

Assessment of sterilization effect and the alteration of surface texture and physical properties of gutta-percha cone after short-term chemical disinfection

Nan-Sim Pang¹, Il-Young Jung¹, Yoon-Jung Yu², Kee-Yeon Kum^{3*}

¹Department of Conservative Dentistry, ²Department of Oral Biology,
Oral Science Research Center, Yonsei University, ³Department of Conservative Dentistry,
³Dental Research Institute, Seoul National University

ABSTRACT

The purposes of this study were firstly to identify the microbial species on gutta-percha (GP) cones exposed at clinics using polymerase chain reaction, and secondly to evaluate the short-term sterilization effect of three chemical disinfectants. It also evaluated the alteration of surface texture and physical properties of GP cones after 5-min soaking into three chemical disinfectants. 150 GP cones from two endodontic departments were randomly selected for microbial detection using PCR assay with universal primer. After inoculation on the sterilized GP cones with the same microorganism identified by PCR assay, they were soaked in three chemical disinfectants: 5% NaOCl, 2% Chlorhexidine, and Chloraprep for 1, 5, 10, and 30 minutes. The sterilization effect was evaluated by turbidity and subculture. The change of surface textures using a scanning electron microscope and the tensile strength and elongation rate of the GP cones were measured using an Instron 5500 (Canton). Statistical analysis was performed.

Four bacterial species were detected in 29 GP cones (19.4%), and all the species belonged to the genus *Staphylococcus*. All chemical disinfectants were effective in sterilization with just 1 minute soaking. On the SEM picture of NaOCl-soaked GP cone, a cluster of cuboidal crystals was seen on the cone surface. The tensile strength of NaOCl-soaked group was significantly higher than the other groups ($p < 0.05$). Also, all disinfectants significantly increased the elongation rate of GP cones compared to the fresh GP cone ($p < 0.05$). Present data demonstrate that three chemical disinfectants are useful for rapid sterilization of GP cone just before obturation. [J Kor Acad Cons Dent 31(2):133-140, 2006]

Key words : PCR, Surface texture, Physical properties, Sterilization effect, Gutta-percha cone

- Received 2006.1.11., revised 2006.1.14., accepted 2006.1.20. -

I . INTRODUCTION

Root canal filling is an important procedure in endodontic treatment to prevent root canal re-infection for successful endodontic prognosis. Care must be taken during this procedure to avoid root canal cross-infection by instruments or filling

* Corresponding Author: **Kee-Yeon Kum**

Dept. of Conservative Dentistry, Dental Research Institute,
College of Dentistry, Seoul National University
28-2 Yeongun-dong, Chongro-gu, Seoul, Korea, 110-749
Tel: 82-2-2072-2656 Fax: 82-2-2072-3851
E-mail: kum6139@snu.ac.kr

※ This study was supported by faculty grant of 2005 Yonsei dental college.

materials. Gutta-percha (GP) cones, now widely used to fill root canals, may become contaminated by pathogens during manufacturing and exposed to the air in clinics for several months. It is controversial whether the sterilization process is necessary because of anti-bacterial characteristic of a component of cone itself^{1,2)}, however, the sterilization of cones prior to root canal filling has been recommended in most cases.

The chemical agents that have been used to sterilize GP cone prior to canal filling are diverse. Recently, numerous experiments have been performed to seek chemical agents that act in a shorter time and sterilize various bacteria, and it has been reported that NaOCl, glutaraldehyde, chlorhexidine, etc. are effective³⁻⁷⁾. However, in these experiments, GP cone was infected with artificially selective bacteria and the sterilization effect among different sterilizers was compared. Therefore, it is necessary to identify the microbial species on GP cones exposed at the outpatient clinics and evaluate which chemical solution is more effective on such bacteria.

On the other hand, the physical changes of GP cones after sterilization have been pointed out. Moller *et al.* have claimed when the cone was treated with 70% isopropyl alcohol, 5% chloramine, and 0.5% chlorhexidine, the tensile strength of the cone decreased evidently, and thus the physical property of gutta-percha cone could be altered by the surface sterilizer⁸⁾. In addition, Lee *et al.* reported that from 1 day after the treatment with 70% isopropyl alcohol, 2.5%, and 5% NaOCl, the tensile strength and the elongation rate of the cone (evidently) decreased, and such alteration could mediate an effect during canal filling in clinics⁹⁾. Therefore, the purpose of this study was first, to identify the type of bacteria that contaminate GP cone exposed at two hospital-based dental clinics by polymerase chain reaction (PCR), and secondly to evaluate the short-term sterilization effect of three chemical disinfectants. It also analyzed the alteration of the surface texture and physical properties after short-term sterilization, particularly the tensile strength and the elongation rate.

