



ORIGINAL ARTICLE

Crotonis Fructus Extract Inhibits 12-O-Tetradecanoylphorbol-13-Acetate-Induced Expression of Matrix Metalloproteinase-9 via the Activator Protein-1 Pathway in MCF-7 Cells

Hyun-Kyung Song^{1,*}, Guem-San Lee^{2,*}, Sueng Hyuk Park³, Eun-Mi Noh¹, Jeong-Mi Kim¹, Do-Gon Ryu³, Sung Hoo Jung⁴, Hyun Jo Youn⁴, Young-Rae Lee^{1,5,6}, Kang-Beom Kwon^{1,3}

¹Center for Metabolic Function Regulation, Wonkwang University School of Medicine, Iksan; Departments of ²Herbology and ³Korean Physiology, Wonkwang University School of Korean Medicine, Iksan; ⁴Department of Surgery, Biomedical Research Institute of Chonbuk National University Hospital, Research Institute of Clinical Medicine of Chonbuk National University, Jeonju; ⁵Department of Oral Biochemistry and Institute of Biomaterials-Implant, Wonkwang University School of Dentistry, Iksan; ⁶Integrated Omics Institute, Wonkwang University, Iksan, Korea

Purpose: Metastatic cancers spread from the primary site of origin to other parts of the body. Matrix metalloproteinase-9 (MMP-9) is essential in metastatic cancers owing to its major role in cancer cell invasion. *Crotonis fructus* (CF), the mature fruits of *Croton tiglium* L., have been used for the treatment of gastrointestinal disturbance in Asia. In this study, the effect of the ethanol extract of CF (CFE) on MMP-9 activity and the invasion of 12-O-tetradecanoylphorbol-13-acetate (TPA)-treated MCF-7 cells was examined. **Methods:** The cell viability was evaluated using the 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. The expression of MMP-9 was examined by Western blotting, zymography, and real-time polymerase chain reaction. An electrophoretic mobility gel shift assay was performed to

detect activator protein-1 (AP-1) DNA binding activity and cell invasiveness was measured by an *in vitro* Matrigel invasion assay. **Results:** CFE significantly suppressed MMP-9 expression and activation in a dose-dependent manner. Furthermore, CFE attenuated the TPA-induced activation of AP-1. **Conclusion:** The results indicated that the inhibitory effects of CFE against TPA-induced MMP-9 expression and MCF-7 cell invasion were dependent on the protein kinase C δ /p38/c-Jun N-terminal kinase/AP-1 pathway. Therefore, CFE could restrict breast cancer invasiveness owing to its ability to inhibit MMP-9 activity.

Key Words: *Crotonis fructus*, Matrix metalloproteinase 9, MCF-7 cells, Neoplasm invasiveness, Transcription factor AP-1

INTRODUCTION

Breast cancer is the most common cancer and the second most common cause of cancer-related death in women [1]. Despite the successful eradication of the primary tumor, cancer can still occur in other areas of the body; therefore breast cancer metastasis is the main cause of death in breast cancer patients [2]. Invasion, which is the initial step in cancer metastasis, allows the tumor to invade into the surrounding tissue and metastasize. Thus, it is a popular target in the study of

metastasis [3].

The degradation of the extracellular matrix, in which matrix metalloproteinases (MMPs) play a key role, is a critical step during cell invasion [4]. MMPs are a family of zinc endopeptidases, of which 26 members have been discovered. Among these, MMP-9 can degrade type IV collagen, which is a major structural component of basement membranes; thus, the control of MMP-9 expression is thought to be important in the regulation of metastasis [5,6].

Crotonis fructus (CF) is the mature fruit of *Croton tiglium* L., which is a leafy shrub native to Southeast Asia. CF has been used to attenuate gastrointestinal diseases such as constipation, visceral pain, and intestinal inflammation [7,8]. Iso-guanosine from *C. tiglium* L. was shown to possess antitumor activity against implanted S-180 ascitic tumors in mice [9]. However, it has remained unclear whether CF extract (CFE) inhibited the invasion of breast cancer cells.

