

# Suppression of CFTR-mediated Cl<sup>-</sup> Secretion of Airway Epithelium in Vitamin C-deficient Mice

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## INTRODUCTION

Hyperoxic inhalation (PO<sub>2</sub> higher than that in the ambient air) is used in a variety of clinical practices including anaesthesia and post-operative recovery. The duration of hyperoxic ventilation can be prolonged depending on the patient conditions such as peripheral O<sub>2</sub> saturation. Also, in patients with respiratory disorders which cause hypoxemia, long-term oxygen therapy is often used. Although the benefits of O<sub>2</sub> supplementation are obvious in clinical situations of fatal hypoxemia, there also are harmful effects of O<sub>2</sub>. For example, the side effects of increasing concentrations of O<sub>2</sub> supplementation are frequently observed as pulmonary injury (1). In neonatal neural tissues such as the retina, the harmful influence of hyperoxic ventilation has also been observed (2). The cough reflex, a defensive respiratory reflex, is also impaired in hyperoxia, and the inhibition of cough reflex is prevented by dietary antioxidants (3).

It is generally thought that tissue injury caused by O<sub>2</sub> is mediated by the formation of reactive oxygen species (ROS), which can react with and damage essential biomolecules via lipid peroxidation, protein sulfhydryl oxidation, and DNA damage (4). Because airway epithelium is constantly and frequently ex-

posed to oxidative stress, it is highly likely that ROS-mediated oxidative stress affects the functions of airway epithelium (5-7). In this context, there is increasing evidence for the protective effects of antioxidant supplementation in respiratory diseases (3, 4, 8). Because humans cannot synthesize ascorbic acid, dietary uptake of vitamin C is essential to cope with oxidative stress and to preserve physiological homeostasis. It has been reported that vitamin C is present in airway surface liquid (ASL), a thin (10-30 μm) layer of fluid covering the luminal surface of the airway epithelium (9-11). The physiological role of vitamin C in ASL is to stimulate Cl<sup>-</sup> secretion via cAMP-activated Cl<sup>-</sup> channels known as cystic fibrosis transmembrane transport regulator (CFTR) in the luminal membrane of the airway epithelium (12). A balanced level of ASL is critical for the protection of the airway epithelium. For transepithelial fluid secretion, an electrogenic Cl<sup>-</sup> secretion model is regarded as the ionic mechanism in which the cAMP-dependent activations of the luminal Cl<sup>-</sup> channel (CFTR) and the basolateral K<sup>+</sup> channel (potassium voltage-gated channel, KQT-like subfamily, member 1, KCNQ1) are critical steps (11, 13).

**Key Words:** Hyperoxia; Airway Epithelium; Cystic Fibrosis Transmembrane Conductance Regulator; Electrolyte Transport; Ascorbic Acid

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While there have been numerous studies on the structural

and biochemical changes in respiratory epithelial cells in response to oxidative stress, a direct investigation on the physiological function (i.e., electrolyte secretion) has been rarely conducted. The studies by Cowley and Linsdell showed that exogenous hydrogen peroxide ( $\text{H}_2\text{O}_2$ , 0.5-2 mM) directly activates the electrogenic  $\text{Cl}^-$  secretion of Calu-3, a cell line model of serous airway epithelial cells (14). In contrast, the oxidative stress caused by pyocyanin, a redox-active phenazine compound, impairs CFTR-dependent  $\text{Cl}^-$  secretion in the bronchial epithelium (15). Consistent with this, vitamin C activates CFTR in primary cultured human airway epithelial cells (12).

Apart from the acute effects, chronic effects of oxidative stress and of vitamin C deprivation on the airway electrolyte transport are very important. To the best of our knowledge, there has been no investigation on the effects of sustained hyperoxic ventilation on the electrolyte secretion of airway epithelium in vivo. Also, the functional changes in the airway epithelium in the vitamin C-deficient animal model have not yet been investigated.

Unlike humans, rodents synthesize vitamin C (ascorbic acid) from glucose in situ. Recently, a mouse line has been generated with a targeted deletion of the gene coding for L-gulonolactone oxidase (Gulo), which catalyzes the final step of ascorbic acid biosynthesis (16). Mice null for Gulo (gulo $^{-/-}$ ) mice and that are not provided with dietary vitamin C supplements become scorbutic, lose weight, and eventually die. The ambient level of oxygen might induce oxidative stress when the intrinsic antioxidant levels are insufficient. In this respect, it was tempting to investigate the airway epithelial secretory function in gulo $^{-/-}$  mice exposed to normal atmospheric conditions.

Based on the results outlined above, we investigated electrolyte secretion and the absorption functions of mouse airway epithelium using the Ussing chamber apparatus to measure short-circuit current ( $I_{sc}$ ). The changes in  $I_{sc}$  ( $\Delta I_{sc}$ ) reflecting the  $\text{Cl}^-$  secretion and  $\text{Na}^+$  absorption functions of airway epithelia were compared to mice exposed to hyperoxic conditions (80%-98%  $\text{PO}_2$ ) and to a normoxic environment. Also, we compared the  $I_{sc}$  values in the airway epithelium of gulo $^{-/-}$  mice according to the duration of vitamin C deficiency.