II. MATERIALS AND METHODS

1. The measurement of the contamination rate of gutta-percha cone and the identification of bacteria by PCR

150 GP cones (Meta Biomed Co. Chung-Ju, Korea) kept exposed in two hospital-based dental clinics were collected and used in this study. From 10 immediately opened GP packs, total 30 cones were collected and contamination was assessed as control group. Each GP cone was placed in tube containing 200 μ l PBS buffer solution, and vortexed for 5 minutes. All GP cones were removed from the tube, 200 μ l PBS buffer solution was inoculated to a Brain Heart Infusion (BHI) agar plate, and cultured for 48 hours in a 37°C incubator. The number of colonies formed from each GP cone (CFU) was counted and the contamination level of total gutta-percha cones was evaluated. The colony pattern formed on the BHI agar plate was examined and classified by microscope.

Polymerase chain reaction (PCR) amplification was performed in a thermal cycler (Perkin-Elmer Inc. Boston, USA) using colony selected from BHI agar plate as a template. Primer, dNTP (Bioneer, Daejeon, Korea), Taq-polymerase (Bioneer, Daejeon, Korea), PCR buffer, and distilled water were added to a PCR tube together. At that time, the universal 16S rRNA primer (TpU1: 5'-AGAGTTTGTATCMTGGCTCAG-3', RTU3: 5'-GWATTACCGCGGCKGCTG-3', Bioneer, Daejeon, Korea) was used as a primer for PCR amplification. The PCR conditions used in this study were as follows: the initial denaturation was at 95°C for 5 minutes. Thirty amplification cycles were then performed: the denaturation reaction at 95°C for 1 minute, the annealing at 56°C for 1 minute, and the extension reaction at 72°C. After the electrophoresis and confirming of PCR products, they were purified using PCR purification system (Bioneer, Daejeon, Korea). Automatic nucleic acid sequencer analyzed the purified DNA. Using the nucleic acid sequence of bacteria thus obtained, the name of bacteria was identified using the

Table 1. Sterilization effect of three chemical agents from *Staphylococcus-inoculated* GP cones

Sterilization Time (Min)	Experimental solutions for sterilization of GP cones									
	5.25% NaOCl		2% CHX		ChloraPrep		Positive control		Negative control	
	T*	C [§]	T	C	T	C	T	C	T	C
1	-	-	-	-	-	-	+	+	-	-
5	-	-	-	-	-	-	+	+	-	-
10	-	-	-	-	-	-	+	+	-	-
30	-	-	-	-	-	-	+	+	-	-

T*: Turbidity, C[§]: Culture

Positive control: Contaminated cone without chemical soaking

Negative control: No artificially contaminated cone

BLAST program of nucleotide database (NCBI).

2. Evaluation of sterilization effect of three chemical disinfectants

The bacteria identified above were inoculated to BHI liquid medium and cultured at 37°C for 24 hours. A GP cone sterilized with EO gas was transferred to the BHI medium that bacteria were cultured, and remained in contact with the bacteria for 2 hours to promote surface contamination. The contaminated GP cone was transferred to a petri dish matted with two layers of filter paper and dried for 24 hours at room temperature. A contaminated GP cone was immersed in each chemical solution for 1 minute, 5 minutes, 10 minutes, and 30 minutes, and dried. As chemical disinfectants, 5.25% NaOCl, 2% chlorhexidine, and ChloraPrep one-step were used. ChloraPrep one-step is a mixed solution (vol: 1 : 1) of 75% isopropyl alcohol and 2% chlorhexidine and is a fast acting, broad spectrum, persistent antiseptic that significantly reduces the number of microorganisms on intact skin. After each decontamination method, all the GP cones were transferred to sterile trial tube containing sterile BHI and incubated at 37°C for 7 days. Growth, as indicated by the turbidity and the subculture of the examined BHI liquid medium, was then recorded. The subculture was performed with the following method:

the observed liquid medium 150 μ l was inoculated to a Brain Heart Infusion (BHI) agar plate, and cultured for 24 hours in a 37°C incubator. The sterilization was confirmed by colony forming state. The positive control group was the GP cone immersed in the BHI liquid medium that bacteria were cultured, and dried without sterilization, and cultured 24 hours in fresh BHI culture medium. The negative control group was a GP cone sterilized with EO gas without bacterial contamination and cultured in BHI liquid medium as is.

