

Implications of Calcineurin/NFAT Inhibitors' Regulation of Dendritic Cells and Innate Immune Cells in Islet Xenotransplantation

Yong-Hee Kim^{1,2}, Won-Woo Lee^{2,3} and Chung-Gyu Park^{2,3*}

¹Department of Microbiology, Kyungpook National University School of Medicine, Daegu; ²Xenotransplantation Research Center (XRC); ³Department of Microbiology and Immunology, Seoul National University College of Medicine, Seoul, Korea

Calcineurin inhibitors, such as cyclosporine and tacrolimus (FK506) are broadly used in organ transplantations as immune suppressants. As the calcineurin/NFAT signaling pathway has been identified as critical pathway in the interleukin-2 (IL-2) production of T cells, inhibition of T-cell derived IL-2 has been considered the major mechanism of calcineurin inhibitors. However, there is increasing evidence that NFAT transcription factor is involved in multiple functions of dendritic cells and innate immune cells as well. NFAT expression is not restricted to T cells, and IL-2 can be produced in dendritic cells and macrophages through the calcineurin/NFAT pathway. Furthermore, it has been discovered that NFAT regulates expressions of several inflammatory mediators, including TNF- α and cyclooxygenase-2 in innate immune cells. Therefore, calcineurin inhibitors may have much broader effects in the transplant recipients than previously being considered. In this review, we reviewed recently discovered roles of NFAT pathway in dendritic cells and innate immune cells, and discussed positive and negative implications of calcineurin inhibitors' broader effects with a focus on islet xenotransplantation.

Key Words: Calcineurin inhibitors, Innate immunity, Islet transplantation, Xenotransplantation

I. INTRODUCTION

Nuclear factor of activated T cells (NFAT) was originally identified in nuclear extracts of activated T cells as a DNA-binding factor which binds to the interleukin-2 (*IL2*) promoter (1). Later, it was shown that inhibition of NFAT is the primary mechanism of calcineurin inhibitors, including cyclosporine and FK506 (tacrolimus) (2, 3). These potent immune suppressants revolutionized allogeneic organ trans-

plantations, dramatically raising 5-year survival rates of allografts since the introduction of cyclosporine into clinical practice. Before recent discoveries, inhibition of NFAT-mediated IL-2 production in T cells had been considered the principal mechanism of action for the calcineurin inhibitors. However, despite its name, the expression of NFAT is not limited to T cells. It has been shown that NFAT is expressed by almost every cell type, including other cells of the immune system (4~8). Due to the recent discoveries, it is now clear that NFAT has important functions in cells

Received: February 20, 2016/ Accepted: February 22, 2016

*Corresponding author: Chung-Gyu Park. Department of Microbiology and Immunology, Xenotransplantation Research Center, Seoul National University College of Medicine, 103 Daehak-ro Jongno-gu, Seoul, 03080, Korea.

Phone: +82-2-740-8308, Fax: +82-2-743-0881, e-mail: chgpark@snu.ac.kr

**This work was supported by a grant from the Korea Healthcare Technology R&D Project through the Korea Health Industry Development Institute (KHIDI), funded by the Ministry for Health and Welfare, Republic of Korea (Grant No. H113C0954).

©This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>).

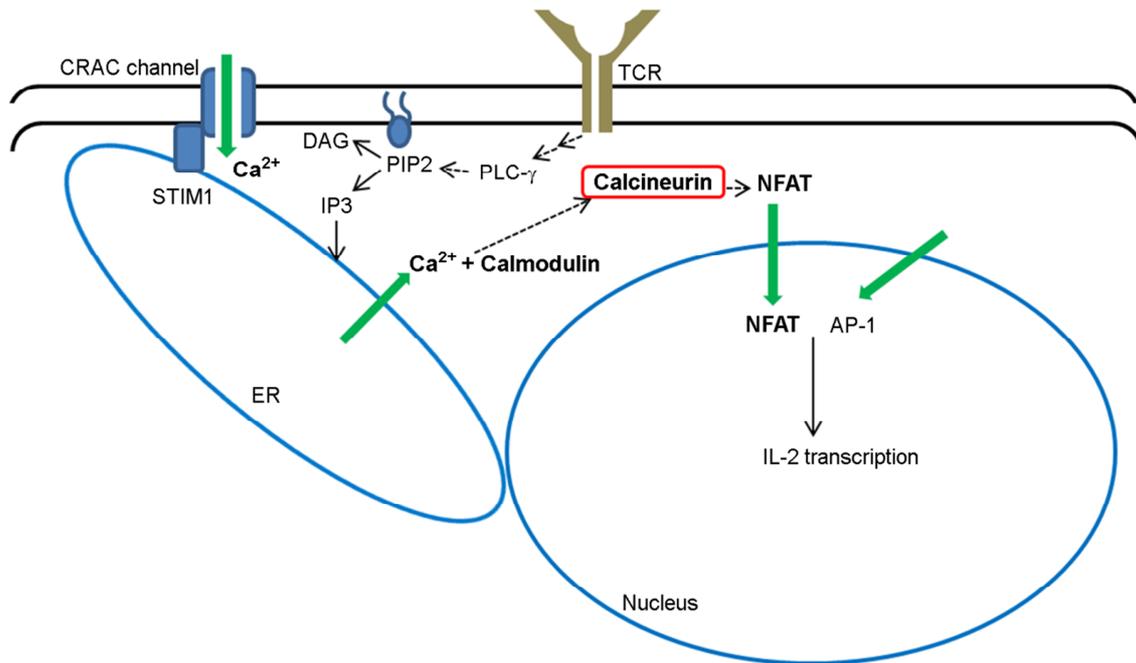


Figure 1. The classical calcineurin/NFAT signaling pathway in T cells

of the innate immune system (9).

During the past two decades, significant progress has been made in the field of xenotransplantation. Especially, successful long-term survival results have been achieved in pre-clinical pig-to-non-human primate islet xenotransplantation (10). Due to successful results in pre-clinical studies, it seems that clinical trials of islet xenotransplantation may take place within a few years (11, 12). Therefore, transplantation of islets is likely to be the first introduction of xenotransplantation into clinic, thereby providing an ultimate solution to the problem of donor organ supply for curing Type I diabetes.

Though the acute rejection of xenogeneic islets is primarily a T cell-dependent process (13), regulations of the innate immunity may bring beneficial effects for achieving long-term graft acceptance in islet xenotransplantation. In this review we overview the actions of calcineurin/NFAT inhibitors in T cells and innate immune cells, and discuss its implications in islet xenotransplantation.

