



Neutralizing Antibodies Against Interferon-Beta in Korean Patients with Multiple Sclerosis

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Background and Purpose Patients treated with interferon-beta (IFN- β) can develop neutralizing antibodies (NAbs) against IFN- β that can negatively affect the therapeutic response. This study assessed the prevalence of NAbs and the impact of NAb positivity on the therapeutic response to IFN- β in Korean patients with multiple sclerosis (MS).

Methods This was a multicenter study involving 150 MS patients from 9 Korean medical centers who were treated with IFN- β for at least 6 months. Sera that had not been influenced by acute treatment were assessed for NAbs using a luciferase reporter gene assay. To evaluate the association between persistent positivity for NAbs and disease activity, NAbs were tested at 2 different time points in 75 of the 150 patients. Disease activity was defined as the presence of clinical exacerbations and/or active MRI lesions during a 1-year follow-up after NAb positivity was confirmed.

Results NAbs were found in 39 of the 150 (26%) MS patients: 30 of the 85 (35%) who were treated with subcutaneous IFN- β -1b, 9 of the 60 (15%) who were treated with subcutaneous IFN- β -1a, and 0 of the 5 (0%) who were treated with intramuscular IFN- β -1a. Thirty of the 39 patients exhibiting NAb positivity were tested at different time points, and 20 of them exhibited persistent NAb positivity. Disease activity was observed more frequently in patients with persistent NAb positivity than in those with transient positivity or persistent negativity [16/20 (80%) vs. 4/55 (7%), respectively; $p < 0.001$]. When disease activity was compared between patients with persistent and transient NAb positivity, the difference was unchanged and remained statistically significant [16/20 (80%) vs. 2/10 (20%), $p = 0.004$].

Conclusions These results further support that persistent NAb positivity is associated with disease activity in MS patients treated with IFN- β .

Key Words multiple sclerosis, neutralizing antibody, disease modifying treatment.

INTRODUCTION

Interferon-beta (IFN- β) is one of the established first-line therapies for multiple sclerosis (MS). However, not all patients with MS experience satisfactory treatment response.¹ Some patients appear to be influenced by the presence of neutralizing antibodies (NAbs) against IFN- β , which has been associated with the occurrence of clinical relapse(s) and/or progression of Expanded Disability Status Scale (EDSS) scores in previous studies, and up to 40% of all patients treated with IFN- β become persistently positive for NAbs.¹⁻⁶ In addition, several molecules-including C-X-C motif chemokine ligand 10 (CXCL10) and soluble tumor-necrosis-factor-related apoptosis-inducing ligand (sTRAIL)-have been reported to be induced by IFN- β and exhibit decreased levels when NAbs are present.^{7,8}

Because diverse therapeutic options are now available for patients with MS, the timely identification of factors that can affect the efficacy of specific first-line agents is vital to op-

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optimize personalized therapeutic strategies.⁹⁻¹¹ Data regarding NABs against IFN- β and IFN- β biomarkers have not been evaluated previously in Korean MS patients. Accordingly, we aimed to investigate the prevalence of NABs against IFN- β and the association between treatment response and persistent NAB positivity. We also evaluated the association between NABs and IFN- β biomarkers in Korean MS patients treated with IFN- β .

METHODS

Patients who fulfilled the 2010 McDonald criteria¹² and clinically isolated syndrome (CIS) suggestive of MS were enrolled. They had received regular treatment with IFN- β (subcutaneous IFN- β -1a 44 mcg thrice weekly; IFN- β -1b 250 mcg every other day; or intramuscular IFN- β -1a 30 mcg weekly) for at least 6 months. Serum samples collected from nine referral hospitals in Korea from 2010 to 2016 were stored at -80°C until analysis. Sera from patients who were treated with corticosteroids within the previous 1 month or with plasma exchange within the previous 3 months were excluded. Serum samples that were available from 75 patients at 2 different time points were analysed for persistent positivity (PP) of NABs. The medical records of these patients were analysed to verify the association between treatment response and persistent NAB positivity. Demographic and clinical information was collected from the participating centers.

