

MMP-1 and TIMP-1 production in MG-63 cells stimulated with *Prevotella nigrescens* Lipopolysaccharide

Won-Kyung Yang¹, Mi-Ri Kim², Won-Jun Shon¹, In-Bog Lee¹, Byeong-Hoon Cho¹, Chung-Moon Um¹, Ho-Hyun Son^{1*}

¹Department of Conservative Dentistry, College of Dentistry, Seoul National University

²Department of Conservative Dentistry, Asan Medical Center

ABSTRACT

The purpose of this study is to monitor the secretion of matrix metalloproteinase-1 (MMP-1) and tissue inhibitor of metalloproteinase-1 (TIMP-1) produced by human osteosarcoma cell line (MG63) stimulated with *Prevotella nigrescens* lipopolysaccharides (LPS), and to compare the level of secretion before and after the treatment of calcium hydroxide on *P. nigrescens* LPS.

LPS was extracted and purified from anaerobically cultured *P. nigrescens*. MG63 cells were stimulated by the LPS (0, 1, 10 $\mu\text{g/ml}$) or LPS (10 $\mu\text{g/ml}$) pretreated with 12.5 mg/ml of Ca(OH)_2 for 3 days.

Total RNA was isolated from the cell, and real-time quantitative polymerase chain reaction (PCR) was performed for quantification of MMP-1 and TIMP-1.

The results were as follows.

1. MMP-1 mRNA expression at 48 hr was highly increased by stimulation with *P. nigrescens* LPS. The increase was dose-dependent.
2. When stimulated with 1 $\mu\text{g/ml}$ of LPS, TIMP-1 mRNA expression was highly increased at 24 hr and 48 hr. However, TIMP-1 expression was suppressed at higher concentration (10 $\mu\text{g/ml}$).
3. When *P. nigrescens* LPS was pretreated with Ca(OH)_2 , MMP-1 and TIMP-1 gene expression was downregulated.

The results of this study suggest that transcriptional regulation of MMP-1 and TIMP-1 by *P. nigrescens* LPS could be one of the important mechanisms in bone resorption of periapical inflammation. The result of calcium hydroxide on MMP-1 and TIMP-1 gene expression suppression shows that calcium hydroxide detoxified bacterial LPS and thus should be used the medication of choice for intracanal dressings in root canal infected with black-pigmented bacteria. [J Kor Acad Cons Dent 29(5):470-478, 2004]

Key words : MMP-1, TIMP-1, *P. nigrescens*, LPS, Ca(OH)_2 , MG63

I. INTRODUCTION

Accumulation of bacterial byproduct, such as lipopolysaccharide (LPS) in the root canal system

can stimulate the host cells in the periapical tissue. These cells such as neutrophils, macrophages and lymphocytes are activated and release inflammatory mediators to initiate and augment subsequent inflammatory reactions leading to periapical tissue breakdown¹⁾.

As is the tissue made up of organized group of cells attached to an extracellular matrix, tissue homeostasis is maintained by coordinating cell growth and proliferation with the production and turnover of the extracellular matrix²⁾. It is con-

* Corresponding author: Ho-Hyun Son

Dept. of Conservative Dentistry,
College of Dentistry, Seoul National University
28 Yoengun-dong, Chongro-gu, Seoul, Korea, 110-749
Tel : 82-2-760-1634 Fax : 82-2-760-3859
E-mail : hhson@snu.ac.kr

ceivable that, since the major component of the organic bone matrix is collagen, collagenolytic enzyme, synthesized by osteoblasts, may also participate in the degradation of the mineralized collagen during bone resorption³.

Matrix metalloproteinases (MMPs) are well known proteolytic enzymes for the degradation of extracellular matrix and this proteolysis is believed to play an essential role in the development and progression of pulpal and periapical inflammation⁴. MMPs, produced by osteoblast-like cells, are likely to be involved in various steps of the bone resorption process⁵.

Tissue inhibitor of metalloproteinase (TIMP) is also secreted from the same cells that secrete MMPs and block the MMP activity^{6,7}. The inhibitor was found in association with bone tissue *in vitro* and *in vivo* in immunohistochemical and biochemical studies^{8,9}, and these inhibitors have been known to inhibit bone resorption by maintaining a delicate balance with activities of extracellular matrix-degrading MMP¹⁰. Resorption of bone may be the results of complex interactions between metalloproteinases and inhibitors produced by cells of bone, a process that is at least in part regulated by local cytokines including bacterial LPS⁵. By detecting the balance between the level of MMP and their inhibitor activation, one of the host degradative pathways in the pathogenesis of bacterial-derived pulpal and periapical lesion will be revealed.

