

## Review Article



**Received:** Nov 12, 2017

**Revised:** Feb 5, 2018

**Accepted:** Feb 13, 2018

### \*Correspondence to

**Dae Woo Kim**

Department of Otorhinolaryngology-Head and Neck Surgery, Seoul Metropolitan Government-Seoul National University Boramae Medical Center, 20 Boramae-ro 5-gil, Dongjak-gu, Seoul 07061, Korea.  
E-mail: kicubi@daum.net

Copyright © 2018. The Korean Association of Immunologists

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Conflict of Interest

The authors declare no potential conflicts of interest.

### Abbreviations

CRS, chronic rhinosinusitis; CRSsNP, chronic rhinosinusitis without nasal polyps; CRSwNP, chronic rhinosinusitis with nasal polyps; CRTH2, chemoattractant receptor homologous molecule expressed on Th2 cells; CysLT, cysteinyl leukotriene; ILC, innate lymphoid cell; NP, nasal polyp; OSM, oncostatin M; PGD2, prostaglandin D2; PGP, Pro-Gly-Pro; STAT, signal transducer and activator of transcription; Th, T helper; TSLP, thymic stromal lymphopoietin

# Pathogenesis of Recalcitrant Chronic Rhinosinusitis: The Emerging Role of Innate Immune Cells

Il Gyu Kong<sup>1</sup>, Dae Woo Kim<sup>2,\*</sup>

<sup>1</sup>Department of Otorhinolaryngology-Head and Neck Surgery, Hallym University Sacred Heart Hospital, Hallym University College of Medicine, Anyang 14068, Korea

<sup>2</sup>Department of Otorhinolaryngology-Head and Neck Surgery, Seoul Metropolitan Government-Seoul National University Boramae Medical Center, Seoul 07061, Korea

## ABSTRACT

Chronic rhinosinusitis (CRS) is a major part of the recalcitrant inflammatory diseases of the upper airway that needs enormous socioeconomic burden. T helper (Th) 2 type immune responses recruiting eosinophils were the most well-known immune players in CRS pathogenesis especially in western countries. By the piling up of a vast amount of researches to elucidate the pathogenic mechanism of CRS recently, heterogeneous inflammatory processes were found to be related to the phenotypes of CRS. Recently more cells other than T cells were in the focus of CRS pathogenesis, such as the epithelial cell, macrophage, innate lymphoid cells, and neutrophils. Here, we reviewed the recent research focusing on the innate immune cells related to CRS pathogenesis.

**Keywords:** Chronic rhinosinusitis; Nasal polyps; Recalcitrant chronic rhinosinusitis; Pathogenesis; Innate immune cell

## INTRODUCTION

Chronic rhinosinusitis (CRS) is chronic inflammation of the sinonasal mucosa and characterized by nasal congestion, rhinorrhea, and a decreased sense of smell persisting for greater than 12 weeks (1). CRS can be subdivided into two major categories based on whether nasal polyps (NPs) are present (chronic rhinosinusitis with nasal polyps [CRSwNP]) or absent (chronic rhinosinusitis without nasal polyps [CRSsNP]). CRS is a common inflammatory disease in paranasal sinus mucosa, affecting more about 10% of the Asian population (2-4). At least 10% of patients with CRS do not respond to medical and surgical treatment regimens and develop a refractory disease. Patients with CRSwNP often have higher disease severity and a risk of recurrence than do patients with CRSsNP (5) and various aspects of cellular responses were investigated to elucidate the immunological phenotypes of CRSwNP.

Phenotypes of CRSwNP, clinically observable characteristics with the presence of NP in the nasal cavity, have been classified merely based on histologic cellular infiltration in NP into eosinophilic or non-eosinophilic NP (6). The degrees of mucosal eosinophil infiltration have repeatedly been reported as a potent predictor of the poor outcome of surgeries and a risk of having comorbidities (7). However, there are no standards to evaluate tissue eosinophilia due to uneven distribution throughout the tissue, and all the reports are using their own methods

### Author Contributions

Conceptualization: Kong IG, Kim DW; Funding acquisition: Kong IG, Kim DW; Writing - original draft: Kong IG; Writing - review & editing: Kim DW.