In the present study, the effects of CFE on 12-O-tetradec-

Correspondence to: Kang-Beom Kwon

Department of Korean Physiology, Wonkwang University School of Korean Medicine, 460 Iksan-daero, Iksan 54538, Korea
Tel: +82-63-850-6917, Fax: +82-63-850-7324
E-mail: dession@wku.ac.kr

*These authors contributed equally to this work.

The present study was supported by Wonkwang University in 2015.

Received: December 9, 2016 Accepted: July 5, 2017

anoylphorbol-13-acetate (TPA)-induced MCF-7 cell invasion were evaluated through measurements of cell invasiveness and MMP-9 activity. In addition, the signaling mechanism was investigated.

METHODS

Cells and materials

MCF-7 cells were obtained from the American Type Culture Collection (Manassas, USA). The cells were grown in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 1% antibiotics. TPA and anti- β -actin antibodies were obtained from Sigma-Aldrich (St. Louis, USA). FBS, high-glucose DMEM, and phosphate-buffered saline were obtained from Gibco-BRL (Gaithersburg, USA).

Preparation of *crotonis fructus* extract

The mature fruits of *C. tiglium* L. were purchased from Kwangmyungdang Medicinal Herbs Co., Ltd. (Ulsan, Korea) and authenticated by professor G.S.L. The fruits were powdered and extracted with 70% aqueous ethanol under reflux for 2 hours. The solvent was removed by evaporation and the extract was freeze-dried. The yield of the final extract was 11.48% (w/w).

Cell viability assay

The effect of CFE on MCF-7 cell viability was determined by using EZ-Cytox Enhanced Cell Viability Assay Kit (DOGEN, Seoul, Korea) in accordance with the manufacturer's instructions. Briefly, 3×10^4 cells/well were treated with 1, 5, 10, 30, 50, or 100 $\mu\text{g}/\text{mL}$ CFE for 24 hours. Thereafter, EZ-cytox was added (10 $\mu\text{L}/\text{well}$), followed by an incubation period of 30 minutes at 37°C. The absorbance at 450 nm was measured by using an ELISA reader (Sunrise™; Tecan, Männedorf, Switzerland).

Western blot analysis

The cells were incubated for 24 hours with TPA, which was followed by treatment with 10 or 30 $\mu\text{g}/\text{mL}$ CFE for 1 hour. The cells were lysed with ice-cold radioimmunoprecipitation assay (RIPA) buffer (Thermo, Rockford, USA), and the protein lysates (20 μg) were resolved by sodium dodecyl sulfate-polyacrylamide gel electrophoresis. The separated proteins were transferred to polyvinylidene difluoride (PVDF) membranes using Western blotting apparatus. The membranes were blocked for 2 hours with 2% bovine serum albumin or 5% skim milk and then incubated with 1 $\mu\text{g}/\text{mL}$ primary antibody overnight at 4°C. The primary antibodies used for detection were: protein kinase C (PKC) α , PKC β , PKC δ , p38, phos-

phorylated p38 (p-p38), c-Jun N-terminal kinase (JNK), p-JNK, extracellular signal-regulated kinase (ERK), p-ERK, p-c-Jun (Cell Signaling Technology, Beverly, USA), sodium potassium ATPase (Na-K ATPase) (Abcam, Cambridge, UK), MMP-9, p50, p65, and proliferating cell nuclear antigen (PCNA) (Santa Cruz Biotechnology, Dallas, USA). Horseradish peroxidase conjugated IgG was used as a secondary antibody and the protein expression levels were evaluated by signal analysis using a MINI HD6 image analyzer (Uvitec, Cambridge, UK).

Gelatin zymography assay

First, the medium was collected and mixed with sample buffer. The gel for electrophoresis contained 0.1% (w/v) gelatin. The gel was washed with 2.5% Triton X-100 solution at 18°C to 23°C (room temperature) for 30 minutes and then incubated at 37°C in 5 mM CaCl_2 , 0.02% Brij, and 50 mM Tris-HCl (pH 7.5) for 16 hours. The gel was stained using Coomassie brilliant blue solution and then photographed using an image analyzer (Fujifilm, Tokyo, Japan). Proteolysis was identified by the presence of white zones in dark blue fields. Densitometric analysis was performed by using Multi Gauge Software for Image Analysis (Fujifilm).