Recently, it was demonstrated that black-pigmented *Bacteroides* (BPB) species play an important role in tissue disintegration of extracellular matrix in pulpal and periapical diseases^{4,11}. The cells of these lesions respond to LPS of the organisms and produce various inflammatory mediators. LPS, localized in the cell wall of Gram-negative bacteria, is known to have a broad spectrum of biochemical and immunochemical activities that can lead to the destruction of host tissue¹². There are increasing evidences that bacterial LPS treated with calcium hydroxide intracanal dressings may have altered biological properties^{13,14}.

Among these BPB, *Prevotella nigrescens* is the

most commonly isolated species from the infected root canals, and thought to be related to endodontic infections¹⁵⁻¹⁷. However, there were little information concerning the effects of *P. nigrescens* LPS on the activation of MMPs produced from osteoblast-like cells, and the influence of calcium hydroxide treatment. Thus, the purpose of this study is to monitor the secretion of MMP and TIMP produced by osteoblastic cells stimulated with *P. nigrescens* LPS and to compare the level of secretion before and after the treatment of calcium hydroxide on *P. nigrescens* LPS.

II. MATERIALS AND METHODS

1. Bacterial cell culture and purification of lipopolysaccharides

The bacterial strain used was *P. nigrescens* ATCC33563, and was cultured in brain heart infusion (BHI) broth supplemented with Hemin (10 $\mu\text{g/ml}$), vitamin K1 (1 $\mu\text{g/ml}$), and yeast. Bacterial cells were grown anaerobically at 37°C. After sufficient growing, the bacterial cell pellet was obtained by centrifugation at 10,000 \times g for 10 min. This pellet was washed with distilled water and then lyophilized. LPS was extracted and purified by the method as described by Eidhin and Mouton in 1993¹⁸. The lyophilized bacteria was mixed with 1 ml of distilled water and then boiled for 15 min, with 5 min of inverted mixing period. Following centrifugation at 12,000 \times g for 5 min, cellular debris was removed, and the supernatant was collected. Then proteinase K (1 mg dissolved in 50 $\mu\text{g/ml}$ water) was added and incubated for 1 h at 60°C. After the tube was boiled for 5 min to precipitate any residual proteinase K, the supernatant was dialyzed against distilled water. After lyophilize the supernatant, the final crude LPS was redissolved to a concentration of 1 mg/ml of distilled water. The purified LPS was confirmed in 12% SDS-PAGE.

2. Osteoblast culture

Human osteogenic sarcoma cell line (MG63) was obtained from the ATCC (CRL-1427). For the experiment, these cells were plated on 100mm dish in α -MEM supplemented with 1 mM sodium pyruvate, 1 mM non-essential amino acid, 10% FBS, and antibiotics at 37°C in 5% CO₂. After confluency, cells were passed on 6 well plate and stimulated by LPS (0, 1, 10 μ g/ml) for various periods of time (8, 12, 24 h). In addition, cells were stimulated with LPS (10 μ g/ml) pretreated with 12.5 mg/ml of Ca(OH)₂ for 3 days.

3. Primer Design

For Real-time PCR amplification, optimal primers were designed using Primer express™ software (PE). They are as follows:

MMP-1 :

Forward: 5' ACTGCTGAGTGGCTACCCA 3' ,

Reverse: 5' GCATCCACGGACTGCATATTT 3' ,

TIMP-1 :

Forward: 5' CGCAGCGAGGAGTTCTCATT 3' ,

Reverse: 5' TGCATTCCTCACAGCCAACA 3' ,

GAPDH :

Forward: 5' GAAGGTGAAGGTCGGAGTC 3' ,

Reverse: 5' GAAGATGGTGATGGGATTTTC 3' ,

4. Isolation of RNA and real-time PCR assay

After stimulation of cells with LPS and Ca(OH)₂ treated LPS for various periods of time, total RNA from the cell was isolated according to the TRIzol Reagent method (Gibco) as described below. The cells were harvested with 0.5 ml of TRIzol reagent and then 100 μ l of chloroform was added and inverted mixed. Following centrifugation at 15,000 rpm for 30 min at 4°C, aqueous phase was collected into a fresh tube and mixed with additional 0.25 ml of isopropyl alcohol. After incubation for 10 min on ice, the sample were centrifuged at 15,000 rpm for 10 min at 4°C to precipitate RNA. The RNA pellet was washed with 1 ml of 75% Ethanol in DEPC and cen-

trifuged at 15,000 rpm for 10 min at 4°C. The isolated RNA was dissolved in DEPC. Extracted total RNA was quantified using spectrophotometric method to determine the concentration of the primer received. RT reaction mixture containing RT primer, oligo(dT)₂₀ primer, 1 μ g of total RNA, Reverse Transcriptase (Invitrogen) was incubated at 42°C for 50 min, after which the cDNA sample was stored at -20°C. Thermal cycling parameters for RT reactions are as follows; incubation for 10 min at 25°C, RT for 30 min at 48°C, RT inactivation for 5 min at 95°C.