II. The Classical Calcineurin/NFAT Signaling Pathway in T Cells

The classical calcineurin/NFAT signaling pathway in T cells and its inhibition by calcineurin inhibitors can be summarized as follows (Fig. 1). Upon antigen binding to a T cell receptor (TCR), cytosolic phospholipase C- γ (PLC- γ) is recruited to the plasma membrane and becomes activated. Activated PLC- γ catalyzes the hydrolysis of the plasma membrane phospholipid phosphatidylinositol 4,5-bisphosphate (PIP2) into inositol-1,4,5-triphosphate (IP3) and diacylglycerol (DAG). IP3 then binds to specific IP3 receptors on endoplasmic reticulum and results in Ca^{2+} release from endoplasmic reticulum Ca^{2+} stores into the cytoplasm. The depletion of endoplasmic reticulum Ca^{2+} is sensed by an endoplasmic reticulum membrane protein called stromal interacting molecule 1 (STIM1), which triggers opening of plasma membrane Ca^{2+} release-activated Ca^{2+} (CRAC) channels. Subsequently, influx of extracellular Ca^{2+}

further increases the cytosolic Ca^{2+} concentration. Cytosolic free Ca^{2+} then binds to a Ca^{2+} -dependent regulatory protein called calmodulin, and these Ca^{2+} -calmodulin complexes can activate a protein serine/threonine phosphatase called calcineurin. The activated calcineurin dephosphorylates phosphoserines in NFAT, thereby exposing a nuclear localization signal that permits NFAT to translocate from the cytoplasm into the nucleus. Inside the nucleus, NFAT proteins interact with multiple transcriptional partners to assemble active transcription complexes. In TCR signaling, NFAT cooperates with AP-1 and induces transcription of IL-2 and other cytokine genes (7, 8).

In fact, the mechanism of NFAT activation was discovered indirectly by mechanism studies of the calcineurin inhibitors, cyclosporine and tacrolimus (2, 3). Cyclosporine binds to a cellular protein called cyclophilin. The cyclosporine-cyclophilin complex binds to inhibit the enzymatic activity of the calcineurin, thereby disrupting the dephosphorylation and nuclear translocation of NFAT. Tacrolimus (FK506) is a widely used calcineurin inhibitor, which binds to its binding protein (FKBP), and then the tacrolimus-FKBP complex binds calcineurin and inhibits its activities, including the NFAT-mediated transcription of IL-2 in T cells, thereby inhibiting T cell proliferation.

The NFAT family consists of five members: NFAT1, NFAT2, NFAT3, NFAT4, and NFAT5 (8). Four of these (NFAT1~4) are regulated by intracellular Ca^{2+} signaling and calcineurin. On the other hand, NFAT5 is a tonicity-responsive protein, and is activated in response to osmotic stress (9).

III. Calcineurin/NFAT Signaling Pathway in Dendritic Cells and Innate Immune Cells

1. NFAT-mediated IL-2 production in dendritic cells and innate immune cells

As described above, calcineurin/NFAT signaling induces IL-2 production in T cells. Recently, this pathway has also been reported to induce IL-2 production in dendritic cells (DCs) and innate immune cells. Similar to the pathway downstream of the TCR, two signal transduction pathways

downstream of pattern recognition receptors (PRRs) have been characterized in terms of their capacity to induce NFAT-mediated IL-2 production in innate immune cells. These are the pathways initiated by dectin-1 and TLR4/CD14 in DCs and macrophages.

Dectin-1 is a carbohydrate receptor which belongs to the C-type lectin family, and recognizes β -glucan. Since β -glucan is a fungal cell wall component, dectin-1 mediates the anti-fungal immune response. Goodridge *et al.* (14) had reported the first evidence for NFAT activation, with dectin-1 stimulation, which is in addition to the previously known function of activating NF- κ B. Upon β -glucan binding, Src-family kinase phosphorylates immunoreceptor tyrosin-based activation motif (ITAM)-like motif at the intracellular tail of dectin-1, thereby creating a docking site for Syk. Recruited tyrosine kinase Syk phosphorylates and activates PLC- γ . Activated PLC- γ then hydrolyzes PIP2 into IP3 and DAG. IP3 induces the endoplasmic reticulum Ca^{2+} release and subsequent calcineurin/NFAT activation, which results in IL-2 production in DCs and macrophages (Fig. 2).

Besides dectin-1, Toll-like receptor 4 (TLR4) and its co-receptor CD14 had been reported to induce NFAT-mediated IL-2 production independently to the canonical MyD88/TRIF pathway (15, 16). Lipopolysaccharide (LPS) engagement of TLR4/CD14 activates Src-family kinase and PLC- γ . Activated PLC- γ then hydrolyzes PIP2 into IP3 and DAG. IP3 induces influx of extracellular Ca^{2+} and subsequent calcineurin/NFAT activation, which results in IL-2 production in DCs (Fig. 2).

Although adjacent T cells of adaptive immunity can utilize the DC-derived IL-2 as a growth factor, interferon- γ -producing activity of natural killer (NK) cells and NKT cells can also be enhanced by DC-derived IL-2 (17~19). Therefore, when calcineurin inhibitors are administered, inhibition of NFAT-mediated IL-2 production not only affects T-cell mediated adaptive immunity, but also downregulates innate immune responses.