Quantitative real-time polymerase chain reaction

Total RNA was isolated using TRIzol® Reagent (Life Technologies, Grand Island, USA). The cDNA was synthesized from 1 μg total RNA by using the PrimeScript™ RT reagent kit (TaKaRa, Shiga, Japan). The RNA expression levels of MMP-9 and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) were determined using the ABI PRISM 7900 sequence detection system and SYBR® Green (Applied Biosystems, Foster City, USA). The following primers (2 μM) were used: MMP-9 (NM 004994), sense 5'-CCTGGAGACCTGAGAACCAATCT-3', antisense 5'-CCACCCGAGTGTAACCATAGC-3'; and GAPDH (NM 002046), sense 5'-ATGGAAA TCCCATCACCATCTT-3', antisense 5'-CGCCCCACTGATTTTGG-3'.

Preparation of nuclear extract

The cells were treated with CFE in the presence or absence of TPA for 3 hours and then pelleted by centrifugation at 1,800 $\times g$ for 4 minutes. The isolation of the cytoplasmic and nuclear extracts was performed using the NE-PER® Nuclear and Cytoplasmic Extraction Reagents (Thermo Scientific, Rockford, USA).

Electrophoretic mobility gel shift assay

Affinity purification of the MCF-7 nuclear protein was per-

formed as described above. The MCF-7 nuclear protein was incubated at room temperature in 10 mM Tris-HCl, 2.5% (v/v) glycerol, 5 mM MgCl₂, 50 ng/μL poly (dI-dC), and 0.05% (v/v) Nonidet P-40 binding buffer, pH 7.5. Then, 1 ng of activator protein-1 (AP-1) electrophoretic mobility gel shift assay (EMSA) probe was added (Panomics Inc., Fremont, USA). After incubation for 20 minutes at room temperature, the samples were resolved on a 4% polyacrylamide gel in 0.5 × Tris-borate buffer. The proteins were electrotransferred onto a 0.45-μm nylon membrane (Hybond-N+; Amersham, UK) at 100 V for 45 minutes at 4°C and then crosslinked to the membrane with CL-1000 ultraviolet crosslinker (UVP Inc., Upland, USA). The blots were developed by using a Chemiluminescent Nucleic Acid Detection Module Kit (Thermo Scientific).

Membrane fractionation

MCF-7 cells (5×10^7) were pretreated with 30 μg/mL CFE for 1 hour and then incubated with TPA for 1 hour at 37°C. The membrane proteins were prepared from cells using the MEM-PER™ plus kit (Thermo Scientific).

Invasion assay

The invasion assay was conducted in 24-well chamber dishes (8-μm pore size) coated with 20 μL of Matrigel that was rehydrated in 0.5 mL DMEM for 30 minutes prior to the experiments. Cells (2×10^5) and chemoattractant (conditioned medium; 0.5 mL) were added to the upper and lower chamber of the dishes, respectively. After incubation, the invading cells were fixed and stained with a Toluidine blue solution. The analyzed data were presented as the mean ± standard error from three individual experiments performed in triplicate.

Statistical analyses

The statistical analyses of data were performed using analysis of variance and Duncan test using the Microsoft 2010 Excel program (Microsoft Corp., Redmond, USA). Values of $p < 0.05$ were considered statistically significant.

RESULTS

Effect of *crotonis fructus* extract on MCF-7 cell viability

To evaluate the cytotoxicity of CFE on MCF-7 cells, the viability of untreated and CFE-treated cells was compared. At all tested concentrations, CFE treatment for 24 hours did not result in any significant changes in cell viability. CFE also did not affect MCF-7 cell proliferation at 100 μg/mL for up to 72 hours (Figure 1A and Supplementary Figure 1A, available online). Thus, the optimal nontoxic concentrations (10 and 30 μg/mL) of CFE were used for all subsequent experiments.