Real-time quantitative PCR was performed by a GeneAmp 5700 sequence detection system (PE, Applied Biosystems) with CYBR Green Master Mix (PE, Applied Biosystems) to assess gene expression. CYBR Green is highly specific to double strand DNA: upon binding to double-strand DNA, it exhibits a large increase in fluorescence. Direct detection of PCR product is monitored by measuring the increase in fluorescence caused by the binding of CYBR Green to double-strand DNA. The reaction mixture (25 μ L) contains 0.5 μ L of the cDNA sample with 0.25 μ M of the PCR primer. Two tube method for target genes and control gene (GAPDH) was used in 96 well plate.

The cycle profile was as follows: 1 cycle at 50°C for 2 min, 1 cycle at 95°C for 1 min, 40 cycles at 95°C for 15 sec, and 60°C for 1 min. After the last cycle, a dissociation protocol was performed as follows: a hold at 95°C for 15 sec, a hold at 60°C for 20 sec, and a slow ramp (20 min) from 60 to 95°C. The dissociation curve was captured during this slow ramp. Experiments were performed in triplicate. The critical threshold cycle (C_T) was defined as the cycle at which fluorescence became detectable above the background and was inversely proportional to the logarithm of the initial number of template molecules. A standard curve was plotted for each primer set with C_T values obtained from amplification of known quantities of the genomic DNA. By the standard curve of C_T vs (logN) starting copy number for all standards, it can be determined the starting copy number of unknowns by interpolation.

III. RESULTS

After stimulated with 0, 1, 10 $\mu\text{g}/\text{ml}$ of *Prevotella nigrescens* LPS at 8, 24 and 48 h, total mRNA of MMP-1 and TIMP-1 were isolated and evaluated by RT-PCR. The threshold cycle was detected and normalized with control gene, GAPDH. The expression of GAPDH was used to compare assay reproducibility utilizing the ratio between raw data and normalized expression as a percent coefficient by employing PCR. The ratio between RNA content and GAPDH of each sample was shown in Table 1 and 2.

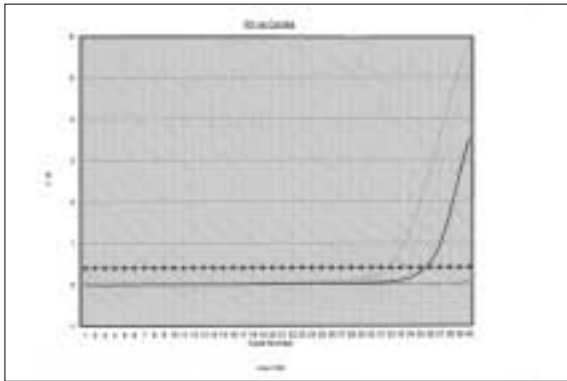


Figure 1. Amplification plots of real time PCR for MMP-1. Dashed line (—x—) indicates the threshold cycle.

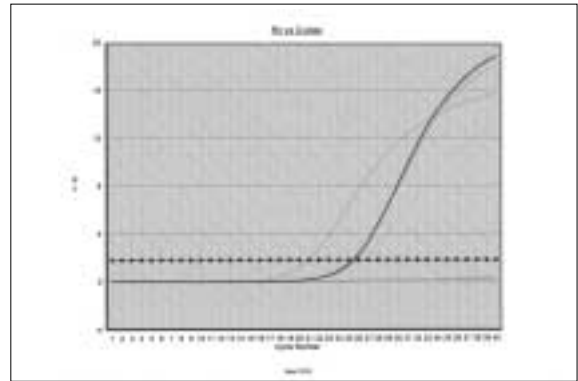


Figure 2. Amplification plots of real time PCR for TIMP-1. Dashed line (—x—) indicates the threshold cycle.

Table 1. The ratio between the mRNA expression of MMP-1 and GAPDH in osteoblast stimulated with *P. nigrescens* LPS. The data were acquired in triplicate of each sample.