## MATERIALS AND METHODS

### Animals

ICR mice, C57BL/6 wild-type mice and gulo $^{-/-}$  mice were maintained in a specific pathogen-free condition in the animal facility at Seoul National University College of Medicine. Gulo $^{-/-}$  mice were maintained with or without 3.3 g/L of vitamin C supplementation in the drinking water. The effects of chronic hyperoxia were tested in ICR mice (Fig. 1). For chronic exposure to hyperoxic conditions (80%-98%  $\text{PO}_2$ ), mice were kept in a semi-tight transparent chamber for 24-28 hr with automatic reg-

ulation of  $\text{PO}_2$  (Biospherix, Lacona, NY, USA). Control mice were kept in the same cage but with a half-open door, i.e., ambient air exposure. For the investigation of airway epithelial function in gulo $^{-/-}$  mice and the immunohistochemistry study, C57BL/6 mice were used as the control group.

### Ussing chamber experiments

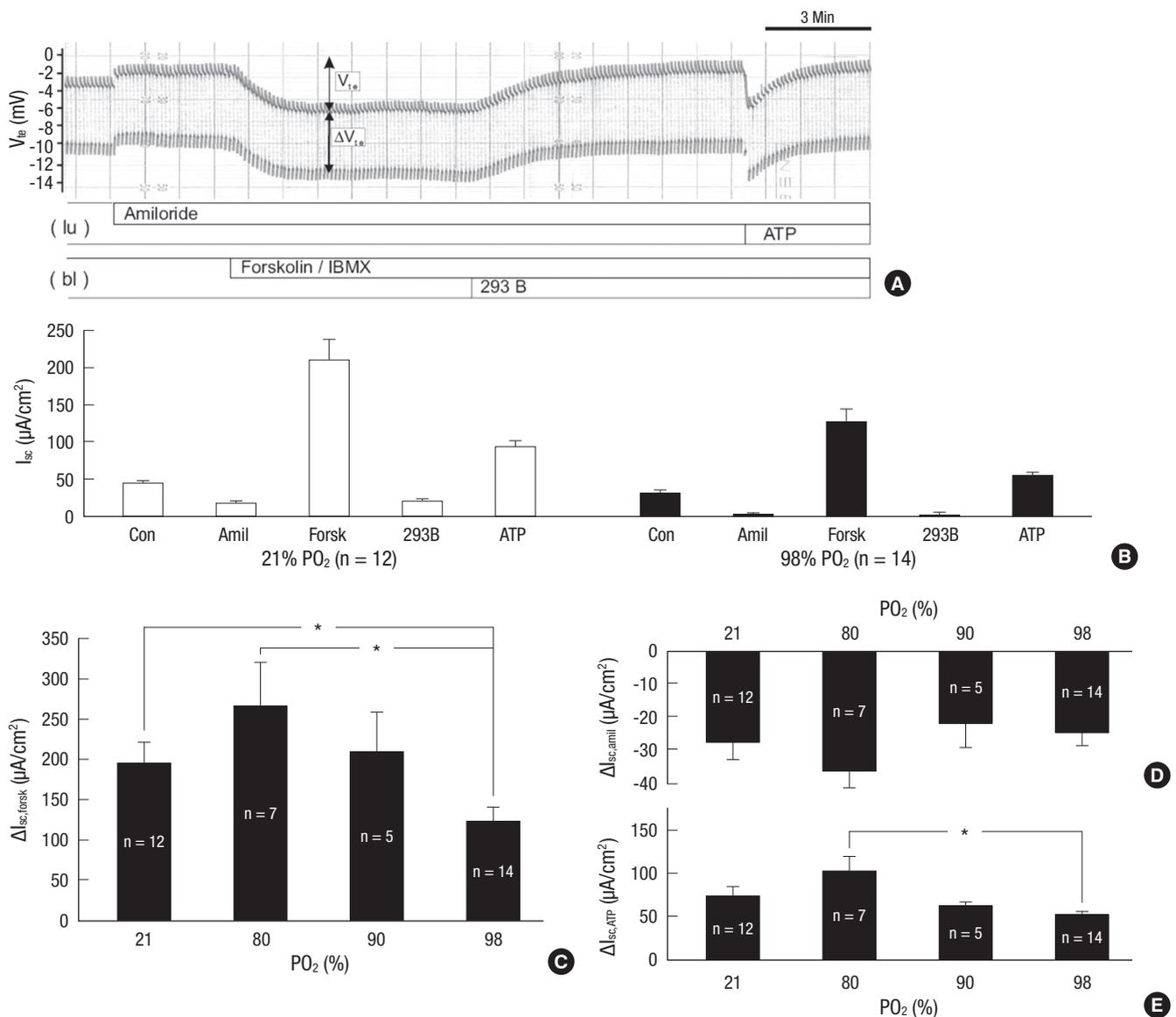
Mice of both genders (body weight, 25-35 g) were sacrificed via inhalation of 100%  $\text{CO}_2$ . The trachea was split along the anterior side, and the pars membranacea of the trachea was mounted into a tissue holder in the Ussing chamber (circular exposed area, 0.64  $\text{mm}^2$ ) with the aid of a dissection microscope. The chamber (2 mL) was maintained at 37°C and continuously perfused with a normal Tyrode's (NT) solution containing (in mM) 145 NaCl, 0.4  $\text{KH}_2\text{PO}_4$ , 1.6  $\text{K}_2\text{HPO}_4$ , 5 HEPES, 5 D-glucose, 1  $\text{MgCl}_2$ , 1.3  $\text{CaCl}_2$  (pH 7.4) on both sides at a flow rate of 10-15 mL/min (For more detail of the instrument, please see supplementary photos in online-only material). Indomethacin (1  $\mu\text{M}$ ) was included in all experimental solutions to inhibit the endogenous formation of prostaglandins. The tissue was allowed to equilibrate for at least 30 min prior to the experiments. Trans-epithelial resistance ( $R_{te}$ ) was determined from the voltage deflection ( $\Delta V_{te}$ ) caused by the injection of current ( $I_{inj}$ , 0.8  $\mu\text{A}$ , 1.4 sec of duration, 0.7 Hz) according to Ohm's law ( $R_{te} = \Delta V_{te}/I_{inj}$ ). The resistance of the empty chamber was subtracted. The equivalent short circuit current ( $I_{sc}$ ) was calculated based on the trans-epithelial voltage ( $V_{te}$ ) and  $R_{te}$  according to Ohm's law ( $I_{sc} = V_{te}/R_{te}$ ). The electrical signs of  $I_{sc}$  and  $V_{te}$  refer to the luminal side. Amiloride, indomethacin, forskolin, and 293B were initially dissolved in dimethylsulfoxide (DMSO) and diluted with NT solution. The final concentration of DMSO was < 0.1%. All the chemicals used in the Ussing chamber study were purchased from Sigma-Korea (Seoul, Korea).