to classify NP into eosinophilic or non-eosinophilic (8). There are quite significant racial and regional differences in the eosinophilic polyp proportion. Moreover, besides the eosinophils, various kinds and degree of inflammatory cells infiltrate in most of the NP, but the function and pathological roles of these cells are not yet completely elucidated (9). In even eosinophilic NP, neutrophils and macrophages exist and may have a role in nasal polypogenesis (3). Therefore, the current subtyping of CRS or NP based on inflammatory cells are too simple to explain the pathogenesis of CRSwNP. Due to these challenges, it is difficult to reach favorable treatment results and needs more research on other immune cells infiltrated in NP. CRS involves multiple pathogenesis mechanisms showing high heterogeneity, and those are suggested to cause different therapeutic responses. Characterization of endotypes of NP also helps to determine primary optimal therapeutic modality, select a good responder to a specific treatment and predict treatment outcomes and risks for comorbidities.

Though aberrant mucosal immune response in CRSwNP is observed and reported in the literature at the various aspects of view, it is unclear whether these phenomena are the cause or result of eosinophilic or neutrophilic inflammation. Furthermore, the primary mechanism of the recalcitrant or treatment-nonresponsive CRSwNP is not known. Moreover, in contrast to NP from Caucasian patients, NP from Asian patients are characterized by a much lesser portion of eosinophilic inflammation, suggesting other cells than eosinophils can play more important roles in CRS pathogenesis. Neutrophils and macrophages are more frequently observed in these population and expected to have potential roles in pathogenesis (10,11). In this article, the recent research results in possible roles of innate immune cells are reviewed, and clues for the key mechanisms of recalcitrant CRSwNP were summarized.

## EPITHELIAL CELLS

Epithelial-derived innate cytokines such as IL-25 and IL-33 may also participate in the evolution of NP (12). These cytokines are reported to exaggerate T helper (Th) 2 type immune responses in the typical Th2-prone diseases such as atopic dermatitis and bronchial asthma (13-17). IL-33 is secreted by immune cells such as macrophages and dendritic cells as well as epithelial cells. IL-33 is constitutively expressed in epithelial barrier tissues including lung, skin, stomach and salivary gland in steady-state (18,19). IL-33 expression can be further enhanced during inflammation. In allergic lower airway inflammation, IL-33 production is increased in the airway epithelial cells in asthma and as chronic obstructive pulmonary disease (COPD) (20,21). The expression of IL-33 has also been observed in hepatocytes during acute hepatitis and in endothelial cells from the colitis tissue, in macrophages and dendritic cells during allergic inflammation and infection (22-24). However, epithelial cells appear to be the most important major source of IL-33 as the level is suspected much higher in epithelial cells (21,23).

Biologically active full-length IL-33 plays a role in mucosal inflammation recruiting neutrophils via chemokines including C-X-C motif chemokine ligand (CXCL)-1 and CXCL-2 (25-27). Several studies sought to investigate the expression and role of IL-33 in CRS. There have been conflicting results on the expression of IL-33 in CRS. It was reported that IL-33 mRNA was highly expressed in nasal mucosa but was not elevated in NP or other inflamed areas of the sinuses in CRSwNP (28-30). *In vitro* cell culture study, epithelial cell culture from NP, IL-33 mRNA levels were 3-fold higher in the recalcitrant group than the responsive group, and it was far more enhanced by CpG stimulation (31). A significant upregulation of ST2, a receptor for IL-33 was demonstrated in ethmoid mucosa from CRSwNP, but the concentration of IL-33

protein was not significantly different between NP and control tissue (32). Of note, the recent research conducted by our group demonstrated IL-33 was upregulated in neutrophil-dominant subtypes of CRSwNP compared to eosinophilic NP or controls in the Korean population (33). Accordingly, IL-33 level in tissue homogenate was correlated with Th1/Th17 inflammatory cytokines (33). However, in this study, protein levels of IL-5 and quantities of eosinophil infiltration were inversely related with levels of IL-33 which was shown in the previous studies conducted in the countries of eosinophil-dominant NPs, suggesting the difference of IL-33 role and immune mechanisms in subtypes of CRSwNP that shows regional differences.