MMP-1	LPS ($\mu\text{g}/\text{ml}$)	8 hr	24 hr	48 hr
(-)control	0	0.22	1.99	1.21
<i>E. coli</i>	1	1.14	11.77	19.94
	10	1.57	12.94	27.90
<i>P. nigrescens</i>	1	4.20	3.01	15.12
	10	8.19	8.15	16.22

Table 2. The ratio between the mRNA expression of TIMP-1 and GAPDH in osteoblast stimulated with *P. nigrescens* LPS. The data were acquired in triplicate of each sample.

TIMP-1	LPS ($\mu\text{g}/\text{ml}$)	8 hr	24 hr	48 hr
(-) control	0	4.36	1.54	2.15
<i>E. coli</i>	1	17.46	1.91	2.08
	10	0.18	0.15	1.00
<i>P. nigrescens</i>	1	5.73	20.59	18.10
	10	1.36	3.83	3.15

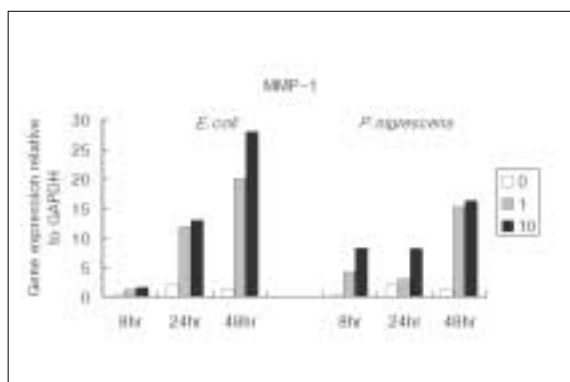


Figure 3. MMP-1 gene expression relative to GAPDH at different LPS concentrations ($\mu\text{g/ml}$) and different periods of time.

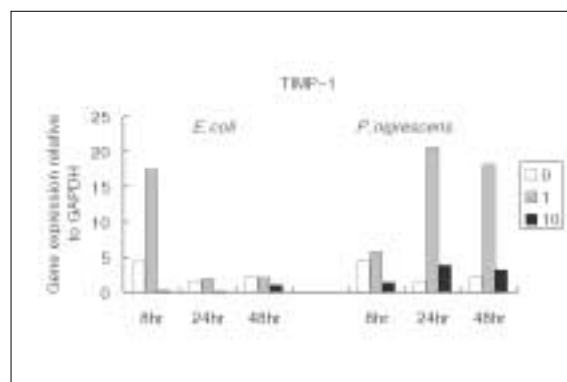


Figure 4. TIMP-1 gene expression relative to GAPDH at different LPS concentrations ($\mu\text{g/ml}$) and different periods of time.

Table 3. The ratio between the mRNA expression of MMP-1 and GAPDH in osteoblast-like cells stimulated with *P. nigrescens* LPS pretreated with $\text{Ca}(\text{OH})_2$. The data were acquired in triplicate of each sample.

MMP-1	24 hr	48 hr	72 hr
<i>E. coli</i>	0.72	0.69	0.88
<i>P. nigrescens</i>	0.65	0.39	0.35

Table 4. The ratio between the mRNA expression of TIMP-1 and GAPDH in osteoblast-like cells stimulated with *P. nigrescens* LPS pretreated with $\text{Ca}(\text{OH})_2$. The data are acquired in triplicate of each sample.

TIMP-1	24 hr	48 hr	72 hr
<i>E. coli</i>	0.11	0.63	1.02
<i>P. nigrescens</i>	0.06	0.34	0.49

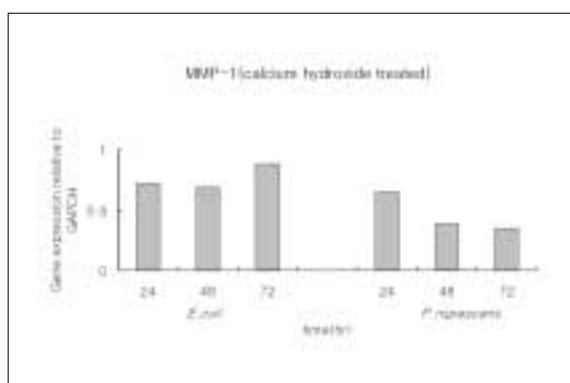


Figure 5. MMP-1 gene expression relative to GAPDH in osteoblasts stimulated with 10 $\mu\text{g/ml}$ of *P. nigrescens* LPS pretreated with $\text{Ca}(\text{OH})_2$ at different periods of time.