NABs were measured using a commercially available luciferase reporter gene assay (iLite, Galway, Ireland), while IFN- β biomarkers (CXCL10 and sTRAIL) were assessed using ELISA kits (R&D Systems, Minneapolis, MN, USA) according to the manufacturer's instructions.

Disease activity was defined as the presence of at least one active MRI lesion [new or enlarging lesion(s) on T2-weighted or fluid-attenuation-inversion-recovery imaging, or gadolinium-enhanced lesions on T1-weighted imaging], and/or the occurrence of clinical relapses during a 1-year follow-up from the confirmation of NAB positivity.¹³

The chi-squared or Fisher's exact test was used to compare patients with and without NABs. Student's *t*-test or the Mann-Whitney U test was performed to compare continuous variables of IFN- β biomarkers, with $p < 0.05$ considered to be statistically significant.

The Institutional Review Board of National Cancer Center approved the study protocol (NCC 2015-0032), and written informed consent was obtained from all patients.

RESULTS

Demographics

In total, 150 MS patients were recruited from 9 medical centers in Korea; their demographic information is summarized in Table 1. The male-to-female ratio was 2:3, and the median age at sampling was 38 years (range 14–64 years). The median disease duration was 8 years (range 2–29 years), and the median duration of IFN- β treatment was 52 months (range 7–190 months). A total of 146 patients fulfilled the 2010 McDonald criteria, and 2 of them had primary progressive MS. Remained 4 patients had CIS suggestive of MS. The median EDSS score at the last visit was 1.5 (range 0.0–8.0). Of the 150 MS patients in this study, subcutaneous IFN- β -1b, IFN- β -1a, or intramuscular IFN- β -1a was used in 85, 60, and 5 individuals, respectively.

Prevalence of NABs in Korean patients with MS

Among 150 Korean MS patients, 226 tests for NABs were performed, with 39 (26%) patients exhibiting NAB positivity: 30 of the 85 (35%) who were treated with subcutaneous IFN- β -1b, 9 of the 60 (15%) who were treated with subcutaneous IFN- β -1a, and 0 of the 5 (0%) who were treated with intramuscular IFN- β -1a (Fig. 1).

Table 1. Demographics

Males:females	60:90
Current age (years)	38 (14–64)
Disease duration (years)	8 (2–29)
Current EDSS score	1.5 (0.0–8.0)
Clinically isolated syndrome: relapsing MS: progressive MS	4:144:2
Median duration of IFN- β therapy (months)	52 (7–190)
sc IFN- β -1b: sc IFN- β -1a: IM IFN- β -1a	85:60:5

Data are *n* or median (range) values.

EDSS: Expanded Disability Status Scale, IFN- β : interferon-beta, IM: intramuscular, MS: multiple sclerosis, sc: subcutaneous.

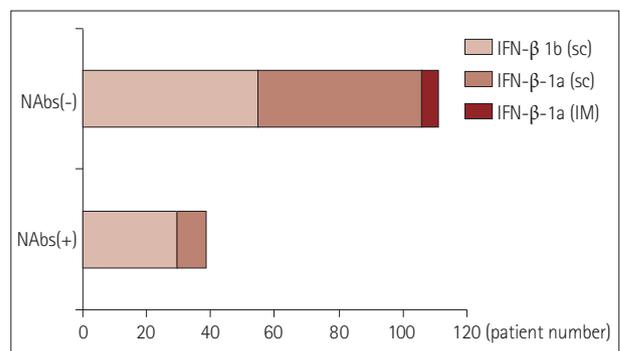


Fig. 1. NABs positivity rates according to different preparations of IFN- β . IFN- β : interferon-beta, IM: intramuscular, NABs: neutralizing antibodies, sc: subcutaneous.

Influence of duration of IFN-β therapy

Fig. 2 demonstrates the influence of treatment duration. In the 226 single tests, the positivity rate for NABs was highest in patients with treatment durations of 12–23 months [22/64 (34.4%)], followed by durations of 24–35 months [15/44 (34.1%)], >35 months [19/85 (22.4%)], and <12 months [3/33 (9.1%)]. The positivity rate for NABs in patients treated with subcutaneous IFN-β-1b was highest for those treated for 12–23 months (45%), whereas for subcutaneous IFN-β-1a it was highest for those treated for 24–35 months (22.2%).