IL-25 has been demonstrated to induce Th2 type immune responses. IL-25 is produced by eosinophils, mast cells, and airway epithelial cells (34,35) and stimulates Th2 airway inflammation characterized by airway hyperreactivity, mucus production, airway eosinophilia, and increased serum IgE (36,37). A recent investigation reported increased expression of IL-25 and its receptor in the airways of asthmatics after allergen provocation (38).

In CRS, some groups reported IL-25mRNA levels in CRSwNP, which was controversial. Our recent study showed IL-25 expression was upregulated at the protein level in the tissue of CRSwNP homogenate compared to normal healthy controls. These controversies are needed to be more investigated, however, given that Th1/Th2/Th17 inflammatory markers were positively correlated with IL-25mRNA level in Asian NP, the subtypes and regional or racial differences and severity of disease should be further elucidated in the future.

## INNATE LYMPHOID CELLS (ILCS)

ILCs are RAG-independent cells with the absence of lineage markers for T/B cells and for other innate cells with lymphoid morphology (39). Three ILC groups have been identified in both mouse and humans, related to their dependence on the transcriptional repressor inhibitor of DNA binding 2 (Id2) and the IL-2R $\gamma$  chain (40). ILC2 is characterized by the capacity to produce large amounts of IL-5 and IL-13 when activated by the pro-Th2 cytokines IL-25, IL-33 or thymic stromal lymphopoietin (TSLP). Unlike T cells, these responses do not require antigen for activation, and they rapidly mobilize these cytokines, as named with 'innate.' ILC2s are regarded as innate counterparts of Th2 cells sharing same functional module by their mutual production of signature cytokines such as IL-5 and IL-13 (41).

About the role of ILCs in the pathogenesis of CRS, there are a vast amount of researches that are going on. ILC2s are most thoroughly studied in the context of lung immune cell homeostasis, immunity, and inflammation. In recent report, ILC2 are the central players in CRS pathogenesis. As ILC2s have been identified as a major innate producer of IL-13 and IL-5, in response to IL-25 and IL-33 expression (42,43), its role in NP pathogenesis has been suggested.

In CRSwNP mucosa, it has been reported chemoattractant receptor homologous molecule expressed on Th2 cells (CRTH2)<sup>+</sup>ST2<sup>+</sup> ILC2s are increased (44). TSLP activates human ILC2 by directly upregulating GATA3 and signal transducer and activator of transcription (STAT) 5, resulting in the production of high amounts of type 2 cytokines (44,45). This observation is highly relevant in the context of CRS and also asthma, because TSLP protein expression was significantly increased in epithelial cells derived from NP of CRS patients and in the airway epithelium and lamina propria of asthmatic patients, particularly in patients with severe forms of diseases (46).

IL-33- and IL-25-activated ILC2s can induce eosinophilic airway inflammation accompanied by airway hyper-responsiveness even in Rag knockout mice, which means ILC2s function independent of acquired immunity (47,48). ILC2s are abundant and also have a close relationship with higher tissue and blood eosinophilia in NP, clinically related to worsening nasal symptom scores and asthma comorbidity (49,50). A recent study reported that there was spatial co-localization between ILC2s and eosinophils in NP. A co-culture of eosinophils and ILC2s augmented the activation of eosinophils and prolonged their survival, and in return, pre-activated eosinophils enhanced IL-5 production of ILC2s in an IL-4 dependent manner.

CRTH2 is the one of marker for human ILC2 (39). CRTH2 is known as the prostaglandin D2 (PGD2) receptor 2, binds PGD2 to induce eosinophilic Th2 inflammation, including ILC2 chemotaxis (51-53). PGD2 induces the production of type 2 cytokines, IL-4, IL-5, and IL-13 in ILC2 by the activation of CRTH2. Mast cells are the most important source of PGD2 in NPs. Activated mast cell supernatant induced migration of ILC2 and Th2 cytokine production in ILC2 through CRTH2 (52). Mast cells produce other pro-inflammatory arachidonic acid metabolites cysteinyl leukotrienes (CysLTs) including LTC4 and LTD4 (54). ILC2 also expresses CysLT receptor 1, and CysLTs strongly induced type 2 cytokines in ILC2s (55). Given that mast cells were increased and activated in CRSwNP, and often found activated and degranulated in eosinophilic NPs (56), mast cells have potential to enhance ILC2 to induce type 2 immune responses in NPs.