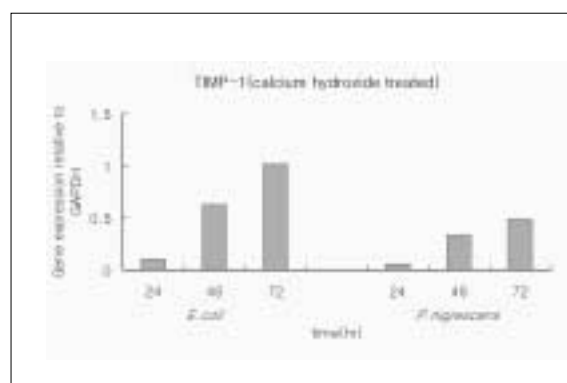


Figure 6. TIMP-1 gene expression relative to GAPDH in osteoblasts stimulated with 10 $\mu\text{g/ml}$ of *P. nigrescens* LPS pretreated with $\text{Ca}(\text{OH})_2$ at different periods of time.

IV. DISCUSSION

A hallmark of periapical disease is its ability to degrade the surrounding extracellular matrix,

resulting in destruction of bone in the periapical tissue. There are increasing evidences that MMPs are essential enzymes responsible for degradation of extracellular matrix in the pathological condi-

tions associated with inflammations¹⁹⁾. Numerous studies have examined MMPs secreted from periodontal ligament fibroblast²⁰⁻²²⁾ and correlated the level of expression with the invasive ability of the disease^{4,6)}. Individual osteoblasts, like fibroblasts, are capable of both synthesizing and degrading their respective organic matrices *in vivo*²³⁾. MMPs and TIMPs are known to be important in homeostatic matrix remodeling by osteoblasts.

However, MMP and TIMP production under stimulation of bacterial LPS in osteoblastic cell has not been well studied.

Conflicting reports exist concerning the production of MMP-1 by normal human osteoblasts. Rifas et al. reported that human osteoblasts are unable to produce MMP-1 either constitutively or in response to bone resorptive agents²⁴⁾. De Bart et al. also reported that the mRNA for MMP-1 was not detectable under normal conditions in MG63 cells by Northern blot analysis²⁵⁾.

In the present study, the expression of MMP-1 mRNA was low, but obvious in the negative control. This result consisted with the findings of Panagakos et al. that the expression of MMP-1 was low and essentially invariant over a culture period of human osteoblastic cell line²⁶⁾. This constitutive MMP-1 seems to serve as a remodeling element of connective tissue under physiological conditions²⁷⁾. Otsuka et al. also demonstrated that rat osteosarcoma cell line produced collagenolytic enzymes²³⁾.

MMP-1 mRNA was highly expressed in this study after 48 h stimulated with *Prevotella nigrescens* LPS, and the increase was dose-dependent. The results consisted with many other studies. Nakata et al. demonstrated that the production of MMP-1 was accelerated by sonicated bacterial extracts²⁸⁾. Shin et al. also confirmed this result by detecting high concentration of MMP-1 in relation to clinical symptoms²⁹⁾.

The activity of MMPs is under strict regulation by a group of tissue inhibitor of metalloproteinases (TIMPs), of which TIMP-1 to TIMP-4 have been identified³⁰⁾. TIMP-1, a glycoprotein with a molecular mass of 28kDa, can form a stoichiometric 1:1 complex with MMP-1 and inhibit the

activity of MMP-1¹⁹⁾. Because the same cells that produce MMP-1 are also capable of synthesizing TIMP-1, the overall molar ratio between MMP-1 and TIMP-1 plays a critical role in connective tissue remodeling³¹⁾.

In this study, TIMP-1 was expressed by MG-63 osteoblastic cells. Otsuka²³⁾, Johansen³²⁾, Uchida³³⁾ also reported that osteoblastic cells synthesized TIMP-1. These results back up our findings.

During the experimental period of time, TIMP-1 expression was highly increased at 24 h and 48 h under the 1 $\mu\text{g/ml}$ of *P. nigrescens* LPS. However, TIMP-1 secretion was suppressed when exposed to higher concentration (10 $\mu\text{g/ml}$) of bacterial LPS. The reason of the downregulation of TIMP-1 could be a perturbation in cell-extracellular matrix interaction by the high virulence of bacterial LPS and in turn it culminated in tissue destruction. A better understanding of TIMP activity will be helpful in understanding the etiology of the periapical lesion and its treatment strategies.