3. SEM observation of the surface texture of GP cone after 5 min soaking in chemical disinfectants

GP cones were immersed in 3 types of chemical disinfectants (5.25% NaOCl, 2% chlorhexidine, ChloraPrep one-step) for 5 minutes. Their surfaces were compared with that of a fresh GP cone by a scanning electron microscope (X500 and X10000, JSEM-820, Tokyo, Japan).

4. Assessment of the alteration of the physical property of GP cone after chemical sterilization

By using a Universal testing machine (Instron 5500, Canton, USA), the tensile strength and elongation rate of GP cones soaked in 3 types of

Table 2. A tensile strength of gutta-percha cone soaked in chemical solutions for 5 minutes (MPa)

Group	Mean (S.D)
Control	0.2091 (0.0147) ^a
2% CHX	0.2053 (0.0135) ^a
Chloraprep	0.2169 (0.0159) ^{a,b}
5.25% NaOCl	0.2264 (0.0183) ^b

Control: GP cone with no chemical soaking
 Different letters denote statistical significance ($p < 0.05$).

chemical disinfectants for 5 minutes were measured. In each experiment group, 40 cones were used. During the measurement of the tensile strength and the elongation rate, a rubber coating grip was used, the crosshead speed was 1.5 mm/min, and the room temperature was maintained at 25°C. The statistical association between the type of chemical disinfectants and the change of physical properties of GP cones was determined by One-way ANOVA and Duncan grouping with a significance level of 0.05.

III. RESULT

PCR identification of bacterial species on the surface GP cones

The immediately opened control GP cones showed negative cultures in all cases. Among total 150 GP cones opened in clinics, bacteria were detected on the surface of 29 gutta-percha cones (19.4%). The numbers of GP cone showing total 1-10 bacteria colonies were 13 (8.7%), the cones showing 11-100 bacterial colonies were 12 (8%), and the cones detected over 101 colonies were 4 (2.7%).

In addition, examining the pattern or color of the colonies formed on BHI agar plate, 1 or 2 types of bacteria were detected on each GP cone. From these colonies, 35 PCR products were puri-

Table 3. The elongation rate change of gutta-percha cone in chemical agents for 5 minutes. Elongation measures the percentage change in length before fracture (%)

Group	Mean (S.D)
Control	167.9 (22.5) ^a
5.25% NaOCl	216.9 (16.6) ^b
2% CHX	269.7 (31.9) ^c
Chloraprep	326.3 (42.0) ^d

Control: GP cone with no chemical soaking
 Different letters denote statistical significance ($p < 0.05$).

fied, and finally, 32 bacteria were identified. Most of them were *Staphylococcus spp.* In particular, *Staphylococcus epidermidis* (n = 20/32, 62.5%) was the most prevalent one. In addition, *Staphylococcus caprae* (n = 8/32, 25%), *Staphylococcus capitis* (n = 4/32, 12.5%), and *Staphylococcus xylosus* (n = 1/32, 3.1%) were also detected.

The sterilization effect of chemical solutions

Three chemical disinfectants showed sterilization effect after only 1 minute immersion, which was confirmed not only by the measurement of the turbidity but also by the subculture of the liquid culture medium. Detailed results are shown in Table 1.

SEM Observation of the surface texture of GP cone after 5-min soaking in chemical disinfectants

In comparison with GP cone that was not sterilized, the surface of GP cone immersed for 5 minutes in 2% chlorhexidine and Chloraprep one-step was slightly wrinkled with the pattern of shrinkage along the long axis of cone, and the adhesion or release of other materials on the surface was not detected. However, on GP cone immersed in 5.25% NaOCl, the precipitate with cuboidal crystal was attached to the overall sur-