It has been reported that IL-25 induces corticosteroid resistance through the induction of ILC2 in allergic pulmonary inflammation model (57). IL-25 induced-ILC2 transferring was sufficient to induce pulmonary allergic inflammatory responses.

In allergic asthma model induced by allergen co-administered with IL-33, the ILC2-related TSLP-STAT5 pathway has been suggested to be critical for steroid resistance to airway inflammatory disease. Steroid resistance of ILC2s was recently reported as TSLP dependent (58), where IL-33 in combination with antigen resulted in ILC2s with increased resistance to steroid treatment than T cells. This process required STAT5 and could be inhibited with pimozide (a STAT5-inhibiting antipsychotic drug). Treatment-recalcitrant ILC2 may contribute to disease heterogeneity which might lead to recurrence and exacerbation of CRSwNP.

The role of ILCs in non-eosinophilic CRSwNP was not clear, and there are very few report that ILC3s are more frequently found in CRSwNP. In the recent study, the role of ILC3 in obesity-associated asthma was reported. Airway hyperreactivity in obesity-related asthma mouse model was dependent on IL-17A and the NLRP3 inflammasome, and the possible link with ILC3 was demonstrated that this phenotype could be adoptive transferred by IL-17 producing ILCs. Though the role of ILC3 in NP pathogenesis is not clear, this suggests that ILC3s have the potentials in non-eosinophilic NP pathogenesis, which needs further investigations (59).

## MACROPHAGE

Macrophages are generated in the bone marrow by progenitor cells. The life-span of macrophage is varying from hours to possibly years depending on the type of immune response. Tissue macrophages are differentiated from circulating monocytes when they enter tissue. Tissue macrophages have a role functioning as the phagocyte ingesting pathogens, a scavenger of dead cells and debris, and a leading player of tissue remodeling after damage (60,61).

Based on their function, macrophages are subdivided into M1 macrophage, mediating host defense and anti-tumor immunity, and M2 macrophage, suppressing immune response with regulating wound healing and some other macrophages that have immunosuppressive activity, regulatory macrophage that secrete IL-10, tumor-associated macrophages that suppressed anti-tumor immune response, and the monocytic subset of myeloid-derived suppressor cells. Macrophages can have important roles in both immune induction and resolution (62).

Focusing on the chronological order of CRSwNP, comparing mature ethmoidal polyps or normal mucosa, subepithelial eosinophils and M2 type macrophages were significantly increased in numbers in the early stage polyps confined to middle turbinate (63).

In a recent report, it was demonstrated that group V phospholipase A2 and transglutaminase-2 was co-localized in macrophages of human NP tissue obtained from patients with Th2 type eosinophilic inflammation, and their co-expression positively correlated with the number of eosinophils in each tissue specimen (64). In human M2 macrophage activated with IL-4, group V phospholipase A2 PLA2G5 regulates transglutaminase activity of human IL-4-activated M2 macrophages through PGE2 generation. Group V phospholipase A2 is a functionally relevant enzyme that may have therapeutic value for the treatment of human Th2 inflammatory CRSwNP.

The number of CD68<sup>+</sup> CD163<sup>+</sup> alternatively activated (M2) macrophages was increased in eosinophilic NP. In eosinophilic Th2 milieu, M2 macrophage expresses tumor necrosis factor- $\alpha$ -induced protein 8-like 2 (TIPE2), a new candidate of immune modulating protein, by IL-4 and IL-13 stimulation. TIPE2<sup>+</sup> CD163<sup>+</sup> CD68<sup>+</sup> M2 macrophage correlated with eosinophilic inflammation and clinical refractoriness (65). In tissue remodeling by Th2-induced coagulopathy, M2 macrophages, alternatively activated by Th2 cytokines (66), produce coagulation factor XIIIa which acts enzymatically and contribute to forming a tight tetrameric complex (FXIIIa2B2), cross-linking process of fibrin, culminating in edematous remodeling patterns in NP (67).