LPS is known to exert profound effect on the protein expression by host cells. Wahl et al. has demonstrated that LPS stimulates collagenase release via a mechanism involving PGE₂ and cAMP³⁴⁾. This can be the one of the mechanisms that bacterial LPS affect on the development of periapical bone resorption. *Prevotella nigrescens* LPS was used in this study because infection of the root canals is frequently associated with BPBs and among the isolated black-pigmented bacteroides species, this microorganism is most often isolated from infections of endodontic origin^{15,17)}. Gharbia et al. also reported that *P. nigrescens* is the predominant cultivable BPB from both coronal and apical portions of infected root canals¹⁶⁾. *E.coli* LPS was used as a positive control because its chemical structure and stimulatory effects have been investigated extensively¹²⁾.

There have been increasing evidences that calcium hydroxide may detoxify bacterial LPS by altering its biological properties^{13,14,35)}. Use of calcium hydroxide as a routine intracanal dressing has been advocated in recent years. Calcium hydroxide treatment was effective in suppression of dev-

astating effect of MMP in this study. The fact that the expression of MMP was downregulated when stimulated with either *P. nigrescens* or control *E. coli* LPS pretreated with calcium hydroxide indicate that it is effective in preventing extracellular matrix degradation by reducing MMP production from osteoblast. Calcium hydroxide seems to be clinically beneficial when LPS is suspected in the root canal.

IV. CONCLUSION

The mRNA expressions of MMP-1 and TIMP-1 produced by Human osteosarcoma cell line (MG63) stimulated with *P. nigrescens* LPS (0, 1, 10 $\mu\text{g/ml}$) and calcium hydroxide, pretreated *P. nigrescens* LPS (10 $\mu\text{g/ml}$) for various periods of time, were monitored by real-time PCR.

The results were as follows.

1. MMP-1 mRNA expression at 48 h was highly increased by stimulation with *P. nigrescens* LPS. The increase was dose-dependent.
2. When stimulated with 1 $\mu\text{g/ml}$ of LPS, TIMP-1 mRNA expression was highly increased at 24 h and 48 h. However, TIMP-1 expression was suppressed at higher concentration (10 $\mu\text{g/ml}$).
3. When *P. nigrescens* LPS was pretreated with Ca(OH)_2 , MMP-1 and TIMP-1 gene expression was downregulated.

The results of this study are suggestive that transcriptional regulation of MMP-1 and TIMP-1 by *P. nigrescens* LPS is one of the important mechanisms in bone resorption of periapical inflammation. The result of calcium hydroxide on MMP-1 and TIMP-1 gene expression suppression shows that calcium hydroxide detoxified bacterial LPS and thus should be used as a medication of choice for intracanal dressings in root canal infected with black-pigmented bacteria.

