), two patients had been receiving stable doses of both Anakinra (100 mg/day) and colchicine (2 mg/day), and two patients had been receiving stable doses of both colchicine and a nonsteroidal anti-inflammatory drug. We obtained information such as age, gender, MEFV mutation types, and the concentrations of some analytes of the study population from the records of the Ankara Numune Education and Training Hospital's laboratory information system. Patients with impaired renal and thyroid function; diabetes mellitus; rheumatic disease; gut, musculoskeletal, and skin diseases; liver disease; malignancy; and pregnancy were excluded from the study. For the healthy controls, the exclusion criteria included a clinical suspicion of infections (body temperature out of the range of  $36^{\circ}\text{C}$ - $38^{\circ}\text{C}$ , heart rate  $> 90$  rate/minute, respiratory rate  $> 20$ /minute, and white blood count  $> 12,000/\text{mm}^3$  or  $< 4,000/\text{mm}^3$ ); the presence of liver disease, kidney disease, rheumatic disease, or malignancy; pregnancy, and smoking. Blood samples were sent by physicians from Ankara Numune Education and Training Hospital, Department of Rheumatology.

### Samples

Overnight fasting venous blood samples were collected into red top tubes (Becton Dickinson, UK) after a 12-hour fast. The serum samples were allowed to clot before centrifugation. After centrifugation at  $4^{\circ}\text{C}$  for 15 minutes at 3,500 rpm, serum was aliquoted and immediately frozen at  $-80^{\circ}\text{C}$  (Sanyo, Japan).

### Determination of ChT concentrations

ChT concentrations were determined by using commercially available ELISA (Sunred, Shanghai, China). The intra-assay and inter-assay confidence interval were  $< 10\%$  and  $< 12\%$ , respectively.

### Determination of ChT activity

ChT enzyme activity was measured based on the method described by Kurt et al. (12) Briefly, 25 mL of serum was incubated with 100 mL of 22 mmol/L 4-methylumbelliferyl-b-D-N,N,N-triacetylchitotriose in McIlvain's phosphate-citrate buffer, pH: 5.2, for 1 hour at  $37^{\circ}\text{C}$ . The reaction was terminated by the addition of 120 mL 0.5 mol/L  $\text{Na}_2\text{CO}_3$ - $\text{NaHCO}_3$  buffer, pH: 10.7, and the fluorescence of 4-methylumbelliferone was measured using a Microfluor 2 fluorimeter (Bio-Tek Instruments, Neufahrn, Germany; excitation: 355 nm, emission: 460 nm). Serum ChT activities were expressed as nanomoles of the substrate hydrolyzed per milliliter per hour (nmol/mL/hr).

### Statistical analysis

Data normality was assessed by the Shapiro-Wilk test. Two-sided independent samples *t*-tests and Mann-Whitney U-tests were used to compare the differences in the continuous and nonparametric variables, respectively. A  $\chi^2$  analysis was used to compare the differences in the categorical variables, and the Pearson method was used to assess the correlation between C-reactive protein (CRP) and the activity and concentrations of chitotriosidase in the participants. Analyses were performed using IBM SPSS software (release 22.0, IBM, SPSS Inc., Chicago, IL, USA).  $P < 0.05$  was considered as statistically significant.

### Ethics statement

Ethics committee approval was received for this study from the institutional review board of Ankara Numune Training and Research Hospital (No. E-15-534). Written informed consent was obtained from all of patients and controls.

## RESULTS

The patients had a median duration of illness of 72 (2-360) months. The baseline characteristics of the study population are given in Table 1. The genotype and frequency of the detected mutations in the patients are provided in Table 2. Median ChT activity was 10.00 (6.00-15.00) nmol/mL/hr and 14.00 (6.25-20.75) nmol/mL/hr in the patients and controls, respectively. A statistically significant difference was found in the ChT activity between the controls and the patients ( $P = 0.027$ ). The median ChT concentrations were 65.40 (46.20-84.92) pg/mL and 125.00 (75.72-143.95) pg/mL in the patients and controls, respectively. There was also a statistically significant difference between the controls and patients in terms of ChT concentrations ( $P < 0.001$ ). The results are expressed as median percentiles (25th-75th). ChT activity, ChT levels and CRP concentrations in controls and patients have been given with boxplot in Fig. 1A, 1B, and 1C, respectively. The mean C-reactive protein (CRP) was  $2.58 \pm 1.83$  mg/dL and  $34.20 \pm 4.5$  mg/dL in patients with an acute attack and an attack-free period, respectively. A statistically sig-