### Immunofluorescence microscopy

Mice were perfused with heparinized PBS, and the main trachea tissues were fixed in 4% paraformaldehyde at 4°C. Frozen sections with 5  $\mu\text{m}$  thicknesses were post-fixed, and non-specific signals were blocked with 5% normal serum. Cut tissues were incubated with anti-CFTR antibody or anti-KCNQ1 antibody (Abcam, Cambridge, MA, USA) at 4°C overnight in a humidified chamber and then incubated with AlexaFluor 555-conjugated secondary antibody (Invitrogen, Camarillo, CA, USA) for 1 hr at room temperature. Nuclei were counterstained with 4',6-diamidino-2-phenylindole (DAPI) and observed with confocal fluorescence microscopy (IX-81, Olympus, Japan) using image software (Fluoview 1000, Olympus, Tokyo, Japan).

### Statistics

The data is presented as the representative original recordings and graphs of the mean  $\pm$  SEM. For statistical analysis, ANOVA



**Fig. 1.** Ussing chamber experiments in tracheal epithelia obtained from control and hyperoxia-exposed mice. **(A)** Original recordings of the transepithelial voltage ( $V_{te}$ ). The upper border of the trace is  $V_{te}$ , the downward deflection ( $\Delta V_{te}$ ) is the response to current injections from which the transepithelial resistance ( $R_{te}$ ) and the equivalent short-circuit current ( $I_{sc}$ ) were calculated. The bars below indicate the luminal (lu) or basolateral (bl) application of amiloride (10  $\mu M$ ), forskolin (2  $\mu M$ )/IBMX (100  $\mu M$ ), chromanol 293B (293B 10  $\mu M$ ) or ATP (50  $\mu M$ ). **(B)** Summary of  $I_{sc}$  measured in the initial control and during each phase of drug application as demonstrated above. Open and closed bar graphs indicate results from normoxia (21% PO<sub>2</sub>) and hyperoxia (98% PO<sub>2</sub>, 36 hr)-exposed mice, respectively. **(C-E)** Summaries of the changes in  $I_{sc}$  ( $\Delta I_{sc}$ ) caused by forskolin/IBMX **(C)**, amiloride **(D)**, and ATP **(E)**, as demonstrated in the above trace. Data from the groups of mice exposed to different levels of oxygen tension (21%, 80%, 90%, and 98% PO<sub>2</sub>) are compared. Numbers of tested tissues are directly indicated in the figure. The asterisks indicate statistical significance ( $P < 0.05$ , paired t-test).

followed by a post hoc t-test was applied (Figs. 1C, 2C, D). Unpaired t-test was applied to the corresponding data between control and test groups in Figs. 1B and 2B, and a  $P$  value  $< 0.05$  was considered statistically significant.