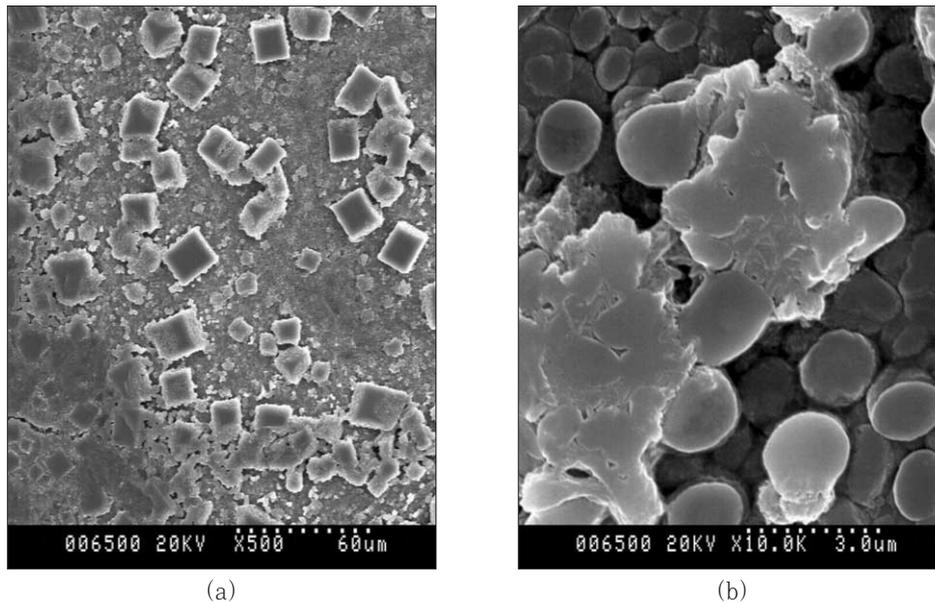


Figure 1. SEM images of the surface of gutta-percha cone after 5-min sterilization with 5.25% NaOCl. A cluster of cuboidal crystals (A) and some dissolved component of cone (B) were shown on the surface of GP cone (Magnification X500 and $\times 10000$).

face, and it was found to be particularly abundant in the defect area or folded area of cone (Figure 1a). Under X10000 magnification, a large quantity of round shaped structure was detected in the deep level of defect area of GP cone. It seemed that it was detected as the pattern of the partial lysis of some components of GP cone (Figure 1b).

The alteration of the physical property of GP cone after 5-min soaking in chemical disinfectants

Table 2 & 3 show the average values of the elongation rate and the tensile strength in each experimental group. In regard to tensile strength, the NaOCl-soaked group showed a significant difference than the other group (Table 2, $p < 0.05$). In terms of elongation rate, all chemical solutions showed significant difference than the control group, especially in the ChloroPrep group (Table 3, $p < 0.05$).

IV. DISCUSSION

Molecular genetic methods, particularly the polymerase chain reaction (PCR), have been widely used for microbial identification purposes. PCR assays are very sensitive and enable a reliable identification of microbial species or strains that are difficult or even impossible to culture¹⁰. In this study, the method of assay should detect the smallest possible number of microorganism. The high sensitivity of the PCR method makes it ideal for this purpose.

Our present study clearly showed that GP cones exposed in clinics can be contaminated by specific organisms, particularly *Staphylococcus spp.* *Staphylococcus epidermidis* detected most frequently in this study, is a normal flora residing in the skin and the mucosa of respiratory system in normal individuals. Nevertheless, in immune suppressed patients or in the case at a site different from the distribution of normal flora such as implanted medical devices, or when its number is

increased suddenly by other reasons, it may cause severe infection as opportunistic microorganism¹¹⁾.

In a previous study¹²⁾, GP cones in over-filled areas were covered with a biofilm structure and a colony of cocci was observed in the cracks of the biofilm structure. These findings suggested that cocci were located in deeper layers of the biofilm structure and these cocci might play an important role in initiating biofilm formation. In addition, the biofilm which form on the extra-radicular area of gutta-percha cone are related to refractory periapical pathosis, and gutta-percha cones might play a role in the initiation of biofilm infection in cases of excessive root filling with periapical lesions^{12,13)}. Therefore, considering the importance of the aseptic root canal and the contamination of the GP cone in clinics, it may be necessary to decontaminate the gutta-percha cones by means of a chemical disinfectant before canal filling. The time needed for disinfection of GP cone is various according to the types and concentrations of chemical disinfectants or the selective microbial species. It has been reported previously that 1 minute short sterilization with 5.25% NaOCl eradicated spores as well as bacteria. Because of its superior antibacterial effect, the sterilization of the GP cone with 5.25% NaOCl prior to canal filling has been recommended for a long time⁴⁻⁷⁾. However, Short et al. have shown that the cluster of cuboidal crystals on the surface of NaOCl-soaked GP cone was detected at various levels and mentioned that at the time of canal filling, it might affect the apical sealing¹⁴⁾. Our present study demonstrated not only cuboidal crystals but also round shaped structures that appeared to be the components of cone on the surface of NaOCl-soaked GP cone under high magnification. Therefore, it is essential to find other chemical disinfectants, which are more stable and clinically useful, to sterilize the contaminated GP cone.