Association between persistent positivity of NABs and disease activity

Seventy-five patients with MS were tested for NABs at 2 different time points at a median interval of 9 months (range 1–102 months), with 20 (26.7%) exhibiting disease activity. Of 39 single-positive NABs patients, 30 were tested for NABs at different time points, and 20 patients showed PP of NABs. Of these 20 patients, 17 were treated with subcutaneous IFN-β-1b, and 3 with subcutaneous IFN-β-1a.

Disease activity was observed more frequently in PP patients than in those with transient positivity (TP) or persistent negativity (PN) for NABs [16/20 (80%) vs. 4/55 (7%), $p<0.001$] (Table 2). When disease activity was compared between PP and TP patients, the difference was essentially un-

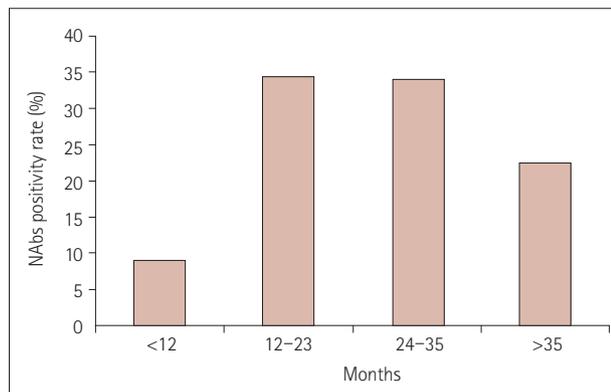


Fig. 2. NABs positivity rates (%) according to the duration of interferon-beta therapy. NABs: neutralizing antibodies.

changed and remained statistically significant [16/20 (80%) vs. 2/10 (20%), respectively; $p=0.004$].

In a sub-analysis of subcutaneous IFN-β-1b treatment, disease activity was observed more frequently in PP than in TP or PN patients [13/17 (76.5%) vs. 3/34 (8.8%), respectively; $p<0.001$]. When disease activity was compared between PP and TP patients, the difference was unchanged and remained statistically significant [13/17 (76.5%) vs. 1/7 (14.3%), respectively; $p=0.009$]. Among patients treated with subcutaneous IFN-β-1a, disease activity was observed more frequently in PP than in TP or PN individuals [3/3 (100%) vs. 1/21 (4.8%), respectively; $p=0.002$]. In these patients, disease activity was observed in 100% (3/3) of PP but 33% (1/3) of TP individuals ($p=0.4$).

NABs and IFN-β biomarkers

Among 90 serum samples from 45 PN patients and 40 serum samples from 20 PP patients, an available total of 114 (79 PN and 35 PP) sera were tested for both CXCL10 and sTRAIL. The level of IFN-β-inducible biomarkers did not differ significantly between PN and PP patients: median 156 pg/mL (range 12–2,636 pg/mL) vs. 97.3 pg/mL (10–1,317 pg/mL), respectively ($p=0.156$), for CXCL10; and median 100 pg/mL (range 2–1,522 pg/mL) vs. 88 pg/mL (27–1,909 pg/mL), respectively ($p=0.491$), for sTRAIL (Fig. 3). Excluding three samples from PN patients and seven from PP patients with clinical relapse status, similar results were observed {CXCL10 [median 148.3 pg/mL (range 12–2,636 pg/mL) vs. 122.7 pg/mL (20–1,317 pg/mL), respectively; $p=0.349$] and sTRAIL [median 98 pg/mL (range 2–1,522 pg/mL) vs. 92.5 pg/mL (27–1,909 pg/mL), respectively; $p=0.311$]}.

DISCUSSION

Twenty-six percent of Korean MS patients in our cohort exhibited NAB positivity: 35% for subcutaneous IFN-β-1b, 15% for subcutaneous IFN-β-1a, and 0% for intramuscular IFN-β-1a. The positivity rate for NABs was highest (34.4%) in patients with treatment durations of 12–23 months. Disease

Table 2. Status of positivity for NABs and disease activity

Status of NABs positivity	Disease activity during after 1 year of IFN-β (clinical relapses and/or active MRI lesions)	p-value
Total 75 tested at 2 different time points, n (%)		
Persistent positive (n=20)	16 (80)	<0.001
Transient positive or persistent negative (n=55)	4 (7)	
Total 30 tested at 2 different time points, n (%)		
Persistent positive (n=20)	16 (80)	0.004
Transient positive (n=10)	2 (20)	

IFN-β: interferon-beta, NABs: neutralizing antibodies.