2. NFAT-mediated production of inflammatory mediators and prolongation of innate immune cells' survival

There have been reports revealing the role of NFAT in

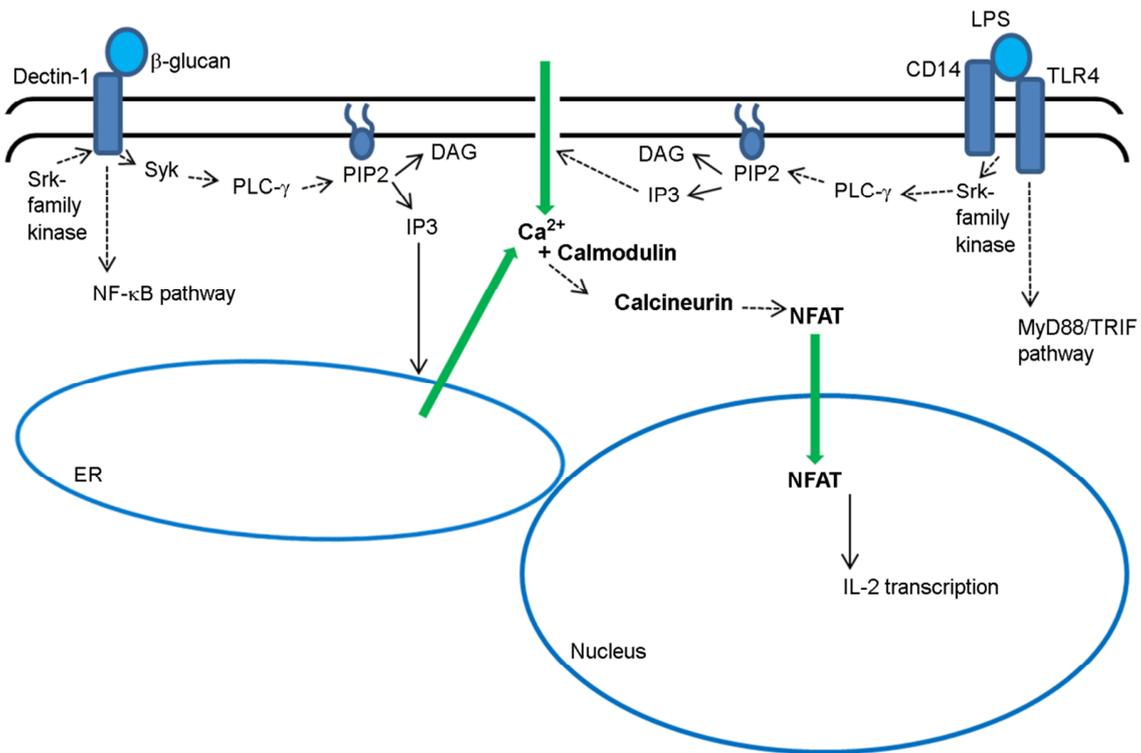


Figure 2. NFAT-mediated IL-2 production in dendritic cells and macrophages

regulating several key modulators of innate immunity, and calcineurin inhibitors may suppress innate immune responses with IL-2-independent mechanisms.

Tumor necrosis factor- α (TNF- α) is a prototypic pro-inflammatory cytokine, and NFAT1 and NFAT2 are critically involved in the expression of TNF- α in mast cells and NK cells (20, 21). IL-3 and granulocyte-macrophage colony-stimulating factor (GM-CSF) are required for differentiations of myeloid lineage innate immune cells. Furthermore, GM-CSF regulates the release of neutrophils from the bone marrow (22). It had been revealed that NFAT contribute to the transcriptions of IL-3 and GM-CSF (21, 23, 24). In addition, it has been discovered that NFAT activation regulates the induction of cyclooxygenase-2 (COX-2) (14), which is required for the production of key inflammatory mediators including prostaglandins. Besides, NFAT transcriptionally controls *Ptges1* that codes a protein called microsomal prostaglandin E synthase 1 (mPGES-1), a key

enzyme in PGE2 biosynthesis (25). Indeed, it had been proven that cyclosporine abolished COX-2 upregulation and PGE2 release in human neutrophils (26). Taken as a whole, inflammatory response can be profoundly decreased by the calcineurin inhibitor, and thus has considerable implications for the clinic (27).

Recently, the mechanism of mast cells' prolonged survival had been proven to be linked to the NFAT-dependent transcriptional induction of anti-apoptotic Bcl-2 family protein A1 (28). Previously reported A1-mediated prolonged survival of mast cells on IgE crosslinking (29) was abrogated by inhibition of calcineurin/NFAT by cyclosporine (28). In addition, anti-apoptotic functions of A1 had previously been reported in macrophages and neutrophils (30~32). Therefore, additional immune suppressive function of calcineurin inhibitors by inhibiting prolonged survival of innate immune cells is anticipated in organ transplant recipients.

IV. Positive Implications in Clinical Islet Transplantation

1. NFAT-mediated cytokines targeted in clinical islet transplantation (IL-2 and TNF- α)

In 2000, Shapiro *et al.* (33) reported successful clinical islet allotransplantation with the Edmonton protocol, and it became a standard immune suppression regimen. Tacrolimus was included as the calcineurin inhibitor, and daclizumab was used as an IL-2 receptor antagonist. In addition to the well-known suppression of T cell-derived IL-2, suppression of DC-mediated IL-2 by tacrolimus might have contributed to the potent immune suppression. Recently, several groups are conducting clinical trials of islet transplantation to improve the immune suppression regimen (34~36). They additionally used etanercept, which is a recombinant fusion protein of TNF receptor and IgG1 Fc domain (37), and functions as a decoy receptor that binds to and sequesters TNF- α (38). In addition, most of successful pre-clinical islet xenotransplantations targeted TNF- α with etanercept or adalimumab, an anti-TNF- α monoclonal antibody (39~45). Therefore, aforementioned NFAT-mediated expression of TNF- α and its inhibition by calcineurin inhibitors have

additional positive implication to the importance of TNF- α blockade in clinical islet transplantation.

2. Alleviation of Instant Blood-Mediated Inflammatory Reaction (IBMIR)

Clinical islet transplantation is performed to the liver through the portal vein. In this situation, the infused islets have direct contact with the blood stream, and results in Instant Blood-Mediated Inflammatory Reaction (IBMIR) which causes a considerable amount of early islet loss. The IBMIR is a multifaceted phenomenon comprising activation of the coagulation pathway, complement system, and platelets quickly followed by the recruitment and infiltration of neutrophils and monocytes (46~48) (Fig. 3). Since calcineurin inhibitors suppress NFAT-mediated development and release of inflammatory cells, they can reduce the recruitment of neutrophils and monocytes and alleviate the IBMIR. In addition, suppression of NFAT-mediated TNF- α can further decrease the recruitment and activation of neutrophils and monocytes. In other arms of IBMIR, activated platelets release a soluble form of CD40 ligand (sCD40L), and activate CD40-expressing neutrophils (49). Crist *et al.* had reported that NFAT1 is a key transcriptional regulator of CD40L expression in megakaryocytes, the precursors of