## NEUTROPHIL

Neutrophils have long been described in the phenotype studies of CRSwNP. Neutrophil-dominant CRSwNP were reported mainly in Asian NP. CRSwNP was initially classified according to the presence of NP which reflects eosinophils-infiltrating Th2 inflammation predominantly at the molecular levels in Western countries (68). However, Asian populations such as in China showed multiple inflammatory cells infiltrate, including neutrophils and T cells classified as Th1/Th17 cells (10). Furthermore, mixed Th17/Th2 inflammatory patterns were also demonstrated throughout a single NP tissue, and even eosinophilic NPs encompass neutrophils and expresses its biological markers such as IL-8 and myeloperoxidase (3).

Similar to M1 and M2 in subpopulations of macrophages, neutrophils are grouped into either N1 or N2. Like the functions in the subpopulation of M1 and M2 macrophage, N1 and N2 neutrophils are functioning as N1 to be pro-inflammatory, whereas the N2 as an anti-inflammatory. N1 neutrophils have been shown to express more pro-inflammatory cytokines and chemokines, such as tumor necrosis factor (TNF), IL-1 $\beta$ , C-C motif chemokine ligand (CCL) 3, CCL5, IL-6, and IL-12. Compared to N1 neutrophil, N2 neutrophils are the



immunosuppressive counterpart. N2 neutrophils are related to tumor progression, metastasis, and like M2 macrophage, are the players in tissue repair and inflammation-resolving process. Tumor-associated N2 neutrophils are characterized by high expression of C-X-C chemokine receptor type 4 (CXCR4), vascular endothelial growth factor and matrix metalloproteinase-9 and are induced in the presence of high levels of transforming growth factor- $\beta$  (TGF- $\beta$ ).

The polarization and markers for N1 and N2 were more studied in mice. Till now, there are no clear-cut phenotypical markers for neutrophilic subsets, however, based on several studies mainly in mice, have shown part of results using similar markers that distinguish M2 macrophages (69,70). It will be similar to M2 macrophage: express the mannose receptor C-type 1 (MRC1), arginase 1 (ARG1), chitinase-like 3 (Ym1), IL-10, and TGF- $\beta$ .