Effect of *crotonis fructus* extract on TPA-induced MMP-9 expression/secretion in MCF-7 cells

To examine the effect of CFE on TPA-induced MMP-9 expression, MCF-7 cells were analyzed by Western blot analysis, real-time quantitative polymerase chain reaction (RT-qPCR), and gelatin zymography. Western blot analysis revealed that CFE treatment blocked the upregulation of TPA-induced MMP-9 protein expression in MCF-7 cells. Zymography was performed to determine the effects of CFE on TPA-induced MMP-9 secretion and showed that TPA increased MMP-9 secretion in MCF-7 cells. However, CFE significantly diminished TPA-induced MMP-9 secretion (Figure 1B). RT-qPCR revealed that TPA induced an increase in MMP-9 expression levels in MCF-7 cells, whereas CFE blocked TPA-induced MMP-9 upregulation in a dose-dependent manner (Figure 1C). These results indicated that CFE exerted potent inhibitory effects against the TPA-induced upregulation of MMP-9 expression in MCF-7 cells.

Effect of *crotonis fructus* extract on TPA-induced PKC activation in MCF-7 cells

To determine the effects of CFE on TPA-induced PKC activation, we analyzed the PKC isotype levels in the membrane fraction. CFE inhibited the membrane localization of PKCδ induced by TPA (Figure 2), which suggested that CFE inhibited TPA-induced MMP-9 expression via the suppression of PKCδ activation in MCF-7 cells.

Effect of *crotonis fructus* extract on TPA-induced MAPK activation in MCF-7 cells

To further understand the signaling pathway involved in the inhibitory action of CFE on TPA-induced MMP-9 activation, we used Western blotting to evaluate the effect of CFE on the TPA-induced activation of mitogen-activated protein kinase (MAPK). The treatment with CFE showed inhibitory effects on p38 and JNK, but not on ERK (Figure 3A). These results suggested that the p38 and JNK pathways were involved in the suppression of TPA-induced MMP-9 expression by CFE.

Effect of *crotonis fructus* extract on TPA-induced activation of NF-κB and AP-1 in MCF-7 cells

It has been reported that the MMP-9 promoter has nuclear factor-κB (NF-κB) and AP-1 binding sites; these are known to be involved in MMP-9 gene induction by TPA [10,11]. To clarify the transcription mechanism by which CFE inhibited MMP-9 expression, the effect of CFE on the TPA-induced activation of NF-κB and AP-1 was evaluated using Western blotting. As shown in Figure 3B, TPA upregulated the expres-

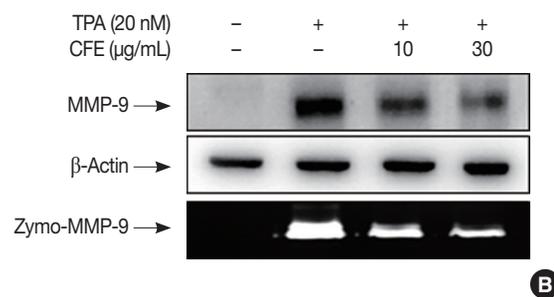
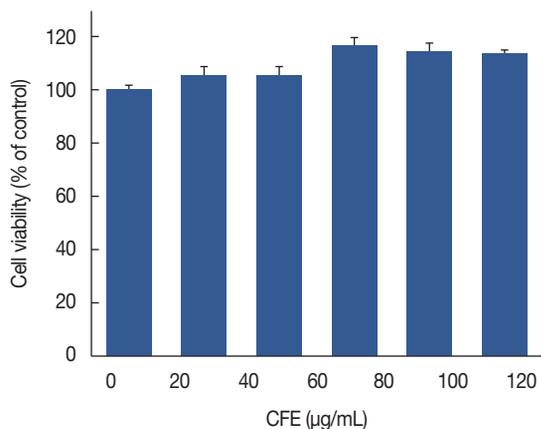


Figure 1. CFE inhibits TPA-induced MMP-9 expression in MCF-7 cells. (A) To assess the cytotoxicity of CFE, cells were treated with various concentrations of CFE for 24 hours. An EZ-cytox enhanced cell viability assay kit was used to detect the. (B) CFE inhibits TPA-induced MMP-9 expression in MCF-7 cells. MCF-7 cells grown in monolayer culture were treated with the indicated CFE concentrations in the presence of TPA for 24 hours. Cell lysates were analyzed by Western blot with anti-MMP-9 antibody and β-actin as a loading control. Conditioned medium was prepared and used for gelatin zymography (Zymo) to assess the effect of CFE on MMP-9 activity in MCF-7 cells. Cells were pretreated with CF for 1 hour and then stimulated with TPA for 24 hours. (C) MMP-9 mRNA levels were analyzed by RT-qPCR, and GAPDH was used as an internal control. Values are shown as mean ± SEM of three independent experiments.