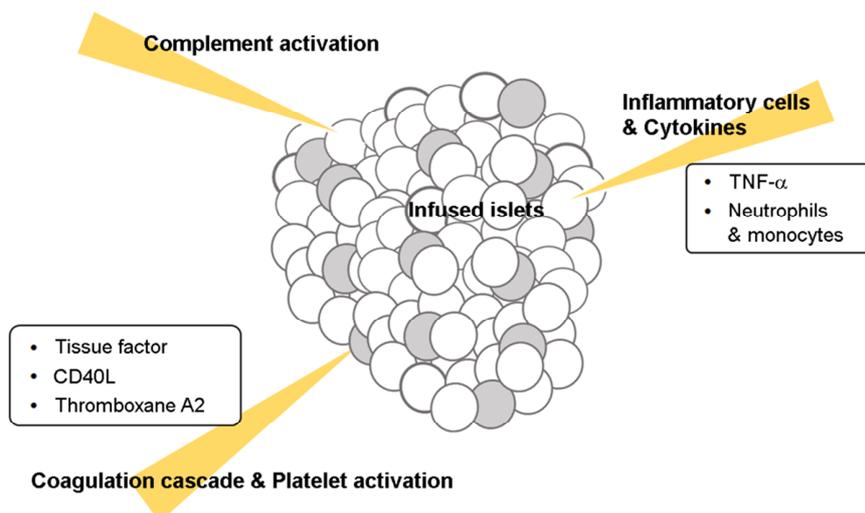


Figure 3. Instant Blood-Mediated Inflammatory Reaction (IBMIR) and NFAT-mediated targets

blood platelets, and that biochemical inhibition of NFAT activity in megakaryocytes diminishes platelet CD40L (50, 51). Additionally, since calcineurin inhibitors suppress NFAT-mediated expression of COX-2, thromboxane A2 production and subsequent platelet aggregation can be diminished. Furthermore, calcineurin inhibitors including cyclosporine inhibit NFAT-mediated expression of tissue factor (52), which interacts with coagulation factor VII and initiates the coagulation pathway (53). To summarize, calcineurin inhibitors may contribute to alleviation of IBMIR by suppressing activations of coagulation pathway, platelets, neutrophils and monocytes, thereby minimizing early islet loss in clinical islet transplantation.

V. Negative implications of Innate Immune Regulation by calcineurin inhibitors

1. Increased susceptibility to opportunistic infections

Since calcineurin inhibitors suppress not only T cells but also dendritic cells and innate immune cells, potent suppression of immune rejection in transplantation could be anticipated. However, additional suppression of innate immune cells may elevate susceptibility to opportunistic infections in the recipients. Indeed, correlation between intense NFAT suppression and recurrent infections in cyclosporine-treated patients has been reported (54). Recent discoveries of calcineurin/NFAT signaling pathway activation downstream of pattern recognition receptors (PRRs) may explain the greater susceptibility of opportunistic infections (27, 55). The fact that conditional deletion of calcineurin in neutrophils decreased resistance to infection with *Candida albicans* in mice suggests the importance of NFAT-mediated innate immune responses (56). In addition to aforementioned dectin-1 and CD14/TLR4, expression of another PRR nucleotide-binding oligomerization domain 1 (Nod1) and neutrophil phagocytic killing activity was significantly reduced in cyclosporine-treated mice (57). These interactions between calcineurin/NFAT signaling pathway and PRRs may explain the increased susceptibility to opportunistic fungal or bacterial infections in transplant recipients treated with calcineurin inhibitors.

2. Homeostasis dysregulation with interruption of innate immunity

It is well-known that NFAT is involved in the development of T cell and B cells (58~61). Though the role of NFAT in the regulation of hematopoiesis of innate immune cells is largely unknown, members of the NFAT family are expressed in CD34⁺ hematopoietic stem cells and their differentially regulated expression during the lineage-specific differentiation of myeloid cells have been reported (62, 63). Recently, Fric *et al.* revealed that NFAT is a potent negative regulator of myeloid cell development (64). Therefore, calcineurin inhibitors may disrupt the hematopoiesis and homeostasis of the innate immune cells.

Since soluble mediators of innate immunity do not discriminate between the host and the graft, immune responses against transplanted grafts may provoke host tissue damage. Anti-inflammatory cytokine IL-10 has a role in regenerative healing, and its expression can be upregulated through calcineurin/NFAT pathway (14, 56, 65). Therefore, calcineurin inhibitors may interrupt homeostatic healing of host tissue damage in the graft recipients.

3. Unfavorable effects of calcineurin inhibitors on regulatory T cells

Recently, Shin *et al.* reported long term survival (167 ~ >603 days) of pig islet xenografts in non-human primates with infusion of autologous regulatory T cells in conjunction with other immune suppressants (44). Regulatory T cells can suppress diverse immune responses, and have physiological functions in self-tolerance. Notably, the induction of regulatory T cells may induce graft-specific tolerance in transplant recipients (66).

Regulatory T cells have higher dependence on IL-2 than effector T cells for their maintenance (67~69). As mentioned earlier, calcineurin/NFAT signaling is required for the expression of not only T cell-derived IL-2 but also DC-derived IL-2. Therefore, calcineurin inhibitors may not be beneficial in terms of regulatory T cell maintenance. Foxp3 is a lineage-defining transcription factor of CD4⁺ regulatory T cells, and has crucial roles for the suppressive function of

these cells. NFAT interact with Foxp3 as a transcriptional partner (70~73), and NFAT-Foxp3 transcriptional complex induces the expression of IL-2 receptor α chain (CD25) and cytotoxic T lymphocyte antigen 4 (CTLA-4) (71), which are involved in the suppressive function. In addition, it has been reported that NFAT was essential for the peripheral conversion of CD4⁺Foxp3⁻ T cells to CD4⁺Foxp3⁺ regulatory T cells (74), and the Foxp3 induction was completely blocked by cyclosporine (75).

4. Unfavorable effects of calcineurin inhibitors on neovascularization

In contrast to transplantation of vascularized solid organs, transplantation of islets requires neovascularization to the islet cells for engraftment. It has recently been shown that NFAT regulates the expression of hypoxia-inducible factor 1 α (HIF-1 α) in mast cells (76). HIF-1 α is critical for adaptation to oxygen deficit and it regulates angiogenesis (77). PGE2 and vascular endothelial growth factor (VEGF) stimulate endothelial cell proliferation, migration and, eventually, neovascularization. However, this angiogenesis had been shown to be inhibited by cyclosporine (78). As described earlier, NFAT regulates PGE2 biosynthesis. In addition, engagement of VEGF receptors on endothelial cell by VEGF induces the expression of additional VEGF and VEGF receptors through the calcineurin/NFAT signaling pathway (79). In other words, calcineurin inhibitors may have unfavorable effects on the neovascularization and engraftment of the transplanted islets.