**Table 1.** Baseline characteristics of study population

Characteristics	Patients (n = 80)	Controls (n = 80)	P value
Age, yr	34.86 ± 9.5	35.43 ± 6.44	0.429
Gender (male/female)	37/43	38/42	0.721
<sup>b</sup> ChT activity, nmol/mL/hr	10.00 (6.00-15.00)	14.00 (6.25-20.75)	0.027
<sup>b</sup> ChT concentration, pg/mL	65.40 (46.20-84.92)	125.00 (75.72-143.95)	< 0.001
Creatinin, mg/dL	0.84 ± 0.11	0.87 ± 0.13	0.086
<sup>b</sup> CRP, mg/dL	17.0 (8.25-24.00)	1.50 (1.00-2.00)	< 0.001
WBC, 10 <sup>3</sup> mL	7.93 ± 2.09	7.88 ± 1.37	0.837
Hb, g/dL	13.82 ± 1.84	14 ± 1.04	0.105
Total cholesterol, mg/dL	177.43 ± 43.18	180.25 ± 34.79	0.651
<sup>b</sup> Triglyceride, mg/dL	95.00 (76.00-167.25)	93.00 (74.50-135.25)	0.382
HDL-C, mg/dL	49.63 ± 11.24	51.40 ± 12.26	0.715
LDL-C, mg/dL	104.75 ± 30.63	107.19 ± 33.71	0.635
Disease onset (age)	26.50 ± 11.23		
Family history (yes/no)	51/29		
Fever (yes/no)	69/11		
Abdominal pain (yes/no)	74/6		
Chest pain (yes/no)	37/43		
Erysipeloid (yes/no)	20/60		

Results are expressed as mean ± SD and <sup>b</sup>median (25th-75th percentiles) with 95% confidence intervals.

ChT = chitotriosidase, CRP = C-reactive protein, WBC = white blood cell, Hb = hemoglobin, HDL = high density lipoprotein, LDL = low density lipoprotein.

**Table 2.** Genotype and frequency of detected mutations in patients

Mutation types	Genotypes	Patients	
		No.	%
Homozygote	M694V	22	27.5
	M680I	5	6.25
	A744S	1	1.25
Compound homozygote	M694V/R202Q	3	3.75
Heterozygote	M694V	10	12.5
	E148Q	4	5
	V726A	3	3.75
Compound heterozygote	M694V/V726A	10	12.5
	M680I/M694V	9	11.25
	M694V/E148Q	4	5
	V726A/R761H	4	5
	M694V/R202Q	2	2.5
	E148Q/M680I	2	2.5
	V726A/A744S	1	1.25
Total		80	100

nificant difference was observed in patients with an acute FMF attack compared to those in an attack-free period in terms of C-reactive protein (CRP) ( $P < 0.001$ ). However, we did not find any significant difference in terms of the activity ( $P = 0.136$ ) and concentrations ( $P = 0.331$ ) of ChT between patients with an acute attack and an attack-free period. We also did not find any statistically significant correlation between ChT activity ( $P = 0.978$ ,  $r = 0.003$ ) or concentrations ( $P = 0.446$ ,  $r = -0.87$ ) and C-reactive protein concentrations in the patients.