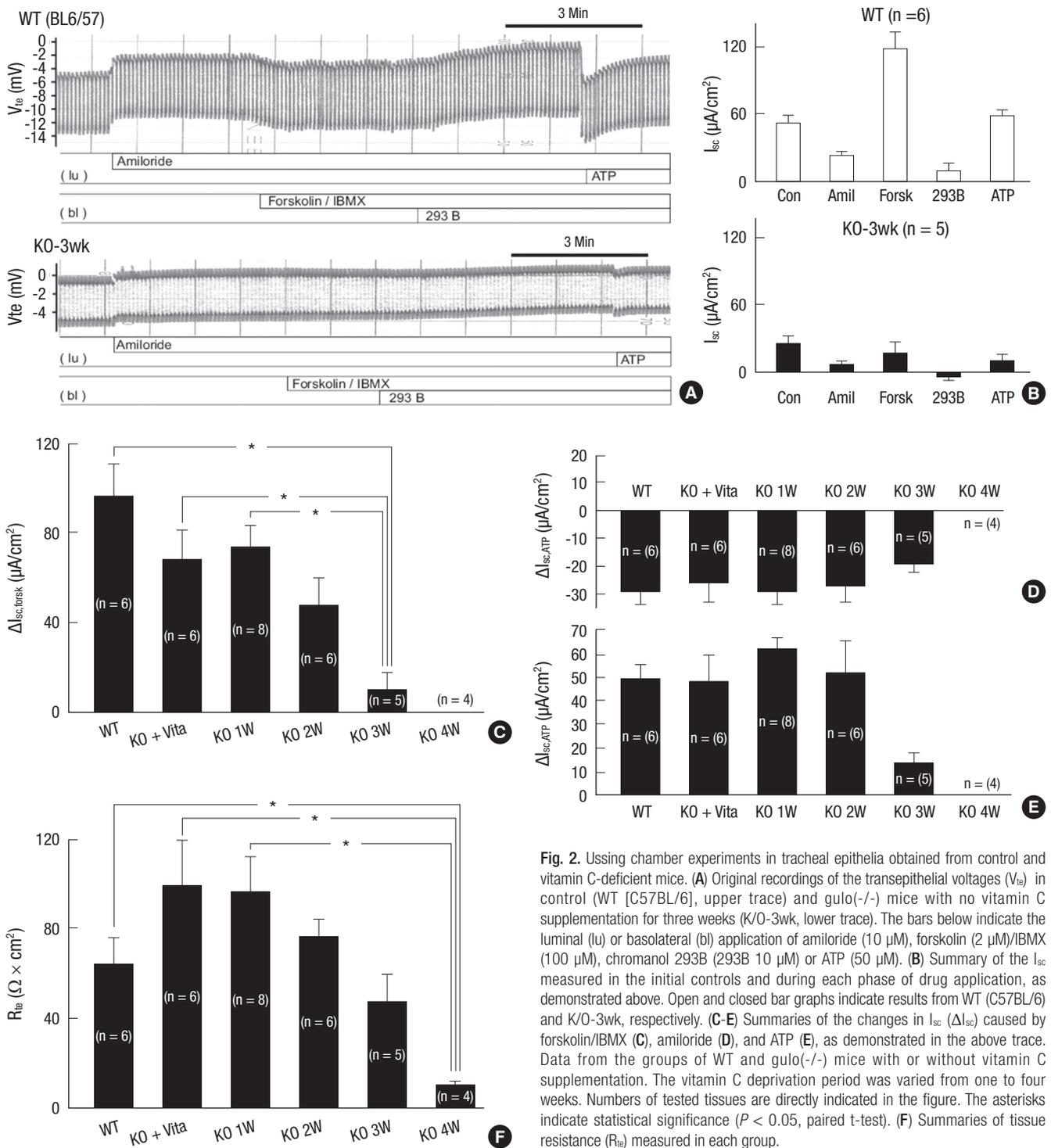
#### Ethics statement

All study protocols were in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996) and also conformed to Seoul National University College of Medicine guidelines for the care and use of animals. The ani-

mal protocol for experiments was reviewed and approved by the IACUC committee of the Seoul National University (SNU-090925-2).