CFE = *Crotonis fructus* extract; TPA = 12-O-tetradecanoylphorbol-13-acetate; MMP-9 = matrix metalloproteinase-9; Zymo = zymography; RT-qPCR = real-time quantitative polymerase chain reaction; GAPDH = glyceraldehyde 3-phosphate dehydrogenase; SEM = standard error of the mean.

**p* < 0.01 vs. TPA.

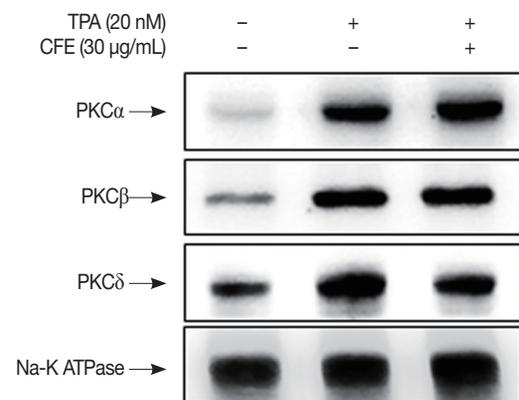
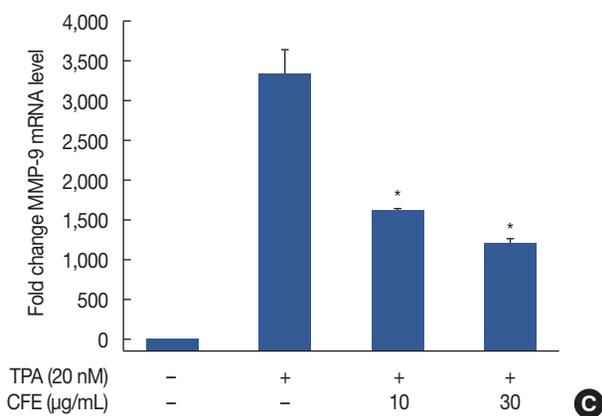


Figure 2. CFE inhibits TPA-induced activation of PKCδ. MCF-7 cells were pretreated with CFE for 1 hour and then with TPA for 1 hour. Western blot analysis was performed to detect the levels of PKCα, PKCβ, PKCδ, and Na-K ATPase as a loading control in the membrane fractions.

CFE = *Crotonis fructus* extract; TPA = 12-O-tetradecanoylphorbol-13-acetate; PKC = protein kinase C.

sion of p65 and p50 (NF-κB subunits) and p-c-Jun (AP-1 subunit) in the nuclear fraction. The pretreatment of cells with

CFE resulted in the inhibition of TPA-induced p-c-Jun expression, but not that of NF-κB activation. Additionally, the DNA binding activity of AP-1 was confirmed. The treatment with CFE suppressed TPA-induced AP-1 DNA binding activity (Figure 3C). These results indicated that CFE specifically blocked AP-1 activation in MCF-7 cells.

Effect of *crotonis fructus* extract on TPA-induced cell invasion in vitro

The upregulation of MMP expression is known to contribute significantly to cancer invasion [12,13]. In this study, we investigated the inhibitory effects of CFE on the invasive ability of MCF-7 cells induced by TPA by performing a Matrigel invasion assay. The cells pretreated with 20 nM TPA were subsequently incubated with CFE for 24 hours. TPA-induced MCF-7 cell invasion was suppressed by CFE treatment (Figure 4). Additionally, CFE slightly inhibited the migration and invasion of MDA-MB-231 cells, a highly metastatic cell line, at nontoxic concentration (Supplementary Figure 2, available online).