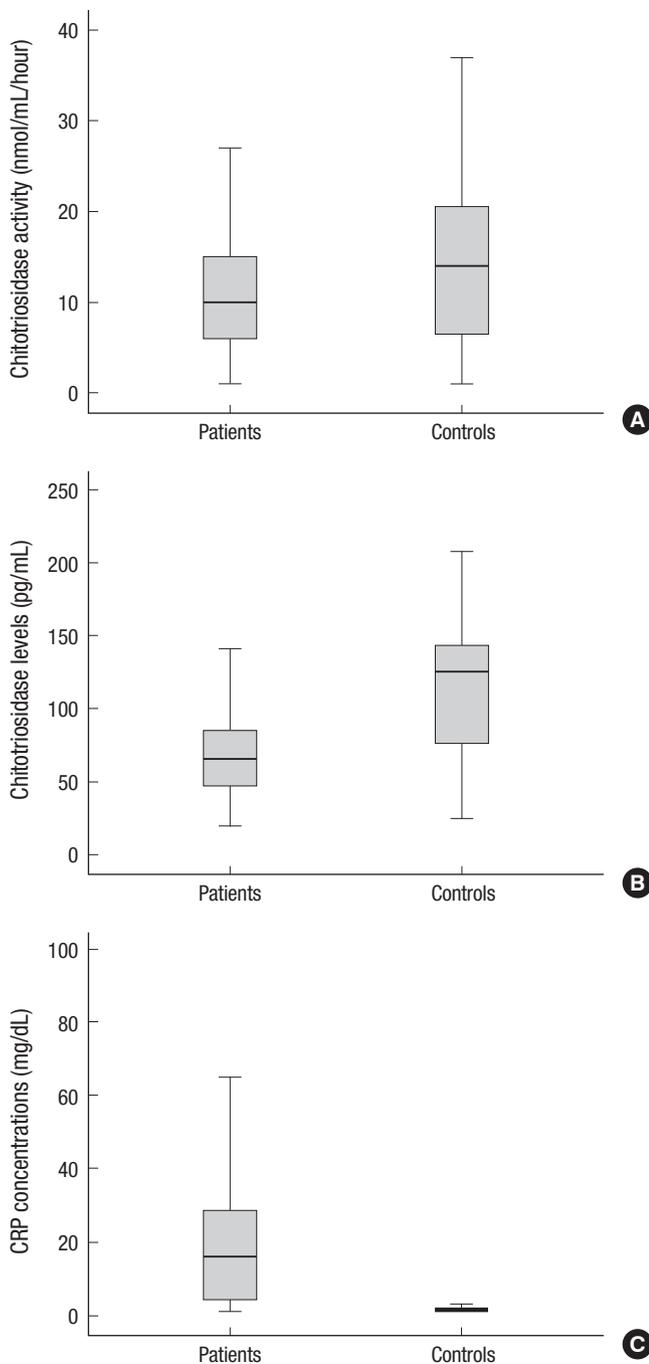
## DISCUSSION

Human ChT is a chitin fragmenting enzyme. Although the physiological functions of this enzyme are still unclear, there is evidence that ChT plays a role in the innate immune system (16-

18). Different studies have demonstrated changes in ChT activity and concentrations in multiple diseases (10-13). However, these changes have not been concurrently evaluated in patients with FMF. FMF is a chronic inflammatory and autosomal recessive disorder. Turkey is one of the countries that has the highest number of patients with FMF. Proteins encoded by Mediterranean fever (*MEFV*) called pyrin and marenostrin are secreted by neutrophils and macrophages. These proteins regulate the secretion of proteins in neutrophils and macrophages. Point mutations (single substitutions) in the *MEFV* gene have an important role in FMF pathogenesis. However, the diagnosis of FMF has been made principally according to clinical findings, and has been frequently applied to the *MEFV* mutation analysis in the case of patients in whom FMF is suspected (19-21).

Taylan et al. (22) demonstrated increased concentrations of ChT in patients with FMF. Contrary to this study, we found lower ChT activity and concentrations in patients with FMF. To date, it has been thought that the main, and even the only, source of serum ChT is activated macrophage (12,23-25). However, it has been demonstrated that polymorphonuclear neutrophils are also a source of ChT (26). Colchicine inhibits the chemotaxis of neutrophils and the release of arachidonate and 5-lipoxygenase in macrophages (27). In our study, 76 patients were receiving colchicine. We thought that the reason for the reduced ChT concentrations and activity in patients with FMF was colchicine use. Colchicine might reduce the expression and activity of ChT by preventing neutrophil leucocytes chemotaxis and macrophage activity.