## RESULTS

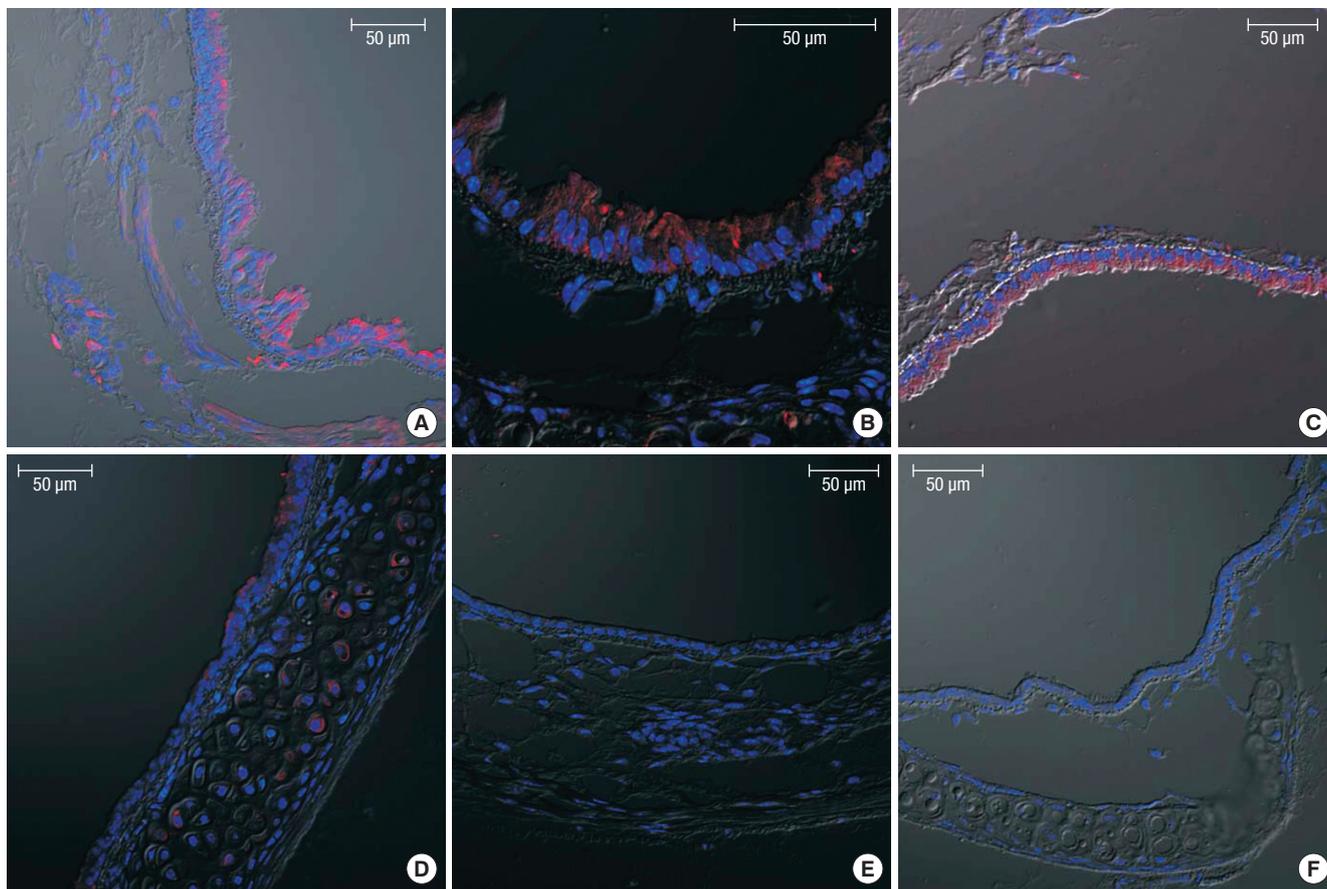
An original trace of the Ussing chamber experiments with an ICR mouse is shown in Fig. 1A, in which the  $V_{te}$  and  $\Delta V_{te}$  are directly indicated. After the equilibrium period in NT solution, a stable level of  $V_{te}$  was confirmed. The initial spontaneous  $V_{te}$  was reduced by the luminal application of amiloride (10  $\mu M$ ),



**Fig. 2.** Ussing chamber experiments in tracheal epithelia obtained from control and vitamin C-deficient mice. **(A)** Original recordings of the transepithelial voltages ( $V_{te}$ ) in control (WT [C57BL/6], upper trace) and *gulo*<sup>-/-</sup> mice with no vitamin C supplementation for three weeks (K/O-3wk, lower trace). The bars below indicate the luminal (lu) or basolateral (bl) application of amiloride (10  $\mu$ M), forskolin (2  $\mu$ M)/IBMX (100  $\mu$ M), chromanol 293B (293B 10  $\mu$ M) or ATP (50  $\mu$ M). **(B)** Summary of the  $I_{sc}$  measured in the initial controls and during each phase of drug application, as demonstrated above. Open and closed bar graphs indicate results from WT (C57BL/6) and K/O-3wk, respectively. **(C-E)** Summaries of the changes in  $I_{sc}$  ( $\Delta I_{sc}$ ) caused by forskolin/IBMX **(C)**, amiloride **(D)**, and ATP **(E)**, as demonstrated in the above trace. Data from the groups of WT and *gulo*<sup>-/-</sup> mice with or without vitamin C supplementation. The vitamin C deprivation period was varied from one to four weeks. Numbers of tested tissues are directly indicated in the figure. The asterisks indicate statistical significance ( $P < 0.05$ , paired t-test). **(F)** Summaries of tissue resistance ( $R_{te}$ ) measured in each group.

which blocks the epithelial  $Na^+$  channels (ENaC). In the presence of amiloride, cAMP-dependent  $Cl^-$  secretion was induced via application of forskolin (2  $\mu$ M) and isobutylmethyl xanthine (IBMX, 100  $\mu$ M) to the basolateral side. The addition of forskolin/IBMX induced a negative shift in  $V_{te}$  and increased the lumen negative  $I_{sc}$ . It is well known that luminal  $Cl^-$  channels (CFTR) and basolateral  $K^+$  channels (KCNQ1) are activated by cAMP-dependent signalling pathways (17, 18). Consistent with

this model, the application of the KCNQ1 channel blocker (100  $\mu$ M of chromanol 293B) to the basolateral side suppressed the cAMP-induced  $I_{sc}$ . Then, ATP (50  $\mu$ M) was applied to the luminal side of the tracheal epithelium, which is known to induce a transient increase in cytosolic  $Ca^{2+}$  concentration ( $[Ca^{2+}]_c$ ) via phospholipase C (PLC)/inositol-trisphosphate ( $InsP_3$ )-coupled signalling pathways inducing the release of stored  $Ca^{2+}$  from the endoplasmic reticulum (ER) (19). This luminal ATP-induced



**Fig. 3.** Decreased expression of CFTR in the tracheal epithelia from vitamin C-deprived mice. Using confocal microscopy, the expression of CFTR (red) was examined with immunostaining in the tracheal epithelia of wild type (A), vitamin C-supplemented *gulo*<sup>-/-</sup> (B), and vitamin C-deprived *gulo*<sup>-/-</sup> mice (C-F). Nuclei were counterstained with DAPI (blue). The expressions of CFTR in luminal membranes decreased in K/O-2wk (D) and almost disappeared in K/O-3wk mice (E). Note that the columnar shape of the epithelium changes to cube-like or flattened in K/O-4wk (F). Scale bar, 50  $\mu$ m.