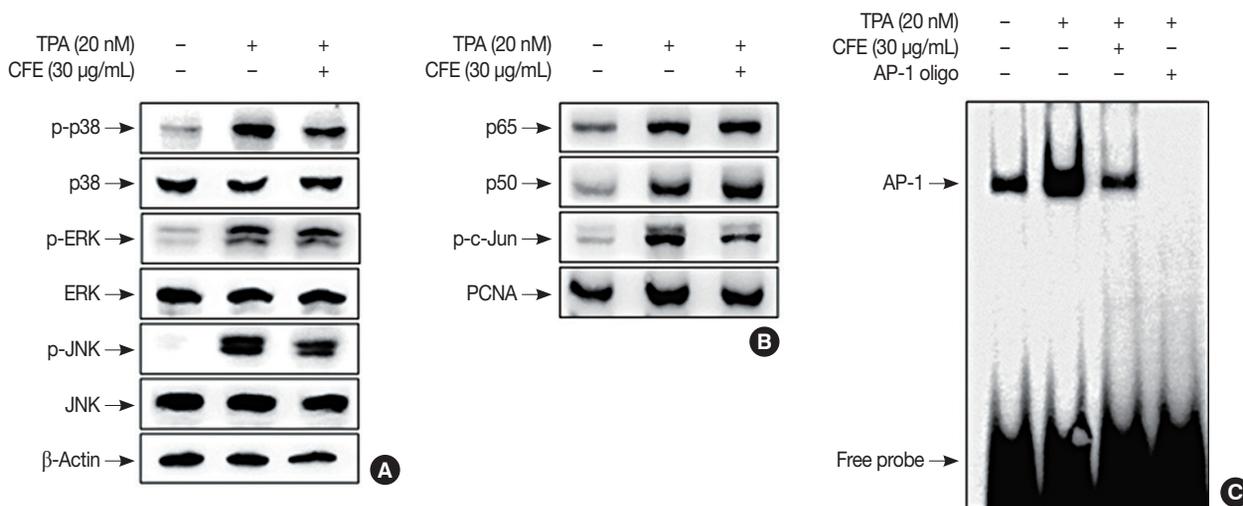


Figure 3. CFE inhibits TPA-induced AP-1 activation in MCF-7 cells. (A) MCF-7 cells were pretreated with CFE for 1 hour and then stimulated with TPA for 15 minutes. The expressions of p-p38, p38, p-JNK, JNK, p-ERK and ERK were analyzed by using Western blotting. (B) Cells were pretreated with CFE for 1 hour before TPA treatment. After stimulation with TPA for 3 hours, the cell nuclear extracts were obtained and subjected to Western blotting to determine the nuclear levels of NF-κB (p50 and p65) and AP-1 (p-c-Jun) subunits. (C) AP-1 DNA binding activity was determined by electrophoretic mobility gel shift assay.

CFE = *Crotonis fructus* extract; TPA = 12-O-tetradecanoylphorbol-13-acetate; AP-1 = activator protein-1; p = phosphorylated; JNK = c-Jun N-terminal kinase; ERK = extracellular signal-regulated kinase; NF-κB = nuclear factor-κB; PCNA = proliferating cell nuclear antigen.

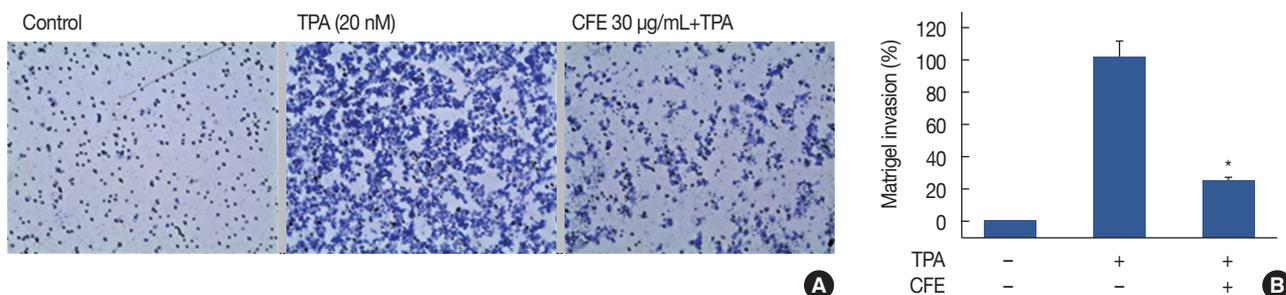


Figure 4. CFE inhibits TPA-induced Matrigel invasion in MCF-7 cells. Cells were seeded in the upper chamber of chamber dishes and were either treated with CFE or left untreated. After 24 hours of incubation, (A) cells on the bottom of the chamber membrane were fixed, stained. (B) The number of counted cells was graphed on the basis of the TPA treatment group. Values represent mean ± SEM of three independent experiments.