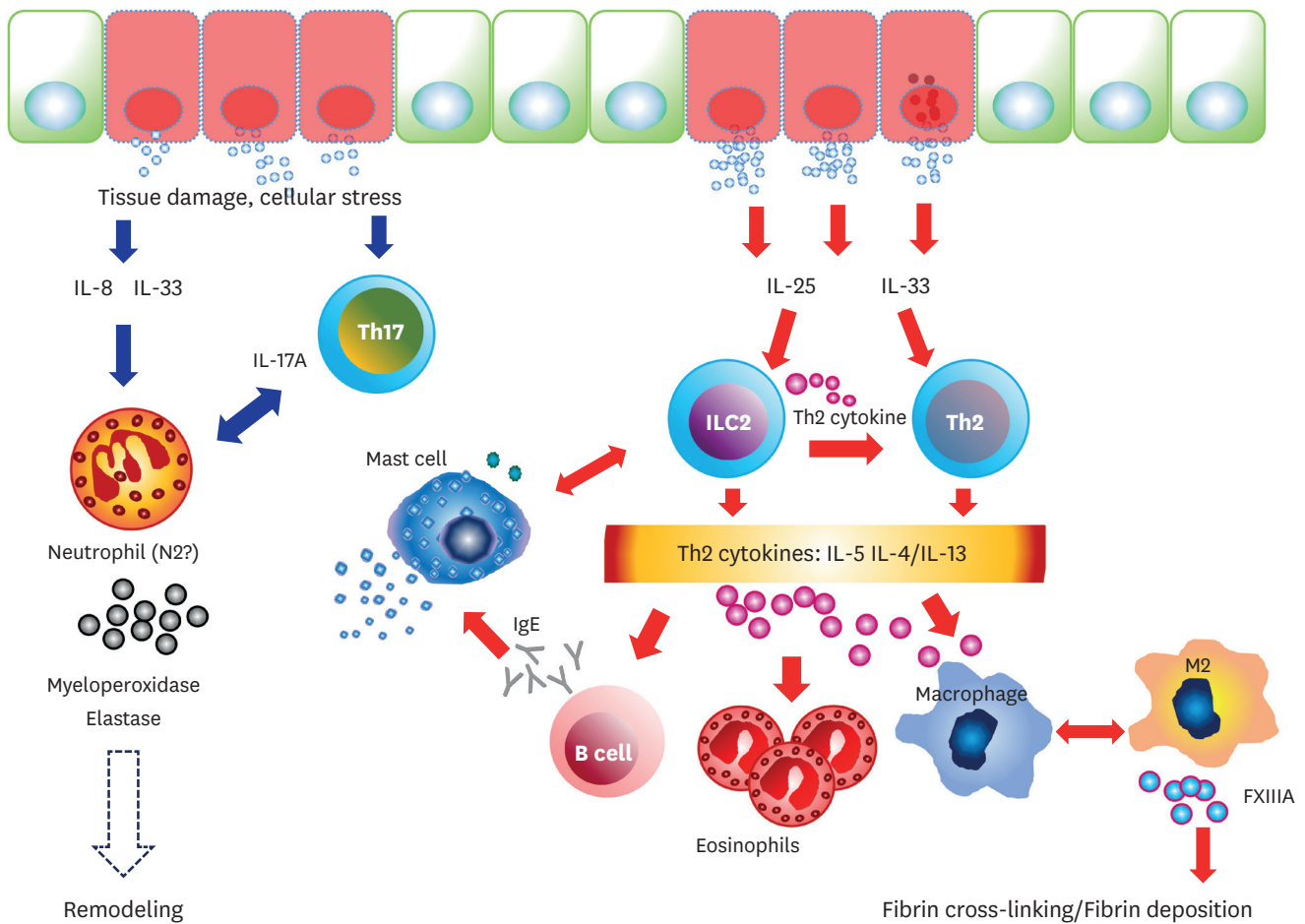
Recently the pathogenic role of neutrophils in CRSwNP was reported disrupting epithelial barrier by producing oncostatin M (OSM) (71,72). These studies demonstrated OSM-producing neutrophil by flow cytometry analysis and immunofluorescence imaging. This neutrophil population produces OSM by the stimulation with granulocyte-macrophage colony stimulating factor (GM-CSF), however, not by the stimulation with IL-25, IL-33, or TSLP. It was suggested neutrophil itself could produce GM-CSF, thus there exists autocrine pathway to secrete OSM in NP. These OSM-producing neutrophils from NP expressed the Th2 type immune response marker ARG1 analyzed by FACS, indicating a phenotype resembling N2 neutrophils more than N1 neutrophils.

Neutrophils are traditionally regarded as the cell of acute inflammation with relatively short life span in tissue, and the reason and the role of accumulation of neutrophil in CRSwNP pathogenesis are not known (73). The similar chronic neutrophilic inflammation is observed in lung diseases such COPD or cystic fibrosis (CF). The mechanism of prolonged excessive neutrophil accumulation is suspected to be driven by tripeptide N-acetyl Pro-Gly-Pro (PGP), the product derived from the breakdown of the extracellular matrix of lung tissue in the disease state (74). Neutrophil product secreted in the inflammatory process may attribute this process, including various proteolytic enzymes that may alter the protease-antiprotease balance. Additionally, cigarette smoking selectively inhibits leukotriene A4 hydrolase aminopeptidase activity, which naturally degrades PGP and resolving inflammation, resulting in the accumulation of PGP and neutrophils (75). However, whether this process is taken place in neutrophil-dominant CRSwNP is largely unknown.

Neutrophils are not classically associated with Th2 immune responses and are considered a Th1 immune response effector cell. However, although the previous study has been conducted with NP in Western countries which tend to be highly eosinophilic, they had significant numbers of neutrophils (71). In parallel, increased neutrophilia within NP has been linked to decreased responsiveness to corticosteroid treatment and neutrophilic subsets of NP are also reported especially in Asian polyps (76,77), neutrophils may contribute to the pathogenesis and treatment resistance of NPs even though little is known.