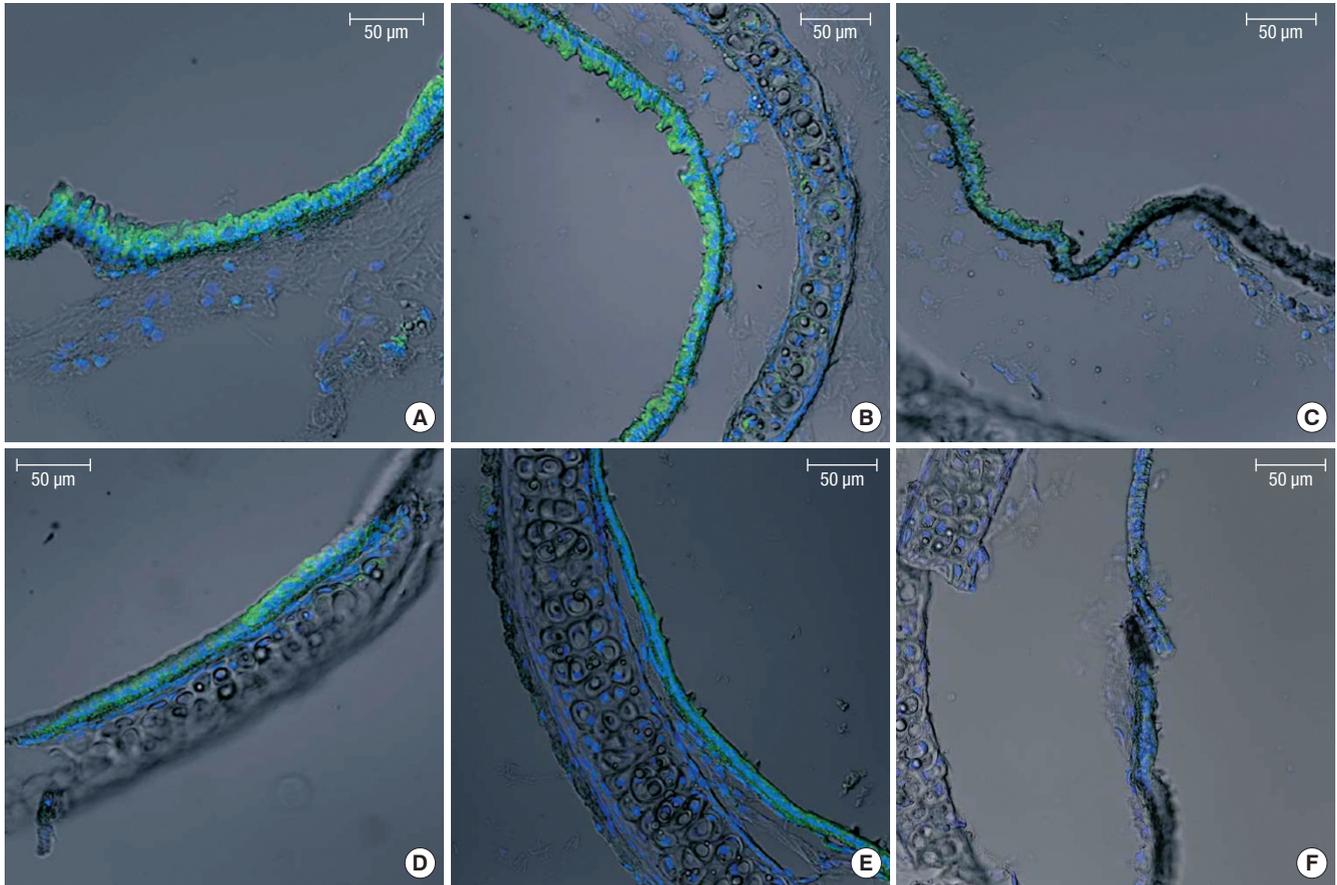
increase in  $[Ca^{2+}]_c$  ( $\Delta[Ca^{2+}]_c$ ) is known to induce  $Cl^-$  secretion via  $Ca^{2+}$ -activated  $Cl^-$  channels ( $Cl_{Ca}$ ) in the luminal membrane. Consistently, transient increases in  $V_{te}$  and  $I_{sc}$  were observed in response to the luminal ATP application (19, 20). The  $I_{sc}$  values measured in each phase of the above protocol showed a decreasing tendency in the mice exposed to strong hyperoxic conditions ( $PO_2$ , 98%) for 36 hr (Fig. 1B).