REFERENCES

1. Trowbridge H. Immunological aspects of chronic inflammation and repair. *J Endod* 16:54-61, 1990.
2. Yang YY, Tsai HF, Lu SC, Huang YF, Chang YC. Regulation of tissue inhibitors of metalloproteinase-1 gene expression by cytokines in human gingival fibroblasts. *J Endod* 28:803-805, 2002.
3. Overall CM, Sodek J. Initial characterization of a neutral metalloproteinase, active of native 3/4 collagen fragments, synthesized by ROS 17/2.8 osteoblastic cells, periodontal fibroblasts, and Identified in gingival crevicular fluid. *J Dent Res* 66:1271-1282, 1987.
4. Chang YH, Lai CC, Yang SF, Chan Y, Hsieh YS. Stimulation of matrix metalloproteinases by black-pigmented bacteroides in human pulp and periodontal ligament cell cultures. *J Endod* 28:90-93, 2002.
5. Van der Zee E, Jansen I, Hoebe K, Beertsen W, Everts V. EGF and IL-1 α modulate the release of collagenase, gelatinase and TIMP-1 as well as the release of calcium by rabbit calvarial bone explants. *J Perio Res* 33:65-72, 1998.
6. Cury JD, Campbell EJ, Lazarus CJ, Albin RJ, Welgus HG. Selective up-regulation of human alveolar macrophage collagenase production by lipopolysaccharide and comparison to collagenase production by fibroblasts. *J Immunol* 141:4306-4312, 1988.
7. Seguir S, Gogly B, Bodineau A, Godeau G, Brousse N. Is collagen breakdown during periodontitis linked to inflammatory cells and expression of matrix metalloproteinases and tissue inhibitors of metalloproteinases in human gingival tissue? *J Periodontol* 72:1398-1406, 2001.
8. Lin SK, Chiang CP, Hong CY, Lin CP, Lan WH, Hsieh CC, Kuo MYP. Immunolocalization of interstitial collagenase(MMP-1) and tissue inhibitor of metalloproteinase-1(TIMP-1) in radicular cysts. *J Oral Pathol Med* 26:458-463, 1997.
9. Evert V, Hoegen K, Beertsen W. The release of tissue inhibitor of metalloproteinases by calvarial bone explants and its immunolocalization. *Bone Miner* 22:43-55, 1993.
10. Hill PA, Reynolds JJ, Meikle MC. Inhibition of stimulated bone resorption in vitro by TIMP-1 and TIMP-2. *Biochem Biophys Acta* 1177:71-74, 1993.
11. Hosoya S, Matsushima K. Stimulation of interleukin-1 β production of human dental pulp cells by *Porphyromonas endodontalis* lipopolysaccharides. *J Endod* 23:39-42, 1997.
12. Koga T, Nishihara T, Fujiwara T, Nisizawa T, Okahashi N, Noguchi T, Hamada S. Biochemical and immunological properties of lipopolysaccharide(LPS) from *Bacteroides gingivalis* and comparison with LPS from *Escherichia coli*. *Infect Immun* 47:638-647, 1985.
13. Safavi KE, Nichols FC. Alteration of biological properties of bacterial lipopolysaccharide by calcium hydroxide treatment. *J Endod* 20:127-129, 1994.
14. Safavi KE, Nichols FC. Effects of calcium hydroxide on bacterial lipopolysaccharide. *J Endod* 19:76-78, 1993.
15. Baumgartner JC, Watkins BJ, Bae KS, Xia T. Association of black-pigmented bacteria with endodontic infections. *J Endod* 25:413-415, 1999.
16. Gharbia S, Haapasalo M, Shah H, et al. Characterization of *Prevotella intermedia* and *Prevotella nigrescens* isolates from periodontic and endodontic origin. *J Periodontol* 65:56-61, 1994.
17. Bae KS, Baumgartner JC, Shearer TR, David LL. Occurrence of *Prevotella nigrescens* and *Prevotella intermedia* in infections of endodontic origin. *J Endod* 23:620-623, 1997.
18. Eidhin DN, Mouton C. A rapid method for preparation of rough and smooth lipopolysaccharides from

- Bacteroides*, *Porphyromonas* and *Prevotella*. *FEMS Microbiol Lett* 110:133-138, 1993.
19. Brikedal-Hansen H, Moore WBI, Bodden MK, Winsor LJ, Brikedal-Hansen B, DeCarlo A, Engler JA. Matrix metalloproteinases: A review. *Crit Rev Oral Biol Med* 4:197-250, 1993.
 20. Alvares O, Klebe R, Grant G, Cochran DL. Growth factor effects on the expression of collagenase and TIMP-1 in periodontal ligament cells. *J Periodontol* 66:552-558, 1995.
 21. Bolcato-Bellemin AL, Elkaim R, Abehsera A, Fausser JL, Haikel Y, Tenenbaum H. Expression of mRNAs encoding for α and β integrin subunit, MMPs, and TIMPs in stretched human periodontal ligament and gingival fibroblast. *J Dent Res* 79:1712-1716, 2000.
 22. Nakaya H, Osawa G, Iwasaki N, Cochran DL, Kamoi K, Oates TW. Effects of bisphosphonate enzymes in human periodontal ligament cells. *J Periodontol* 71:1158-1166, 2000.
 23. Otsuka K, Sodek J, Limeback H. Synthesis of collagenase and collagenase inhibitors by osteoblast-like cells in culture. *Eur J Biochem* 145:123-129, 1984.
 24. Rifas L, Halstead LR, Peck WA. Human osteoblasts in vitro secrete tissue inhibitor of metalloproteinases and gelatinase but not interstitial collagenase as major cellular products. *J Clin Invest* 84:686-694, 1989.
 25. De Bart ACW, Quax PHA, Lowik CWGM, Verheijen. Regulation of plasminogen activation, matrix metalloproteinases and urokinase-type plasminogen activator-mediated extracellular matrix degradation in human osteosarcoma cell line MG63 by interleukin-1 alpha. *J Bone Miner Res* 10:1374-1384, 1995.
 26. Panagakos FA, Kumar S. Differentiation of human osteoblastic cells in culture: Modulation of proteases by extracellular matrix and tumor necrosis factor-alpha. *Inflammation* 19:423-443, 1995.
 27. Lin SK, Wang CC, Huang S, Lee JJ, Chiang CP, Lan WH, Hong CY. Induction of dental pulp fibroblast matrix metalloproteinase-1 and tissue inhibitor of metalloproteinase-1 gene expression by interleukin-1 α and tumor necrosis factor- α through a prostaglandin-dependent pathway. *J Endod* 27:185-189, 2001.
 28. Nakata K, Yamasaki M, Iwata T, Suzuki K, Nakane A, Nakamura H. Anaerobic bacterial extracts influence production of matrix metalloproteinases and their inhibitors by human dental pulp cells. *J Endod* 26:410-413, 2000.
 29. Shin SS, Lee JI, Baek SH, Lim SS. Tissue levels of matrix metalloproteinases in pulps and periapical lesions. *J Endod* 28:313-315, 2002.
 30. Douglas DA, Shi YE, Sang QA. Computational sequence analysis of the tissue inhibitor of metalloproteinase family. *J Protein Chem* 16:237-255, 1997.
 31. Dean DD, Martel-Pelletier J, Pelletier JP, Howell DS, Woessner JF Jr. Evidence for metalloproteinase and metalloproteinase inhibitor imbalance in human osteoarthritic cartilage. *J Clin Invest* 84:678-685, 1989.
 32. Johansen JS, Williamson MK, Rice JS, Price PA. Identification of proteins secreted by human osteoblastic cells in culture. *J of Bone Miner Res* 7:501-512, 1992.
 33. Uchida M, Shima M, Shimoaka T, Fujieda A, Obara K, Suzuki H, Nagai Y, Ikeda T, Yamato H, Kawaguchi H. Regulation of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMPs) by bone resorptive factors in osteoblastic cells. *J Cell Physiol* 185:207-214, 2000.
 34. Whal LM, Olsen SE, Sandberg AL, Mergenhagen SE. Prostaglandin regulation of macrophage collagenase production. *Proc Natl Acad Sci USA* 74:4955-4958, 1977.
 35. Buck RA, Cai J, Eleazer PD, Staat RH, Hurst HE. Detoxification of endotoxin by endodontic irrigants and calcium hydroxide. *J Endod* 27:325-327, 2001.