ChlorPrep one-step (2% chlorhexidine gluconate + 70% isopropyl alcohol) is an approved antiseptic for preoperative skin preparation and has the fast bactericidal effect of alcohol and the substantive effect of chlorhexidine. Our results

showed that all three chemical disinfectants including ChlorPrep one-step were efficient for sterilization of GP cones by immersion only for 1 minute, which was confirmed by the turbidity measurement and subcultures. We contribute this result to the contamination of GP cone by *Staphylococcus*, which is a different species from the spore-forming bacteria, fungus with a thick lipid layer, and small virus lacking the envelope that need more powerful sterilization agents. Therefore, GP cone could be easily sterilized in three chemical disinfectants in the present study. Time is more critical when it is necessary to disinfect extra accessory cones during compaction techniques for multi-rooted tooth, because it needs 5 to 10 min for cone disinfection. Therefore, in this study we evaluated the changes of surface textures and physical properties of gutta-percha cones only after 5-min soaking in chemical solutions.

As the factors that alter the physical property of GP cone, the content of gutta-percha, storage conditions, the surface treatment with chemical solutions, etc. have been suggested. Similarly, in the result of our study, it was found that the treatment with chemical solutions affected the tensile strength and elongation rate of gutta-percha cone. Particularly, the result of the treatment with NaOCl was different from that of the study reported by Lee⁹⁾. This difference is supposed to result from sterilization time. In the study performed by Lee, the period immersed in NaOCl was from 1 day to 30 days and this long-term sterilization treatment generated difference from our results. On the other hand, Valois has shown by atomic force microscopy that only 1 minute treatment with 5.25% NaOCl changed the elasticity of GP cone¹⁵⁾.

On root canal filling, the alteration of physical property of GP cone may affect the outcome of root canal treatment. In other words, in the cases of the cone with a high elongation rate, it was thought that the change of the shape of cone itself to irregular canal shape may occur more readily by the same load and it could compact the

space between cones more densely. Nonetheless, other physical properties of cone such as modulus of elasticity, compressive strength, etc. have to be considered together. Hence, it is considered that additional clinical studies have to be performed in regard to the actual effect of the change of the physical property of cone by sterilization. In conclusion, our present study demonstrates that three chemical solutions are useful for short-term sterilization of gutta-percha cone before canal filling. Further research is, however, needed to determine the clinical relevance of the changes of physical properties on gutta-percha cone after chemical disinfection.

REFERENCES

1. Moorer WR, Genet JM. Evidence for antibacterial activity of endodontic gutta-percha cones. *Oral Surg* 53:503-507, 1982.
2. Delivanis Pd, Mattison GD, Mendel RW. The survivability of F43 strain of streptococcus sanguis in root filled with gutta-percha and Procosol cement. *J Endod* 9:408-412, 1983.
3. Cardoso CL, Kotaka CR, Guilhermetti M, Hidalgo MM. Rapid sterilization of gutta-percha cones with glutaraldehyde. *J Endod* 24:561-569, 1998.
4. Cardoso CL, Kotaka CR, Redmerski R, Guilhermetti M, Queiroz AF. Rapid decontamination of gutta-percha cones with sodium hypochlorite. *J Endod* 25:498-501, 1999.
5. de Motta PG, de Figueiredo CB, Maltos SM, Nicoli JR, Ribeiro Sobrinho AP, Maltos KL, Carvalhais HP. Efficacy of chemical sterilization and storage conditions of gutta-percha cones. *Int Endod J* 34:435-0, 2001.
6. de Souza RE, de Souza EA, Sousa-Neto MD, Pietro RC. In vitro evaluation of different chemical agents for the decontamination of gutta-percha cones. *Pesqui Odontol Bras* 17:75-77, 2003.
7. Siqueira JF JR, da Siliva CH, Cerqueira M das D, Lopes HP, de Uzeda M. Effectiveness of four chemical solutions in eliminating Bacillus subtilius spores on gutta-percha cones. *Endod Dent Traumatol* 14:124-126, 1998.
8. Moller B, Orstavik D. Influence of antiseptic storage solution on physical properties of endodontic gutta-percha points. *Scand J Dent Res* 93:158-161, 1985.
9. Lee MS. An experimental study of the effect of the various antiseptic storage solutions on physical properties of gutta-percha cone. MS Thesis, 1989, Yonsei Dental College.
10. Siqueira JF JR, Rocas IN. PCR methodology as a valuable tool for identification of endodontic pathogens. *J Dent* 31:333-339, 2003.
11. Gill SR, Fouts DE, Archer GL: Insights on evolution of virulence and resistance from the complete genome analysis of an early Methicillin-Resistant Staphylococcus aureus strain and a biofilm-producing Methicillin-Resistant staphylococcus epidermidis strain. *J Bacteriol* 187:2426-38, 2005.
12. Noiri Y, Ehara A, Kawahara T, Takemura N, Ebisu S. Participation of bacterial biofilms in refractory and chronic periapical periodontitis. *J Endod* 28:679-683, 2002.
13. Takemura N, Noiri Y, Ehara A, Kawahara T, Noguchi N, Ebuisu S. Single species biofilm-forming ability of root canal isolates on gutta-percha points. *Eur J Oral Sci* 112:523-529, 2004.
14. Short RD, Dorn SO, Kuttler S. The crystallization of sodium hypochlorite on gutta-percha cones after the rapid-sterilization technique: an SEM study. *J Endod* 29: 670-673, 2003.
15. Valois CR, Silva LP, Azevedo RB. Effects of 2% chlorhexidine and 5.25% sodium hypochlorite on gutta-percha cones studied by atomic force microscopy. *Int Endod J* 38:425-429, 2005.