Familial Mediterranean fever is characterized by periodic attacks that last 1-3 days. The major inflammatory process in acute FMF attacks is the activation of neutrophils in serosal and syno-



**Fig. 1.** Box plots for patients and controls. (A) Chitotriosidase activity. (B) Chitotriosidase levels. (C) CRP levels. The image of each group shows the box with median (horizontal line within the box); the interquartile range (IQR), corresponding to the 25th-75th percentiles (lower and upper limit of the box); nearest observations within 1.5 IQRs (the whiskers) and outliers (circles within 3 IQR).

vial surfaces. It has been determined that CRP is the only acute phase protein raised during the acute attacks of patients with FMF (28). In our study, we found higher CRP concentrations in patients with an acute FMF attack than in those in an attack-free period. However, we did not find any statistically significant differences in the ChT activity and concentrations in patients

with an acute FMF attack compared to those in an attack-free period. Additionally, we did not find any correlation between CRP and the activity and concentration of ChT. Our results are inconsistent with the results of Taylan et al. (22). The discrepancy between results might rely on the great differences in the study population number, the selection criteria for the patients, and differences in the received doses of colchicine. We believe that both ChT activity and concentrations cannot be used to determine and evaluate the acute attack period of FMF.

One of the limitations of the study was that the study population was made up of the same ethnic group, which cannot be generalized to other ethnic groups. Also, we did not adjust our data for body mass index. Another limitation was the lack of data about the patients' ChT polymorphism.

In conclusion, the findings of the present study demonstrate that FMF patients have decreased ChT concentrations and activity compared to healthy controls. Additionally, we did not find any statistically significant correlation between enzyme activity and concentrations and disease activity. Our study is the first report that shows decreasing ChT concentrations and activity in FMF patients. According to our findings, serum ChT enzyme activity and concentrations may not be considered as a biomarker to evaluate disease activity and the prognosis of disease. However, more studies are needed to evaluate the changes in enzyme activity and the role of ChT in patients compared to colchicine negative patients.

## DISCLOSURE

The authors have no potential conflicts of interest to disclose.

## AUTHOR CONTRIBUTION

Conception and design of the study: Doğan HO, Omma A. Performing the experiments or case collection: Doğan HO, Boğdaycıoğlu N, Yavuz H, Demirpençe Ö, Aydın H. Analysis of data: Doğan HO, Turhan T, Karaaslan Y, Bakır S. Writing, revision, final approval: all authors.