국문초록

Prevotella nigrescens lipopolysaccharides로 자극된 MG63 세포에서 분비되는 기질금속단백질 MMP-1과 TIMP-1의 수준에 관한 연구

양원경¹ · 김미리² · 손원준¹ · 이인복¹ · 조병훈¹ · 엄정문¹ · 손호현^{1*}

서울대학교 치과대학 치의학과 보존학교실¹, 아산 병원 보존과²

본 연구의 목적은 *Prevotella nigrescens* lipopolysaccharides (LPS)로 자극된 MG63 osteosarcoma 세포에서 생성, 분비되는 기질금속단백효소인 MMP-1과 그 억제제인 TIMP-1을 측정할 뿐아니라 수산화칼슘으로 처리한 *P. nigrescens* LPS에 의한 기질금속단백효소와 그 억제제의 분비수준의 변화를 알아보는데 있다. 혐기성 조건에서 배양한 *P. nigrescens*로부터 LPS를 추출하여 순수정제한 다음 0, 1 그리고 10 $\mu\text{g}/\text{ml}$ 의 LPS 농도로 MG63 세포를 자극하거나 또는 수산화칼슘으로 처리한 10 $\mu\text{g}/\text{ml}$ 의 LPS로 세포를 다양한 자극하여 다양한 시간이 경과한 다음 세포로부터 분비되는 MMP-1과 TIMP-1의 RNA 수준을 real time-PCR 방법으로 측정하였다. 실험결과 MMP-1의 mRNA 수준은 48시간에서 최고에 달하였고 그 분비정도는 LPS의 농도에 비례하였다. TIMP-1 mRNA는 1 $\mu\text{g}/\text{ml}$ 의 세균성 LPS 자극시 24시간 및 48시간에서 높은 증가를 보였으나 고농도인 10 $\mu\text{g}/\text{ml}$ 의 LPS로 자극한 경우 오히려 그 발현이 억제되었다. 또한 수산화칼슘으로 전처리한 *P. nigrescens* LPS로 자극한 MG 63 세포에서는 MMP-1과 TIMP-1의 분비가 억제되었다. 이러한 결과를 통해 볼 때 *P. nigrescens* LPS에 의한 MMP-1과 TIMP-1의 발현조절이 치근단 질환에서 발생하는 치조골 흡수 기전중 하나로 사료된다. 뿐만 아니라 *P. nigrescens*에 의해 분비되는 기질금속단백효소를 매개로 하는 염증반응 감소에 수산화칼슘이 효과적으로 작용하는것으로 확인되어 치근단 질환에 관여하는 세균성 LPS를 제거하기 위해 임상적으로 사용되는 근거가 될 수 있다.