VI. Effects on Beta-cell Function

Islet transplantation aims to supply insulin-producing β -cells and normalize blood glucose levels in Type 1 diabetes patients. To achieve this goal, transplanted β -cells have to maintain viability, and produce and secrete insulin. In addition, insulin has to be utilized by the target cells, including muscle cells. Although the molecular mechanisms are not completely understood, calcineurin is involved in regulation of replication and survival of β -cells (80), and production and secretion of insulin (81, 82). Skeletal muscle

is the primary site for glucose uptake in response to insulin (83), and is composed of a mixture of three myofiber types which have variable insulin sensitivity. NFAT has been reported to be responsible for the transcriptional activation and repression of distinct myosin fibers, thereby increasing insulin-sensitive myofibers and decreasing insulin-resistant myofibers in the skeletal muscle (84~86).

New-onset diabetes mellitus after transplantation (NODAT) occurs in 15~30% of recipients after renal transplantation with immunosuppressive drugs (87~89), and use of calcineurin inhibitor is one of its risk factors (90). Since calcineurin/NFAT is involved in above-mentioned functions of β -cells, calcineurin inhibitors may induce β -cell death (91), diminished insulin production and secretion (81, 92), and impaired insulin sensitivity (93, 94). Considering the reversibility after withdrawal of the drug (95), impaired insulin secretion and insulin resistance seems to be the major mechanisms.

VII. CONCLUSION

Activation of adaptive immunity with T cell-derived IL-2 production had been thought as the principal role of calcineurin/NFAT. However, it has become evident that calcineurin/NFAT has multiple roles in the regulation of dendritic cells and innate immune cells. Although calcineurin inhibitors are widely used in clinical transplantations, our attention on calcineurin inhibitors has not been extended to the recently discovered roles in innate immune system. In islet xenotransplantation, intense immune suppression covering innate immunity and possible alleviation of IBMIR may be beneficial. However, long-term use of calcineurin inhibitors may not be favorable due to the possible effects on opportunistic infections, disruption of homeostasis, regulatory T cells, neovascularization, and β -cell functions.

The complicated functions of NFAT in various cell types are not fully understood. Various cell types do not homogeneously express the five NFAT isomers, thereby varying the effects of calcineurin inhibitors. In addition, the NFAT's transcriptional partners such as AP-1 and Foxp3 are differentially expressed in various cell types, and the balance of these cofactors present may result in the different

outcomes (27). Although it is complicated, thorough research and understanding of the roles of NFAT in diverse immune cells is required. Through the improvement in the understanding on the roles of calcineurin/NFAT, calcineurin inhibitors can be utilized more effectively and safely in the transplant recipients.

REFERENCES

- 1) Shaw JP, Utz PJ, Durand DB, Toole JJ, Emmel EA, Crabtree GR. Identification of a putative regulator of early T cell activation genes. *Science* 1988;241:202-5.
- 2) Jain J, McCaffrey PG, Miner Z, Kerppola TK, Lambert JN, Verdine GL, *et al.* The T-cell transcription factor NFATp is a substrate for calcineurin and interacts with Fos and Jun. *Nature* 1993;365:352-5.
- 3) Clipstone NA, Crabtree GR. Identification of calcineurin as a key signalling enzyme in T-lymphocyte activation. *Nature* 1992;357:695-7.
- 4) Rao A, Luo C, Hogan PG. Transcription factors of the NFAT family: regulation and function. *Annu Rev Immunol* 1997;15:707-47.
- 5) Crabtree GR, Olson EN. NFAT signaling: choreographing the social lives of cells. *Cell* 2002;109 Suppl: S67-79.
- 6) Graef IA, Chen F, Crabtree GR. NFAT signaling in vertebrate development. *Curr Opin Genet Dev* 2001;11: 505-12.
- 7) Hogan PG, Chen L, Nardone J, Rao A. Transcriptional regulation by calcium, calcineurin, and NFAT. *Genes Dev* 2003;17:2205-32.
- 8) Macian F. NFAT proteins: key regulators of T-cell development and function. *Nat Rev Immunol* 2005;5: 472-84.
- 9) Muller MR, Rao A. NFAT, immunity and cancer: a transcription factor comes of age. *Nat Rev Immunol* 2010;10:645-56.
- 10) Park CG, Bottino R, Hawthorne WJ. Current status of islet xenotransplantation. *Int J Surg* 2015;23:261-6.
- 11) van der Windt DJ, Bottino R, Kumar G, Wijkstrom M, Hara H, Ezzelarab M, *et al.* Clinical islet xenotransplantation: how close are we? *Diabetes* 2012;61:3046-55.
- 12) Ekser B, Ezzelarab M, Hara H, van der Windt DJ, Wijkstrom M, Bottino R, *et al.* Clinical xenotransplantation: the next medical revolution? *Lancet* 2012;379: 672-83.
- 13) Vadori M, Cozzi E. The immunological barriers to xenotransplantation. *Tissue Antigens* 2015;86:239-53.
- 14) Goodridge HS, Simmons RM, Underhill DM. Dectin-1 stimulation by *Candida albicans* yeast or zymosan triggers NFAT activation in macrophages and dendritic cells. *J Immunol* 2007;178:3107-15.
- 15) Zandoni I, Ostuni R, Capuano G, Collini M, Caccia M, Ronchi AE, *et al.* CD14 regulates the dendritic cell life cycle after LPS exposure through NFAT activation. *Nature* 2009;460:264-8.
- 16) Zandoni I, Bodio C, Broggi A, Ostuni R, Caccia M, Collini M, *et al.* Similarities and differences of innate immune responses elicited by smooth and rough LPS. *Immunol Lett* 2012;142:41-7.
- 17) Granucci F, Zandoni I, Pavelka N, Van Dommelen SL, Andoniou CE, Belardelli F, *et al.* A contribution of mouse dendritic cell-derived IL-2 for NK cell activation. *J Exp Med* 2004;200:287-95.
- 18) Granucci F, Zandoni I, Ricciardi-Castagnoli P. Natural killer (NK) cell functions can be strongly boosted by activated dendritic cells (DC). *Eur J Immunol* 2006;36: 2819-20.
- 19) Fujii S, Shimizu K, Kronenberg M, Steinman RM. Prolonged IFN-gamma-producing NKT response induced with alpha-galactosylceramide-loaded DCs. *Nat Immunol* 2002;3:867-74.
- 20) Klein M, Klein-Hessling S, Palmetshofer A, Serfling E, Tertilt C, Bopp T, *et al.* Specific and redundant roles for NFAT transcription factors in the expression of mast cell-derived cytokines. *J Immunol* 2006;177:6667-74.
- 21) Aramburu J, Azzoni L, Rao A, Perussia B. Activation and expression of the nuclear factors of activated T cells, NFATp and NFATc, in human natural killer cells: regulation upon CD16 ligand binding. *J Exp Med* 1995; 182:801-10.
- 22) Christopher MJ, Link DC. Regulation of neutrophil homeostasis. *Curr Opin Hematol* 2007;14:3-8.
- 23) Hawwari A, Burrows J, Vadas MA, Cockerill PN. The human IL-3 locus is regulated cooperatively by two NFAT-dependent enhancers that have distinct tissue-