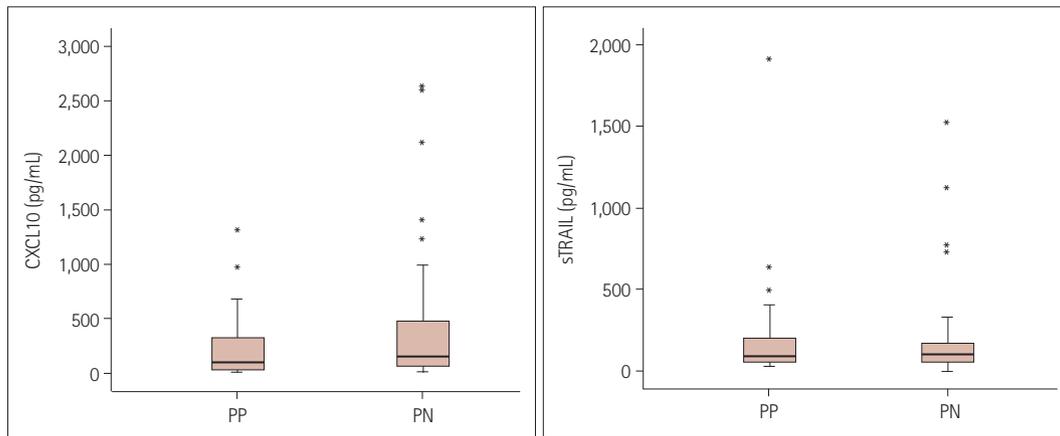


Fig. 3. Levels of CXCL10 and sTRAIL according to the NAb status. CXCL10: C-X-C motif chemokine ligand 10, NAb: neutralizing antibody, PN: persistent negativity for NAb, PP: persistent positivity for NAb, sTRAIL: soluble tumor-necrosis-factor-related apoptosis-inducing ligand.

activity was present in 80% of PP patients, whereas only 7% of TP or PN patients exhibited disease activity ($p < 0.001$). The difference between PP and TP patients remained statistically significant in the presence of disease activity (80% vs. 20%, respectively; $p = 0.004$).

The frequency of NAb positivity in previous studies has varied due to differences in NAb assay methodologies, varying durations of IFN- β treatment, and diverse cohort characteristics. The reported ranges for the rate of NAb positivity have generally been 27–53% in patients treated with subcutaneous IFN- β -1b, 15–35% in those treated with subcutaneous IFN- β -1a, and 2–19% in those treated with intramuscular IFN- β -1a.^{14,15} Our results are comparable with those of previous studies investigating subcutaneous IFN- β -1b and IFN- β -1a, which have been prescribed in Korea since 1998 and 2000, respectively. The prescription of intramuscular IFN- β -1a for MS patients in Korea was approved very recently (2013); therefore, only five patients undergoing this particular treatment could be enrolled in the current study; none of them exhibited NAb positivity.

Cytopathic effect assays to measure myxovirus resistance protein A (MxA), which is stimulated by IFN- β , have been applied in previous investigations.^{4,16} ELISA for MxA protein or real-time polymerase chain reaction for MxA messenger RNA have also been used previously.^{17,18} However, the current study used a luciferase reporter gene assay because this is less time-consuming and more cost-efficient, and has demonstrated lower inter-laboratory variability compared with the previously used methods.^{14,19,20}

The emphasis on therapeutic monitoring of NAb in the decision-making process for switching therapy differs between guidelines from Europe and North America, but an association between NAb and a reduction in therapeutic efficacy of IFN- β is generally recognized.^{21–25} The European guidelines recommend that patients treated with IFN- β should be tested

for NAb after 1 and 2 years of treatment, and altering therapy should be considered when high-titer PP for NAb in repeated measurements at 3 to 6 months was observed.^{21–24} Due to the retrospective design of the current study, we could not unify the time points for testing NAb. Nevertheless, we observed that the positivity rate for NAb was only 9.1% after <1 year of IFN- β therapy, but >34% for a therapeutic period of 1 to 3 years. More importantly, we reconfirmed that PP for NAb was associated with disease activity in MS patients treated with IFN- β therapy. Repeated measurement of NAb may therefore represent an additional indicator that could promote optimal therapeutic decision-making in Korean patients with MS.