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## REFERENCES

- Zadeh N, Getzug T, Grody WW. Diagnosis and management of familial Mediterranean fever: integrating medical genetics in a dedicated interdisciplinary clinic. *Genet Med* 2011; 13: 263-9.
- Shohat M, Halpern GJ. Familial Mediterranean fever--a review. *Genet Med* 2011; 13: 487-98.
- Doğan HO, Koca Y, Erden G, Karaaslan Y, Bozat H. Evaluating MEFV mutation frequency in Turkish familial Mediterranean fever suspected patients and gender correlation: a retrospective study. *Mol Biol Rep* 2012; 39: 6193-6.
- Koo KY, Park SJ, Wang JY, Shin JI, Jeong HJ, Lim BJ, Lee JS. The first case of familial Mediterranean fever associated with renal amyloidosis in Korea. *Yonsei Med J* 2012; 53: 454-8.
- Martinon F, Tschopp J. Inflammatory caspases: linking an intracellular innate immune system to autoinflammatory diseases. *Cell* 2004; 117: 561-74.
- Henderson C, Goldbach-Mansky R. Monogenic autoinflammatory diseases: new insights into clinical aspects and pathogenesis. *Curr Opin Rheumatol* 2010; 22: 567-78.
- Netea MG, Simon A, van de Veerdonk F, Kullberg BJ, Van der Meer JW, Joosten LA. IL-1beta processing in host defense: beyond the inflammasomes. *PLoS Pathog* 2010; 6: e1000661.
- Gasparyan AY, Ayvazyan L, Yessirkepov M, Kitas GD. Colchicine as an anti-inflammatory and cardioprotective agent. *Expert Opin Drug Metab Toxicol* 2015; 11: 1781-94.
- Alanbay I, Ercan CM, Sakinci M, Coksuer H, Ozturk M, Tapan S. A macrophage activation marker chitotriosidase in women with PCOS: does low-grade chronic inflammation in PCOS relate to PCOS itself or obesity? *Arch Gynecol Obstet* 2012; 286: 1065-71.
- Guo Y, He W, Boer AM, Wevers RA, de Bruijn AM, Groener JE, Hollak CE, Aerts JM, Galjaard H, van Diggelen OP. Elevated plasma chitotriosidase activity in various lysosomal storage disorders. *J Inherit Metab Dis* 1995; 18: 717-22.
- Barone R, Di Gregorio F, Romeo MA, Schilirò G, Pavone L. Plasma chitotriosidase activity in patients with beta-thalassemia. *Blood Cells Mol Dis* 1999; 25: 1-8.
- Boot RG, Hollak CE, Verhoek M, Alberts C, Jonkers RE, Aerts JM. Plasma chitotriosidase and CCL18 as surrogate markers for granulomatous macrophages in sarcoidosis. *Clin Chim Acta* 2010; 411: 31-6.
- Kurt YG, Cayci T, Onguru P, Akgul EO, Yaman H, Aydin I, Bodur H, Turker T, Kurt I, Cevik MA, et al. Serum chitotriosidase enzyme activity in patients with Crimean-Congo hemorrhagic fever. *Clin Chem Lab Med* 2009; 47: 1543-7.
- Malaguarnera L. Chitotriosidase: the yin and yang. *Cell Mol Life Sci* 2006; 63: 3018-29.
- Livneh A, Langevitz P, Zemer D, Zaks N, Kees S, Lidar T, Migdal A, Padeh S, Pras M. Criteria for the diagnosis of familial Mediterranean fever. *Arthritis Rheum* 1997; 40: 1879-85.
- Wajner A, Michelin K, Burin MG, Pires RF, Pereira ML, Giugliani R, Coelho JC. Biochemical characterization of chitotriosidase enzyme: comparison between normal individuals and patients with Gaucher and with Niemann-Pick diseases. *Clin Biochem* 2004; 37: 893-7.
- van Eijk M, van Roomen CP, Renkema GH, Bussink AP, Andrews L, Blommaert EF, Sugar A, Verhoeven AJ, Boot RG, Aerts JM. Characterization of human phagocyte-derived chitotriosidase, a component of innate immunity. *Int Immunol* 2005; 17: 1505-12.
- Labadaridis I, Dimitriou E, Theodorakis M, Kafalidis G, Velegraki A, Michelakakis H. Chitotriosidase in neonates with fungal and bacterial infections. *Arch Dis Child Fetal Neonatal Ed* 2005; 90: F531-2.
- Stehlik C, Reed JC. The PYRIN connection: novel players in innate immunity and inflammation. *J Exp Med* 2004; 200: 551-8.
- Liepinsh E, Barbals R, Dahl E, Sharipo A, Staub E, Otting G. The death-domain fold of the ASC PYRIN domain, presenting a basis for PYRIN/PYRIN recognition. *J Mol Biol* 2003; 332: 1155-63.
- Pras M. Familial Mediterranean fever: from the clinical syndrome to the cloning of the pyrin gene. *Scand J Rheumatol* 1998; 27: 92-7.
- Taylan A, Gurler O, Toprak B, Sisman AR, Yalcin H, Colak A, Sari I. S100A12, chitotriosidase, and resolvin D1 as potential biomarkers of familial Mediterranean fever. *J Korean Med Sci* 2015; 30: 1241-5.
- Renkema GH, Boot RG, Strijland A, Donker-Koopman WE, van den Berg M, Muijsers AO, Aerts JM. Synthesis, sorting, and processing into distinct isoforms of human macrophage chitotriosidase. *Eur J Biochem* 1997; 244: 279-85.
- Boot RG, Renkema GH, Strijland A, van Zonneveld AJ, Aerts JM. Cloning of a cDNA encoding chitotriosidase, a human chitinase produced by macrophages. *J Biol Chem* 1995; 270: 26252-6.
- Hollak CE, van Weely S, van Oers MH, Aerts JM. Marked elevation of plasma chitotriosidase activity. A novel hallmark of Gaucher disease. *J Clin Invest* 1994; 93: 1288-92.
- Czartoryska B, Fiszer U, Lugowska A. Chitotriosidase activity in cerebrospinal fluid as a marker of inflammatory processes in neurological diseases. *J Lab Med* 2001; 25: 77-81.
- Grattagliano I, Bonfrate L, Ruggiero V, Scaccianoce G, Palasciano G, Portincasa P. Novel therapeutics for the treatment of familial Mediterranean fever: from colchicine to biologics. *Clin Pharmacol Ther* 2014; 95: 89-97.
- Korkmaz C, Ozdogan H, Kasapçopur O, Yazici H. Acute phase response in familial Mediterranean fever. *Ann Rheum Dis* 2002; 61: 79-81.