- specific activities. *J Immunol* 2002;169:1876-86.
- 24) Brettingham-Moore KH, Rao S, Juelich T, Shannon MF, Holloway AF. GM-CSF promoter chromatin remodelling and gene transcription display distinct signal and transcription factor requirements. *Nucleic Acids Res* 2005; 33:225-34.
- 25) Zanoni I, Ostuni R, Barresi S, Di Gioia M, Broggi A, Costa B, *et al.* CD14 and NFAT mediate lipopolysaccharide-induced skin edema formation in mice. *J Clin Invest* 2012;122:1747-57.
- 26) Vega A, Chacon P, Monteseirin J, El Bekay R, Alba G, Martin-Nieto J, *et al.* Expression of the transcription factor NFAT2 in human neutrophils: IgE-dependent, Ca²⁺- and calcineurin-mediated NFAT2 activation. *J Cell Sci* 2007;120:2328-37.
- 27) Fric J, Zelante T, Wong AY, Mertes A, Yu HB, Ricciardi-Castagnoli P. NFAT control of innate immunity. *Blood* 2012;120:1380-9.
- 28) Ulleras E, Karlberg M, Moller Westerberg C, Alfredsson J, Gerondakis S, Strasser A, *et al.* NFAT but not NF- κ B is critical for transcriptional induction of the prosurvival gene A1 after IgE receptor activation in mast cells. *Blood* 2008;111:3081-9.
- 29) Xiang Z, Ahmed AA, Moller C, Nakayama K, Hatakeyama S, Nilsson G. Essential role of the prosurvival bcl-2 homologue A1 in mast cell survival after allergic activation. *J Exp Med* 2001;194:1561-9.
- 30) Orlofsky A, Somogyi RD, Weiss LM, Prystowsky MB. The murine antiapoptotic protein A1 is induced in inflammatory macrophages and constitutively expressed in neutrophils. *J Immunol* 1999;163:412-9.
- 31) Hamasaki A, Sendo F, Nakayama K, Ishida N, Negishi I, Nakayama K, *et al.* Accelerated neutrophil apoptosis in mice lacking A1-a, a subtype of the bcl-2-related A1 gene. *J Exp Med* 1998;188:1985-92.
- 32) Lin EY, Orlofsky A, Wang HG, Reed JC, Prystowsky MB. A1, a Bcl-2 family member, prolongs cell survival and permits myeloid differentiation. *Blood* 1996;87:983-92.
- 33) Shapiro AM, Lakey JR, Ryan EA, Korbitt GS, Toth E, Warnock GL, *et al.* Islet transplantation in seven patients with type 1 diabetes mellitus using a glucocorticoid-free immunosuppressive regimen. *N Engl J Med* 2000;343:230-8.
- 34) Bellin MD, Barton FB, Heitman A, Harmon JV, Kandaswamy R, Balamurugan AN, *et al.* Potent induction immunotherapy promotes long-term insulin independence after islet transplantation in type 1 diabetes. *Am J Transplant* 2012;12:1576-83.
- 35) Qi M, Kinzer K, Danielson KK, Martellotto J, Barbaro B, Wang Y, *et al.* Five-year follow-up of patients with type 1 diabetes transplanted with allogeneic islets: the UIC experience. *Acta Diabetol* 2014;51:833-43.
- 36) Rickels MR, Liu C, Shlansky-Goldberg RD, Soleimanpour SA, Vivek K, Kamoun M, *et al.* Improvement in beta-cell secretory capacity after human islet transplantation according to the CIT07 protocol. *Diabetes* 2013;62:2890-7.
- 37) Weinblatt ME, Kremer JM, Bankhurst AD, Bulpitt KJ, Fleischmann RM, Fox RI, *et al.* A trial of etanercept, a recombinant tumor necrosis factor receptor:Fc fusion protein, in patients with rheumatoid arthritis receiving methotrexate. *N Engl J Med* 1999;340:253-9.
- 38) Zalevsky J, Secher T, Ezhevsky SA, Janot L, Steed PM, O'Brien C, *et al.* Dominant-negative inhibitors of soluble TNF attenuate experimental arthritis without suppressing innate immunity to infection. *J Immunol* 2007; 179:1872-83.
- 39) Cardona K, Korbitt GS, Milas Z, Lyon J, Cano J, Jiang W, *et al.* Long-term survival of neonatal porcine islets in nonhuman primates by targeting costimulation pathways. *Nat Med* 2006;12:304-6.
- 40) Cardona K, Milas Z, Strobert E, Cano J, Jiang W, Safley SA, *et al.* Engraftment of adult porcine islet xenografts in diabetic nonhuman primates through targeting of costimulation pathways. *Am J Transplant* 2007;7:2260-8.
- 41) Thompson P, Badell IR, Lowe M, Cano J, Song M, Leopardi F, *et al.* Islet xenotransplantation using gal-deficient neonatal donors improves engraftment and function. *Am J Transplant* 2011;11:2593-602.
- 42) Thompson P, Badell IR, Lowe M, Turner A, Cano J, Avila J, *et al.* Alternative immunomodulatory strategies for xenotransplantation: CD40/154 pathway-sparing regimens promote xenograft survival. *Am J Transplant* 2012;12:1765-75.
- 43) Thompson P, Cardona K, Russell M, Badell IR, Shaffer V, Korbitt G, *et al.* CD40-specific costimulation blockade enhances neonatal porcine islet survival in non-