CXCL10 is involved in the recruitment of leukocytes and sTRAIL inhibits autoreactive and antigen-specific T cells, and previous studies have found that the levels of these IFN- β -inducible biomarkers were diminished in the presence of NAb.^{7,8,26,27} A recent study found CXCL10 and sTRAIL to be promising potential biomarkers for the response to IFN- β therapy in a longitudinal follow-up.⁷ Although the mean values of these markers were higher in PN than PP patients in the current study, we did not observe statistically significant differences. This discrepancy could be explained by the limited measurement of IFN- β biomarkers in the only available samples, the uncontrolled timing of assessment due to the retrospective study design. Further prospective longitudinal studies are needed to clarify the clinical implications of IFN- β -inducible biomarkers in Korean MS patients.

The irregular sampling interval of the repeated NAb measurements and the absence of NAb titers were additional limitations of the current study. Nevertheless, the reliable assays used in this real-world, multi-center cohort investigation further confirmed that the proportion of patients exhibiting disease activity was significantly higher in PP patients than in TP or PN patients. In an era of various treatment options

for MS, identifying suboptimal responders to first-line disease-modifying agents is crucial in clinical practice. Integrating the results of NAb testing with clinical and radiological information may be helpful for establishing optimal individualized therapeutic strategies.

Conflicts of Interest

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REFERENCES

1. The IFNB Multiple Sclerosis Study Group. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. I. Clinical results of a multicenter, randomized, double-blind, placebo-controlled trial. *Neurology* 1993;43:655-661.
2. Deisenhammer F. Neutralizing antibodies to interferon-beta and other immunological treatments for multiple sclerosis: prevalence and impact on outcomes. *CNS Drugs* 2009;23:379-396.
3. Sorensen PS, Koch-Henriksen N, Ross C, Clemmesen KM, Bendtzen K; Danish Multiple Sclerosis Study Group. Appearance and disappearance of neutralizing antibodies during interferon-beta therapy. *Neurology* 2005;65:33-39.
4. Rudick RA, Simonian NA, Alam JA, Campion M, Scaramucci JO, Jones W, et al. Incidence and significance of neutralizing antibodies to interferon beta-1a in multiple sclerosis. Multiple Sclerosis Collaborative Research Group (MSCRG). *Neurology* 1998;50:1266-1272.
5. Francis GS, Rice GP, Alsup JC; PRISMS Study Group. Interferon beta-1a in MS: results following development of neutralizing antibodies in PRISMS. *Neurology* 2005;65:48-55.
6. Sorensen PS, Ross C, Clemmesen KM, Bendtzen K, Frederiksen JL, Jensen K, et al. Clinical importance of neutralising antibodies against interferon beta in patients with relapsing-remitting multiple sclerosis. *Lancet* 2003;362:1184-1191.
7. Hegen H, Millonig A, Bertolotto A, Comabella M, Giovannoni G, Guger M, et al. Early detection of neutralizing antibodies to interferon-beta in multiple sclerosis patients: binding antibodies predict neutralizing antibody development. *Mult Scler* 2014;20:577-587.
8. Cepok S, Schreiber H, Hoffmann S, Zhou D, Neuhaus O, von Geldern G, et al. Enhancement of chemokine expression by interferon beta therapy in patients with multiple sclerosis. *Arch Neurol* 2009;66:1216-1223.
9. Wingerchuk DM, Carter JL. Multiple sclerosis: current and emerging disease-modifying therapies and treatment strategies. *Mayo Clin Proc* 2014;89:225-240.
10. Río J, Comabella M, Montalban X. Predicting responders to therapies for multiple sclerosis. *Nat Rev Neurol* 2009;5:553-560.