CFE = *Crotonis fructus* extract; TPA = 12-O-tetradecanoylphorbol-13-acetate; SEM = standard error of the mean.

*p < 0.01 vs. TPA.

DISCUSSION

The main components of CF, the mature fruit of *C. tiglium* L., are croton oils, which are traditionally used as a laxative or purgative. This oil contains a toxic resin, which is a highly toxic substance that must be treated with great caution. Thus, croton oil can lead to initial skin sloughing, inflammation, and pain [14]. For that reason, the croton oils in CF are removed. Although there are limited studies on the effects of CFE, the effectiveness of CFE for the therapy of peptic ulcers and for the stimulation of lipolysis has been reported [15,16].

MMP-9 is a well-known proteinase that disrupts the extra-

cellular matrix, which induces the invasion of cancer cells; thus, it has been widely studied as a major target of cancer metastasis. In the present study, we found that CFE treatment suppressed the TPA-induced protein, mRNA expression, and secretion of MMP-9 (Figure 1). These results were the first to demonstrate the effect of CFE on MMP-9.

As described above, MMP-9 expression was closely correlated with cancer cell invasion. The present findings showed that CFE blocked TPA-induced MCF-7 cell invasion (Figure 4). Additionally, CFE inhibited TPA-induced MCF-7 migration (Supplementary Figure 1B, available online). As this was the first study to demonstrate the inhibitory activity of CFE

on cell invasion, the mechanism of action was also evaluated. The activation of the three major MAPK families, JNK, ERK, and p38 kinase, is required for MMP-9 induction [17,18]. The MMP-9 gene promoter contains both NF- κ B and AP-1 binding sites. AP-1, composed of Jun and Fos subunits, can be activated by p-JNK [19,20]. As shown in Figures 2 and 3, CFE inhibited the TPA-induced phosphorylation of p38, JNK, and c-Jun. These results showed that CFE suppressed TPA-induced MMP-9 expression through the MAPK/AP-1 pathway in MCF-7 cells.

It was previously reported that PKC isoforms contributed to the TPA-induced invasion and migration of MCF-7 cells, suggesting that the TPA-induced upregulation of MMP-9 expression and cell invasion involved the activation of PKC δ and PKC α [21]. In this study, the translocation of PKC δ into the cell membrane was slightly inhibited by CFE treatment (Figure 2A). These results suggested that CFE possessed the ability to regulate the activation of TPA-responsive PKC isozymes, especially PKC δ , in MCF-7 cells.

This study demonstrated that CFE exhibited anti-invasive activity in TPA-induced MCF-7 cells and suppressed TPA-induced MMP-9 expression via the activation of the PKC δ /p38/JNK/AP-1 pathway. Therefore, the anti-invasive effect of CFE on breast cancer cells at nontoxic concentrations indicated its potential as a chemopreventive agent against breast cancer.

CONFLICT OF INTEREST

The authors declare that they have no competing interests.