- human primates. *Am J Transplant* 2011;11:947-57.
- 44) Shin JS, Kim JM, Kim JS, Min BH, Kim YH, Kim HJ, *et al.* Long-Term Control of Diabetes in Immunosuppressed Nonhuman Primates (NHP) by the Transplantation of Adult Porcine Islets. *Am J Transplant* 2015;15:2837-50.
- 45) Jung KC, Park CG, Jeon YK, Park HJ, Ban YL, Min HS, *et al.* In situ induction of dendritic cell-based T cell tolerance in humanized mice and nonhuman primates. *J Exp Med* 2011;208:2477-88.
- 46) Bennet W, Groth CG, Larsson R, Nilsson B, Korsgren O. Isolated human islets trigger an instant blood mediated inflammatory reaction: implications for intraportal islet transplantation as a treatment for patients with type 1 diabetes. *Ups J Med Sci* 2000;105:125-33.
- 47) Nilsson B, Ekdahl KN, Korsgren O. Control of instant blood-mediated inflammatory reaction to improve islets of Langerhans engraftment. *Curr Opin Organ Transplant* 2011;16:620-6.
- 48) Moberg L, Johansson H, Lukinius A, Berne C, Foss A, Källén R, *et al.* Production of tissue factor by pancreatic islet cells as a trigger of detrimental thrombotic reactions in clinical islet transplantation. *The Lancet* 2002;360:2039-45.
- 49) Vanichakarn P, Blair P, Wu C, Freedman JE, Chakrabarti S. Neutrophil CD40 enhances platelet-mediated inflammation. *Thromb Res* 2008;122:346-58.
- 50) Crist SA, Sprague DL, Ratliff TL. Nuclear factor of activated T cells (NFAT) mediates CD154 expression in megakaryocytes. *Blood* 2008;111:3553-61.
- 51) Crist SA, Elzey BD, Ahmann MT, Ratliff TL. Early growth response-1 (EGR-1) and nuclear factor of activated T cells (NFAT) cooperate to mediate CD40L expression in megakaryocytes and platelets. *J Biol Chem* 2013;288:33985-96.
- 52) Armesilla AL, Lorenzo E, Gomez del Arco P, Martinez-Martinez S, Alfranca A, Redondo JM. Vascular endothelial growth factor activates nuclear factor of activated T cells in human endothelial cells: a role for tissue factor gene expression. *Mol Cell Biol* 1999;19:2032-43.
- 53) Johansson H, Lukinius A, Moberg L, Lundgren T, Berne C, Foss A, *et al.* Tissue factor produced by the endocrine cells of the islets of Langerhans is associated with a negative outcome of clinical islet transplantation. *Diabetes* 2005;54:1755-62.
- 54) Billing H, Breil T, Schmidt J, Tonshoff B, Schmitt CP, Giese T, *et al.* Pharmacodynamic monitoring by residual NFAT-regulated gene expression in stable pediatric liver transplant recipients. *Pediatr Transplant* 2012;16:187-94.
- 55) Vandewalle A, Tourneur E, Bens M, Chassin C, Werts C. Calcineurin/NFAT signaling and innate host defence: a role for NOD1-mediated phagocytic functions. *Cell Commun Signal* 2014;12:8.
- 56) Greenblatt MB, Aliprantis A, Hu B, Glimcher LH. Calcineurin regulates innate antifungal immunity in neutrophils. *J Exp Med* 2010;207:923-31.
- 57) Tourneur E, Ben Mkaddem S, Chassin C, Bens M, Goujon JM, Charles N, *et al.* Cyclosporine A Impairs Nucleotide Binding Oligomerization Domain (Nod1)-Mediated Innate Antibacterial Renal Defenses in Mice and Human Transplant Recipients. *Plos Pathogens* 2013;9.
- 58) Berland R, Wortis HH. Normal B-1a cell development requires B cell-intrinsic NFATc1 activity. *Proc Natl Acad Sci U S A* 2003;100:13459-64.
- 59) Neilson JR, Winslow MM, Hur EM, Crabtree GR. Calcineurin B1 is essential for positive but not negative selection during thymocyte development. *Immunity* 2004;20:255-66.
- 60) Winslow MM, Gallo EM, Neilson JR, Crabtree GR. The calcineurin phosphatase complex modulates immunogenic B cell responses. *Immunity* 2006;24:141-52.
- 61) Muller MR, Sasaki Y, Stevanovic I, Lamperti ED, Ghosh S, Sharma S, *et al.* Requirement for balanced Ca/NFAT signaling in hematopoietic and embryonic development. *Proc Natl Acad Sci U S A* 2009;106:7034-9.
- 62) Kiani A, Habermann I, Haase M, Feldmann S, Boxberger S, Sanchez-Fernandez MA, *et al.* Expression and regulation of NFAT (nuclear factors of activated T cells) in human CD34+ cells: down-regulation upon myeloid differentiation. *J Leukoc Biol* 2004;76:1057-65.
- 63) Kiani A, Kuithan H, Kuithan F, Kyttala S, Habermann I, Temme A, *et al.* Expression analysis of nuclear factor of activated T cells (NFAT) during myeloid differentiation of CD34+ cells: regulation of Fas ligand gene expression in megakaryocytes. *Exp Hematol* 2007;35:757-70.
- 64) Fric J, Lim CX, Koh EG, Hofmann B, Chen J, Tay HS,