11. Rudick RA, Polman CH. Current approaches to the identification and management of breakthrough disease in patients with multiple sclerosis. *Lancet Neurol* 2009;8:545-559.
12. Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol* 2011;69:292-302.
13. Hyun JW, Kim SH, Jeong IH, Ahn SW, Huh SY, Park MS, et al. Utility of the rio score and modified rio score in Korean patients with multiple sclerosis. *PLoS One* 2015;10:e0129243.
14. Creeke PI, Farrell RA. Clinical testing for neutralizing antibodies to interferon- β in multiple sclerosis. *Ther Adv Neurol Disord* 2013;6:3-17.
15. Govindappa K, Sathish J, Park K, Kirkham J, Pirmohamed M. Development of interferon beta-neutralising antibodies in multiple sclerosis—a systematic review and meta-analysis. *Eur J Clin Pharmacol* 2015;71:1287-1298.
16. Bertolotto A, Malucchi S, Milano E, Castello A, Capobianco M, Mutani R. Interferon beta neutralizing antibodies in multiple sclerosis: neutralizing activity and cross-reactivity with three different preparations. *Immunopharmacology* 2000;48:95-100.
17. Pachner A, Narayan K, Price N, Hurd M, Dail D. MxA gene expression analysis as an interferon-beta bioactivity measurement in patients with multiple sclerosis and the identification of antibody-mediated decreased bioactivity. *Mol Diagn* 2003;7:17-25.
18. Bertolotto A, Sala A, Caldano M, Capobianco M, Malucchi S, Marnetto F, et al. Development and validation of a real time PCR-based bioassay for quantification of neutralizing antibodies against human interferon-beta. *J Immunol Methods* 2007;321:19-31.
19. Farrell R, Kapoor R, Leary S, Rudge P, Thompson A, Miller D, et al. Neutralizing anti-interferon beta antibodies are associated with reduced side effects and delayed impact on efficacy of interferon-beta. *Mult Scler* 2008;14:212-218.
20. Lam R, Farrell R, Aziz T, Gibbs E, Giovannoni G, Grossberg S, et al. Validating parameters of a luciferase reporter gene assay to measure neutralizing antibodies to IFNbeta in multiple sclerosis patients. *J Immunol Methods* 2008;336:113-118.
21. Sørensen PS, Deisenhammer F, Duda P, Hohlfeld R, Myhr KM, Palace J, et al. Guidelines on use of anti-IFN-beta antibody measurements in multiple sclerosis: report of an EFNS Task Force on IFN-beta antibodies in multiple sclerosis. *Eur J Neurol* 2005;12:817-827.
22. Polman CH, Bertolotto A, Deisenhammer F, Giovannoni G, Hartung HP, Hemmer B, et al. Recommendations for clinical use of data on neutralising antibodies to interferon-beta therapy in multiple sclerosis. *Lancet Neurol* 2010;9:740-750.
23. Bertolotto A, Capobianco M, Amato MP, Capello E, Capra R, Centonze D, et al. Guidelines on the clinical use for the detection of neutralizing antibodies (NAbs) to IFN beta in multiple sclerosis therapy: report from the Italian Multiple Sclerosis Study group. *Neurol Sci* 2014;35:307-316.
24. Hesse D, Sørensen PS. Using measurements of neutralizing antibodies: the challenge of IFN-beta therapy. *Eur J Neurol* 2007;14:850-859.
25. Goodin DS, Frohman EM, Hurwitz B, O'Connor PW, Oger JJ, Reder AT, et al. Neutralizing antibodies to interferon beta: assessment of their clinical and radiographic impact: an evidence report: report of the Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology. *Neurology* 2007;68:977-984.
26. Buttman M, Merzyn C, Rieckmann P. Interferon-beta induces transient systemic IP-10/CXCL10 chemokine release in patients with multiple sclerosis. *J Neuroimmunol* 2004;156:195-203.
27. Lünemann JD, Waiczies S, Ehrlich S, Wendling U, Seeger B, Kamradt T, et al. Death ligand TRAIL induces no apoptosis but inhibits activation of human (auto) antigen-specific T cells. *J Immunol* 2002;168:4881-4888.