REFERENCES

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015;136:E359-86.
2. Yücel B, Bahar S, Kaçan T, Şeker M, Celasun MG, Bahçeci A, et al. Importance of metastasis site in survival of patients with breast cancer. *Austin J Med Oncol* 2014;1:7.
3. Eckhardt BL, Francis PA, Parker BS, Anderson RL. Strategies for the discovery and development of therapies for metastatic breast cancer. *Nat Rev Drug Discov* 2012;11:479-97.
4. Woessner JF Jr. Matrix metalloproteinases and their inhibitors in connective tissue remodeling. *FASEB J* 1991;5:2145-54.
5. Park HL, Ni J, Gerkema FE, Liu D, Belozarov VE, Sang QX. Identification and characterization of human endometase (matrix metalloproteinase-26) from endometrial tumor. *J Biol Chem* 2000;275:20540-4.
6. Mylona E, Nomikos A, Magkou C, Kamberou M, Papassideri I, Keramopoulos A, et al. The clinicopathological and prognostic significance of membrane type 1 matrix metalloproteinase (MT1-MMP) and MMP-9 according to their localization in invasive breast carcinoma. *Histopathology* 2007;50:338-47.
7. Tsai JC, Tsai S, Chang WC. Effect of ethanol extracts of three Chinese medicinal plants with anti-diarrheal properties on ion transport of the rat intestinal epithelia. *J Pharmacol Sci* 2004;94:60-6.
8. Wang X, Lan M, Wu HP, Shi YQ, Lu J, Ding J, et al. Direct effect of croton oil on intestinal epithelial cells and colonic smooth muscle cells. *World J Gastroenterol* 2002;8:103-7.
9. Kim JH, Lee SJ, Han YB, Moon JJ, Kim JB. Isolation of isoguanosine from *Croton tiglium* and its antitumor activity. *Arch Pharm Res* 1994;17:115-8.
10. Eberhardt W, Huwiler A, Beck KE, Walpen S, Pfeilschifter J. Amplification of IL-1 beta-induced matrix metalloproteinase-9 expression by superoxide in rat glomerular mesangial cells is mediated by increased activities of NF-kappa B and activating protein-1 and involves activation of the mitogen-activated protein kinase pathways. *J Immunol* 2000;165:5788-97.
11. Lungu G, Covaldea L, Mendes O, Martini-Stoica H, Stoica G. FGF-1-induced matrix metalloproteinase-9 expression in breast cancer cells is mediated by increased activities of NF-kappaB and activating protein-1. *Mol Carcinog* 2008;47:424-35.
12. Nabeshima K, Inoue T, Shimao Y, Sameshima T. Matrix metalloproteinases in tumor invasion: role for cell migration. *Pathol Int* 2002;52:255-64.
13. Chambers AF, Matrisian LM. Changing views of the role of matrix metalloproteinases in metastasis. *J Natl Cancer Inst* 1997;89:1260-70.
14. Hetter GP. An examination of the phenol-croton oil peel: part I. dissecting the formula. *Plast Reconstr Surg* 2000;105:227-39.
15. Jeon CM, Min GW, Park JH, Yoon CH, Jeong JC, Kang JJ, et al. Effect of the methanol extract of *Dansameum* on the ammonia-induced peptic ulcer in rats. *Korean J Orient Int Med* 2000;21:597-604.
16. Kim MS, Kim HR, So HS, Lee YR, Moon HC, Ryu DG, et al. *Crotonis fructus* and its constituent, croton oil, stimulate lipolysis in OP9 adipocytes. *Evid Based Complement Alternat Med* 2014;2014:780385.
17. Kim KC, Lee CH. MAP kinase activation is required for the MMP-9 induction by TNF-stimulation. *Arch Pharm Res* 2005;28:1257-62.
18. Hong S, Park KK, Magae J, Ando K, Lee TS, Kwon TK, et al. Ascochlorin inhibits matrix metalloproteinase-9 expression by suppressing activator protein-1-mediated gene expression through the ERK1/2 signaling pathway: inhibitory effects of ascochlorin on the invasion of renal carcinoma cells. *J Biol Chem* 2005;280:25202-9.
19. Karin M, Liu Zg, Zandi E. AP-1 function and regulation. *Curr Opin Cell Biol* 1997;9:240-6.
20. Crawford HC, Matrisian LM. Mechanisms controlling the transcription of matrix metalloproteinase genes in normal and neoplastic cells. *Enzyme Protein* 1996;49:20-37.
21. Kim JM, Noh EM, Kwon KB, Kim JS, You YO, Hwang JK, et al. Curcumin suppresses the TPA-induced invasion through inhibition of PKC-alpha-dependent MMP-expression in MCF-7 human breast cancer cells. *Phytomedicine* 2012;19:1085-92.