## CONCLUSION

This article focused on the review of the recent research about key candidate innate immune player cells in the pathogenesis of CRSwNP. The pathogenesis of NP described above is summarized in **Fig. 1**.



**Figure 1.** Summary of innate immune cell responses in chronic rhinosinusitis with NP.

Targeting acquired immunity and mainstay of research targeting Th2 inflammation has been progressed actively, launching the new treatment targets such as monoclonal antibodies against IL-5, IL-4 receptor alpha subunit, IgE, and TSLP in recent clinical trials. The therapies targeting Th2 inflammation showed the beneficial effect especially on eosinophilic NP and/or asthma.

Although the majority of CRSwNP cases in western countries show type 2 inflammation and eosinophilia, non-eosinophilic CRSwNP is more common in Asian countries (3,9). Given that a worse prognosis for steroid therapy in non-eosinophilic CRSwNP versus eosinophilic CRSwNP was demonstrated (76), more studies on innate inflammatory molecules in non-eosinophilic CRSwNP can develop novel therapeutic targets for recalcitrant CRSwNP. CRSwNP is a heterogeneous disease and more precise sub-classification concerning the inflammatory mechanism reflecting prognosis and predicting target treatment modality of this disease might be required in the future.

## ACKNOWLEDGEMENTS

This research was supported by grant support from Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science and ICT (NRF-2016R1C1B1014775 to Il Gyu Kong and NRF-2017R1A2B4003445 to Dae Woo Kim).

## REFERENCES

1. Fokkens WJ, Lund VJ, Mullol J, Bachert C, Alobid I, Baroody F, Cohen N, Cervin A, Douglas R, Gevaert P, et al. EPOS 2012: European position paper on rhinosinusitis and nasal polyps 2012. A summary for otorhinolaryngologists. *Rhinology* 2012;50:1-12.  
[PUBMED](#) | [CROSSREF](#)
2. Kim DH, Han K, Kim SW. Effect of chronic rhinosinusitis with or without nasal polyp on quality of life in South Korea: 5th Korea National Health and Nutrition Examination Survey Korean. *Clin Exp Otorhinolaryngol* 2016;9:150-156.  
[PUBMED](#) | [CROSSREF](#)
3. Wang X, Zhang N, Bo M, Holtappels G, Zheng M, Lou H, Wang H, Zhang L, Bachert C. Diversity of TH cytokine profiles in patients with chronic rhinosinusitis: a multicenter study in Europe, Asia, and Oceania. *J Allergy Clin Immunol* 2016;138:1344-1353.  
[PUBMED](#) | [CROSSREF](#)
4. Blackwell DL, Lucas JW, Clarke TC. Summary health statistics for U.S. adults: national health interview survey, 2012. *Vital Health Stat 10* 2014;1:161.  
[PUBMED](#)
5. Huvenne W, van Bruaene N, Zhang N, van Zele T, Patou J, Gevaert P, Claeys S, Van Cauwenberge P, Bachert C. Chronic rhinosinusitis with and without nasal polyps: what is the difference? *Curr Allergy Asthma Rep* 2009;9:213-220.  
[PUBMED](#) | [CROSSREF](#)
6. Kim DW, Kim JY, Jeon SY. The status of the olfactory cleft may predict postoperative olfactory function in chronic rhinosinusitis with nasal polyposis. *Am J Rhinol Allergy* 2011;25:e90-e94.  
[PUBMED](#) | [CROSSREF](#)
7. Lou H, Meng Y, Piao Y, Wang C, Zhang L, Bachert C. Predictive significance of tissue eosinophilia for nasal polyp recurrence in the Chinese population. *Am J Rhinol Allergy* 2015;29:350-356.  
[PUBMED](#) | [CROSSREF](#)
8. Bachert C, Akdis CA. Phenotypes and emerging endotypes of chronic rhinosinusitis. *J Allergy Clin Immunol Pract* 2016;4:621-628.  
[PUBMED](#) | [CROSSREF](#)
9. Kim DK, Park MH, Chang DY, Eun KM, Shin HW, Mo JH, Shin EC, Jin HR, Shin S, Roh EY, et al. MBP-positive and CD11c-positive cells are associated with different phenotypes of Korean patients with non-asthmatic chronic rhinosinusitis. *PLoS One* 2014;9:e111352.  
[PUBMED](#) | [CROSSREF](#)
10. Zhang N, Van Zele T, Perez-Novo C, Van Bruaene N, Holtappels G, DeRuyck N, Van Cauwenberge P, Bachert C. Different types of T-effector cells orchestrate mucosal inflammation in chronic sinus disease. *J Allergy Clin Immunol* 2008;122:961-968.  
[PUBMED](#) | [CROSSREF](#)
11. Mahdavinia M, Suh LA, Carter RG, Stevens WW, Norton JE, Kato A, Tan BK, Kern RC, Conley DB, Chandra R, et al. Increased noneosinophilic nasal polyps in chronic rhinosinusitis in US second-generation Asians suggest genetic regulation of eosinophilia. *J Allergy Clin Immunol* 2015;135:576-579.  
[PUBMED](#) | [CROSSREF](#)
12. Liao B, Cao PP, Zeng M, Zhen Z, Wang H, Zhang YN, Hu CY, Ma J, Li ZY, Song J, et al. Interaction of thymic stromal lymphopoietin, IL-33, and their receptors in epithelial cells in eosinophilic chronic rhinosinusitis with nasal polyps. *Allergy* 2015;70:1169-1180.  
[PUBMED](#) | [CROSSREF](#)
13. Comeau MR, Ziegler SF. The influence of TSLP on the allergic response. *Mucosal Immunol* 2010;3:138-147.  
[PUBMED](#) | [CROSSREF](#)
14. Kouzaki H, Iijima K, Kobayashi T, O'Grady SM, Kita H. The danger signal, extracellular ATP, is a sensor for an airborne allergen and triggers IL-33 release and innate Th2-type responses. *J Immunol* 2011;186:4375-4387.  
[PUBMED](#) | [CROSSREF](#)
15. Ziegler SF. Thymic stromal lymphopoietin and allergic disease. *J Allergy Clin Immunol* 2012;130:845-852.  
[PUBMED](#) | [CROSSREF](#)
16. Cayrol C, Girard JP. IL-33: an alarmin cytokine with crucial roles in innate immunity, inflammation and allergy. *Curr Opin Immunol* 2014;31:31-37.  
[PUBMED](#) | [CROSSREF](#)
17. Licona-Limón P, Kim LK, Palm NW, Flavell RA. TH2, allergy and group 2 innate lymphoid cells. *Nat Immunol* 2013;14:536-542.  
[PUBMED](#) | [CROSSREF](#)