국문초록

Short-term chemical disinfection 후 가타파차 콘의 멸균 효과와 표면 성장 및 물성 변화의 평가

방난심¹ · 정일영¹ · 유윤정² · 금기연^{3*}

연세대학교 치과대학 ¹치과보존학교실, ²구강생물학교실,

³서울대학교 치의학대학원 치과보존학교실

본 연구의 목적은 첫째 임상에서 진료실에 노출된 가타파차 콘 표면의 오염 균종을 중합효소연쇄반응법(polymerase chain reaction, PCR)을 이용해 동정하고, 둘째 이들 세균으로 오염시킨 가타파차 콘에 대해 3종의 소독제의 short-term sterilization 효과를 비교하였다. 또한 이들 소독제에 5분간 처리된 가타파차 콘 표면 성장의 변화를 주사전자현미경으로 관찰하였고 물성 변화, 특히 인장 강도와 연신률의 변화를 측정, 비교하고자 하였다. 진료실에 수 개월간 노출된 가타파차 콘 150개를 수거하여 배양지에 넣어 배양 후 universal primer를 사용한 PCR assay를 통해 오염 균종을 동정하였다. 실험실 상에서 이 균종을 다시 배양하여 소독된 가타파차 콘에 접종하고 1주일간 배양한 후 3종의 소독제 (5% NaOCl, 2% Chlorhexidine, ChloraPrep)에 1, 5, 10, 30분간 short-term soaking 후 각 소독제의 종류와 적용시간에 따른 멸균 효과를 turbidity test와 subculture를 이용하여 평가하였다. 또한 각 소독제에 5분간 처리된 가타파차 콘 표면 성장의 변화를 주사전자현미경으로 관찰하였으며, 물성 변화를 평가하기 위해 Instron 5500을 이용하여 연신률 및 tensile strength를 측정한 후 통계학적 유의성 여부를 검증하였다. 중합효소연쇄반응법의 분석결과 19.4%의 가타파차 콘이 오염된 것으로 나타났고 대부분이 Staphylococcus 계통이었으며, 3종의 소독제 모두 이들 균종에 대해 1분내에 멸균 효과를 나타냈다. 주사전자현미경상 NaOCl로 소독된 가타파차 콘 표면에는 cuboidal crystal의 침전물이 전반적으로 관찰되었다. Tensile strength는 NaOCl군에서 다른 군에 비해 유의성 있는 증가를 보였으며 ($p < 0.05$), elongation rate는 3종의 소독제 모두 대조군에 비해 유의성 있는 증가를 보였으나 ($p < 0.05$), 특히 ChloraPrep군에서 가장 큰 증가를 보였다. 본 연구 결과 3종의 소독제 모두 근관충진 전 가타파차 콘의 rapid sterilization을 위해 유용하게 사용할 수 있음을 시사한다.

주요어: 중합효소연쇄반응법, 표면성장, 물성변화, 멸균효과, 가타파차 콘