The average values of  $I_{sc}$  change ( $\Delta I_{sc}$ ) induced by the above pharmacological treatments are summarized in Fig. 1C. In mice that were in hyperoxia conditions (80%, 90%, and 98% of  $PO_2$  for 36 hr), the peak amplitude of  $\Delta I_{sc}$  induced by forskolin/IBMX ( $\Delta I_{sc, forsk}$ ) decreased at the highest  $PO_2$  condition compared to the responses of the mice exposed to 80%  $PO_2$  or to the normoxic condition (Fig. 1C). The peak increase in  $I_{sc}$  due to the luminal ATP ( $\Delta I_{sc, ATP}$ ) of 98%  $PO_2$  was smaller than that of 80%  $PO_2$  and not significantly different from that of the control (Fig. 1D). In contrast, the amiloride-sensitive  $I_{sc}$  ( $\Delta I_{sc, amil}$ ) was not affected by chronic hyperoxia (Fig. 1E).

Next, we investigated the  $I_{sc}$  responses of airway epithelia obtained from wild-type C57BL/6 mice (WT), *gulo*<sup>-/-</sup> mice with vitamin C-supplementation for three weeks (K/O+Vit-C), and

*gulo*<sup>-/-</sup> mice reared with no vitamin C-supplementation for 5-7 days (K/O-1wk), 14-15 days (K/O-2wk), 20-22 days (K/O-3wk), and 27-29 days (K/O-4wk). In Fig. 2A, the representative traces of  $V_{te}$  and  $\Delta V_{te}$  are shown for WT and K/O-3wk mice. Also, the averaged  $I_{sc}$  values measured in each phase of the above protocol (control, amiloride, forskolin/IBMX, 293B, and ATP) are summarized for the WT and K/O-3wk mice (Fig. 2B). In general, the responses of airway epithelial  $I_{sc}$  to the above pharmacological agents were smaller in WT mice than those in the ICR mice described above (Fig. 1B). Nevertheless, it was notable that the amplitudes of  $I_{sc}$  were markedly suppressed in K/O-3wk mice (Fig. 2B).

The amplitudes of  $\Delta I_{sc, forsk}$  were compared between the groups of different duration of vitamin C deprivation. Overall,  $\Delta I_{sc, forsk}$  showed a decreasing tendency beginning with K/O-2wk mice, then became significant in K/O-3wk mice, and was completely abolished in K/O-4wk mice (Fig. 2C).  $\Delta I_{sc, ATP}$  was also decreased in the K/O-3wk and K/O-4wk mice (Fig. 2D). In contrast,  $\Delta I_{sc, amil}$  was abolished only in the K/O-4wk group (Fig. 1E). We also noted that the tissue resistance ( $R_{te}$ ) showed a decreasing tendency in K/O-3wk mice and was significantly de-



**Fig. 4.** Persistent expression of KCNQ1 in the tracheal epithelia from vitamin C-deprived mice. Immunofluorescence microscopy was used to compare the expressions of KCNQ1 (green) in WT (A) and *gulo*(-/-) mice in which vitamin C was either supplemented (B) or not for 1, 2, 3, and 4 weeks (C-F). Nuclei were counterstained with DAPI (blue). Scale bar, 50  $\mu$ m.

creased in K/O-4wk mice (Fig. 2F).

The above results suggest that the  $\text{Cl}^-$  secretory function of the mouse airway epithelium is susceptible to ambient levels of oxidative stress when the endogenous antioxidant is deficient. Because the cAMP-activated CFTR is the major pathway of  $\text{Cl}^-$  secretion (11, 13), we compared the expression of CFTR in airway epithelia between WT and *gulo*(-/-) mice. Strong expression of CFTR in the luminal membrane was commonly observed in the control and K/O+Vit-C mice. However, consistent with the decrease in  $\Delta I_{\text{sc, forsk}}$  according to the extent of vitamin C deficiency, the expression of CFTR was decreased in K/O-2wk mice and was nearly absent in K/O-3wk and K/O-4wk mice (Fig. 3).

As seen from the inhibition of  $\Delta I_{\text{sc, forsk}}$  by 293B, KCNQ1 activity is also critical to the maintenance of  $\text{Cl}^-$  secretion through CFTR (18). Therefore, we investigated whether KCNQ1 expression is altered in *gulo*(-/-) mice. However, the expression of KCNQ1 was persistently observed in K/O-3wk and K/O-4wk mice (Fig. 4). While not rigorously analyzed here, morphological changes were also observed in the airway epithelia of *gulo*(-/-) mice; the typical ciliated columnar epithelium became cuboid or flattened in response to increased duration of vita-

min C deficiency (Figs. 3, 4, K/O-3wk and K/O-4wk).