- et al.* Calcineurin/NFAT signalling inhibits myeloid haematopoiesis. *EMBO Mol Med* 2012;4:269-82.
- 65) Mourao-Sa D, Robinson MJ, Zelenay S, Sancho D, Chakravarty P, Larsen R, *et al.* CLEC-2 signaling via Syk in myeloid cells can regulate inflammatory responses. *Eur J Immunol* 2011;41:3040-53.
- 66) Walsh PT, Taylor DK, Turka LA. Tregs and transplantation tolerance. *J Clin Invest* 2004;114:1398-403.
- 67) Malek TR, Bayer AL. Tolerance, not immunity, crucially depends on IL-2. *Nat Rev Immunol* 2004;4:665-74.
- 68) Setoguchi R, Hori S, Takahashi T, Sakaguchi S. Homeostatic maintenance of natural Foxp3(+) CD25(+) CD4(+) regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. *J Exp Med* 2005;201:723-35.
- 69) Guiducci C, Valzasina B, Dislich H, Colombo MP. CD40/CD40L interaction regulates CD4+CD25+ T reg homeostasis through dendritic cell-produced IL-2. *Eur J Immunol* 2005;35:557-67.
- 70) Bopp T, Palmethofer A, Serfling E, Heib V, Schmitt S, Richter C, *et al.* NFATc2 and NFATc3 transcription factors play a crucial role in suppression of CD4+ T lymphocytes by CD4+ CD25+ regulatory T cells. *J Exp Med* 2005;201:181-7.
- 71) Hu H, Djuretic I, Sundrud MS, Rao A. Transcriptional partners in regulatory T cells: Foxp3, Runx and NFAT. *Trends Immunol* 2007;28:329-32.
- 72) Sumpter TL, Payne KK, Wilkes DS. Regulation of the NFAT pathway discriminates CD4+CD25+ regulatory T cells from CD4+CD25- helper T cells. *J Leukoc Biol* 2008;83:708-17.
- 73) Wu Y, Borde M, Heissmeyer V, Feuerer M, Lapan AD, Stroud JC, *et al.* FOXP3 controls regulatory T cell function through cooperation with NFAT. *Cell* 2006;126:375-87.
- 74) Zheng Y, Josefowicz S, Chaudhry A, Peng XP, Forbush K, Rudensky AY. Role of conserved non-coding DNA elements in the Foxp3 gene in regulatory T-cell fate. *Nature* 2010;463:808-12.
- 75) Tone Y, Furuuchi K, Kojima Y, Tykocinski ML, Greene MI, Tone M. Smad3 and NFAT cooperate to induce Foxp3 expression through its enhancer. *Nat Immunol* 2008;9:194-202.
- 76) Walczak-Drzewiecka A, Ratajewski M, Wagner W, Dastych J. HIF-1alpha is up-regulated in activated mast cells by a process that involves calcineurin and NFAT. *J Immunol* 2008;181:1665-72.
- 77) Semenza GL. HIF-1: mediator of physiological and pathophysiological responses to hypoxia. *J Appl Physiol* (1985) 2000;88:1474-80.
- 78) Hernandez GL, Volpert OV, Iniguez MA, Lorenzo E, Martinez-Martinez S, Grau R, *et al.* Selective inhibition of vascular endothelial growth factor-mediated angiogenesis by cyclosporin A: roles of the nuclear factor of activated T cells and cyclooxygenase 2. *J Exp Med* 2001;193:607-20.
- 79) Jinnin M, Medici D, Park L, Limaye N, Liu Y, Boscolo E, *et al.* Suppressed NFAT-dependent VEGFR1 expression and constitutive VEGFR2 signaling in infantile hemangioma. *Nat Med* 2008;14:1236-46.
- 80) Soleimanpour SA, Crutchlow MF, Ferrari AM, Raum JC, Groff DN, Rankin MM, *et al.* Calcineurin signaling regulates human islet {beta}-cell survival. *J Biol Chem* 2010;285:40050-9.
- 81) Ozbay LA, Smidt K, Mortensen DM, Carstens J, Jorgensen KA, Rungby J. Cyclosporin and tacrolimus impair insulin secretion and transcriptional regulation in INS-1E beta-cells. *Br J Pharmacol* 2011;162:136-46.
- 82) Lawrence MC, Bhatt HS, Easom RA. NFAT regulates insulin gene promoter activity in response to synergistic pathways induced by glucose and glucagon-like peptide-1. *Diabetes* 2002;51:691-8.
- 83) DeFronzo RA. Lilly lecture 1987. The triumvirate: beta-cell, muscle, liver. A collusion responsible for NIDDM. *Diabetes* 1988;37:667-87.
- 84) Calabria E, Ciciliot S, Moretti I, Garcia M, Picard A, Dyar KA, *et al.* NFAT isoforms control activity-dependent muscle fiber type specification. *Proc Natl Acad Sci U S A* 2009;106:13335-40.
- 85) Rana ZA, Gundersen K, Buonanno A. Activity-dependent repression of muscle genes by NFAT. *Proc Natl Acad Sci U S A* 2008;105:5921-6.
- 86) McCullagh KJ, Calabria E, Pallafacchina G, Ciciliot S, Serrano AL, Argentini C, *et al.* NFAT is a nerve activity sensor in skeletal muscle and controls activity-dependent myosin switching. *Proc Natl Acad Sci U S A* 2004;101:10590-5.
- 87) Chakkerla HA, Mandarino LJ. Calcineurin inhibition

- and new-onset diabetes mellitus after transplantation. *Transplantation* 2013;95:647-52.
- 88) Cosio FG, Kudva Y, van der Velde M, Larson TS, Textor SC, Griffin MD, *et al.* New onset hyperglycemia and diabetes are associated with increased cardiovascular risk after kidney transplantation. *Kidney Int* 2005;67: 2415-21.
- 89) Kasiske BL, Snyder JJ, Gilbertson D, Matas AJ. Diabetes mellitus after kidney transplantation in the United States. *Am J Transplant* 2003;3:178-85.
- 90) Heisel O, Heisel R, Balshaw R, Keown P. New onset diabetes mellitus in patients receiving calcineurin inhibitors: a systematic review and meta-analysis. *Am J Transplant* 2004;4:583-95.
- 91) Drachenberg CB, Klassen DK, Weir MR, Wiland A, Fink JC, Bartlett ST, *et al.* Islet cell damage associated with tacrolimus and cyclosporine: morphological features in pancreas allograft biopsies and clinical correlation. *Transplantation* 1999;68:396-402.
- 92) Oetjen E, Baun D, Beimesche S, Krause D, Cierny I, Blume R, *et al.* Inhibition of human insulin gene transcription by the immunosuppressive drugs cyclosporin A and tacrolimus in primary, mature islets of transgenic mice. *Mol Pharmacol* 2003;63:1289-95.
- 93) Hjelmessaeth J, Hagen LT, Asberg A, Midtvedt K, Storset O, Halvorsen CE, *et al.* The impact of short-term ciclosporin A treatment on insulin secretion and insulin sensitivity in man. *Nephrol Dial Transplant* 2007;22: 1743-9.
- 94) Kutkuhn B, Hollenbeck M, Heering P, Koch M, Voiculescu A, Reinhard T, *et al.* Development of insulin resistance and elevated blood pressure during therapy with cyclosporine A. *Blood Press* 1997;6:13-7.
- 95) Wahlstrom HE, Akimoto R, Endres D, Kolterman O, Moossa AR. Recovery and hypersecretion of insulin and reversal of insulin resistance after withdrawal of short-term cyclosporine treatment. *Transplantation* 1992; 53:1190-5.
-