## DISCUSSION

In the present study, we found that chronic vitamin C deficiency suppressed  $\text{Cl}^-$  secretion and downregulated CFTR expression in mouse airway epithelium. The cAMP-dependent  $\text{Cl}^-$  secretion ( $\Delta I_{\text{sc, forsk}}$ ) as well as the luminal ATP-induced  $\text{Cl}^-$  secretion ( $\Delta I_{\text{sc, ATP}}$ ) was decreased in *gulo*(-/-) mice. Since the CFTR was fully stimulated by forskolin/IBMX upon ATP application in the present experimental procedure,  $\Delta I_{\text{sc, ATP}}$  was believed to be due to  $\text{Cl}^-$  secretion via CFTR as well as  $\text{Ca}^{2+}$ -activated  $\text{Cl}^-$  channels (19). In this respect, the decreased  $\Delta I_{\text{sc, ATP}}$  might also reflect the downregulation of CFTR. However, we could not conclude that the ion channels other than CFTR were actually affected by hyperoxia or vitamin C deficiency in mice. The inhibitory tendency of chronic hyperoxia on  $\Delta I_{\text{sc, Fsk}}$  also suggested that an imbalance between oxidizing influence and antioxidant capacity could impair the cAMP-dependent  $\text{Cl}^-$  secretion mechanisms (Fig. 1C).

While CFTR-mediated secretion is more susceptible to vitamin C deficiency than it is to other parameters, the sustained

deprivation of vitamin C generally suppresses the transepithelial transport functions. This functional impairment seems to precede the morphological changes in the epithelia of *gulo(-/-)* mice. Notably, the parameter reflecting the integrity of tissue ( $R_{te}$ ) is severely lowered in K/O-4wk mice, indicating that it may be one of the scorbutic symptoms. It is well known that chronic deficiency of vitamin C weakens connective tissue due to impaired collagen synthesis (16). In addition to the decreased  $R_{te}$ , the histological findings of the K/O-3 and -4wk (flattened airway epithelium) mice indicate that a transformation from a ciliated/columnar epithelium occurs during the sustained loss of vitamin C in vivo.

The human respiratory tract is constantly exposed to transient instances of oxidative stress resulting from the inhalation of a variety of foreign materials including atmospheric pollutants and microorganisms. Furthermore, the production of ROS during episodes of infection and inflammation has been implicated in the pathogenesis of a number of pulmonary diseases such as asthma, adult respiratory distress syndrome, chronic obstructive pulmonary disease, and cystic fibrosis (21-24). Since inflammation and oxidative stress are closely related, inflammation also plays a role in the development of chronic lung disease (21, 22). Clinically, oxygen therapy is frequently applied to treat systemic hypoxemia. In this process, harmful effects of hyperoxia are often reported. The risk of hyperoxic stress and its sequel (e.g., bronchopulmonary dysplasia) are more prominently observed in neonates (2, 25).

The concomitant suppressions of  $\Delta I_{sc, forsk}$  and CFTR expression in vitamin C-deprived strongly suggest that chronic oxidative stress negatively regulates CFTR protein expression. Recently, it has been reported that oxidative stress induced by the pharmacological agent tert-butylhydroquinone (BHQ) suppresses CFTR expression in T84, a colonic epithelial cell line (6). The same group also reported that the functional expression of CFTR was suppressed in the human nasal mucosa of cigarette smokers (5). Based on the results of these previous studies, it is highly likely that the suppressions of  $\Delta I_{sc, forsk}$  and CFTR in *gulo(-/-)* mice reflect a transcriptional regulation of CFTR genes under chronic oxidative stress (26, 27). Interestingly, CF-like symptoms such as thickened airway secretions are often seen in chronic inflammatory airway diseases that are not associated with mutations in the CFTR gene, and there is emerging evidence that posttranslational damage to CFTR by reactive oxygen and nitrogen species decreases CFTR function (27).

Antioxidants such as vitamin C may have beneficial effects via restoration of the balanced levels of ROS in tissues, as has been suggested in a study of hepatic ischemic-reperfusion injury model (28). Also, the lack of vitamin C in the ASL of asthmatics has been reported (9). However, careful interpretation is still required for the role of dietary supplementation of vitamin C in human, since there are controversies in the relation between

plasma levels of vitamin C and cancer incidence (29). Furthermore, because we have not directly measured the oxidative stress in this study, the downregulation of CFTR by vitamin C deprivation might owe to the lack of vitamin C per se in addition to the putative oxidative stress.

In summary, this study confirms the inhibitory effects of vitamin C deprivation on the  $Cl^-$  secretion function in murine airway epithelium in vivo. Among the ion channels associated with epithelial  $Cl^-$  secretion, CFTR appears more to be vulnerable to oxidative stress than are other types of channels.

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## AUTHOR SUMMARY

### Suppression of CFTR-mediated Cl<sup>-</sup> Secretion of Airway Epithelium in Vitamin C-deficient Mice

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During surgery or post-operative care, prolonged high oxygen (hyperoxia) supply might damage respiratory system, which is mediated by oxidative stress. Airway epithelium is covered by a thin fluid that is maintained by cooperative actions of membrane ion channels such as cystic fibrosis conductance regulator (CFTR) and K<sup>+</sup> channel (KCNQ1). The CFTR activity of mouse airway epithelia was attenuated when exposed to 98% oxygen for a prolonged period (> 36 hr). Vitamin C, an antioxidant intrinsically produced in mice, might play a protective role. In fact, the CFTR activity in the vitamin C-depleted scurvy mice (gulo<sup>-/-</sup> mice) was decreased in normal ambient air. An immunofluorescence study confirmed the decreased expression of CFTR in gulo<sup>-/-</sup> mice whereas KCNQ1 was preserved. The CFTR-dependent airway surface fluid is susceptible to oxidative stress, which might suggest the benefit of vitamin C supplementation.