18. Moussion C, Ortega N, Girard JP. The IL-1-like cytokine IL-33 is constitutively expressed in the nucleus of endothelial cells and epithelial cells *in vivo*: a novel 'alarmin'? *PLoS One* 2008;3:e3331.  
[PUBMED](#) | [CROSSREF](#)
19. Pichery M, Mirey E, Mercier P, Lefrancais E, Dujardin A, Ortega N, Girard JP. Endogenous IL-33 is highly expressed in mouse epithelial barrier tissues, lymphoid organs, brain, embryos, and inflamed tissues: *in situ* analysis using a novel IL-33-LacZ gene trap reporter strain. *J Immunol* 2012;188:3488-3495.  
[PUBMED](#) | [CROSSREF](#)
20. Préfontaine D, Nadigel J, Chouiali F, Audusseau S, Semlali A, Chakir J, Martin JG, Hamid Q. Increased IL-33 expression by epithelial cells in bronchial asthma. *J Allergy Clin Immunol* 2010;125:752-754.  
[PUBMED](#) | [CROSSREF](#)
21. Byers DE, Alexander-Brett J, Patel AC, Agapov E, Dang-Vu G, Jin X, Wu K, You Y, Alevy Y, Girard JP, et al. Long-term IL-33-producing epithelial progenitor cells in chronic obstructive lung disease. *J Clin Invest* 2013;123:3967-3982.  
[PUBMED](#) | [CROSSREF](#)
22. Chang YJ, Kim HY, Albacker LA, Baumgarth N, McKenzie AN, Smith DE, Dekruyff RH, Umetsu DT. Innate lymphoid cells mediate influenza-induced airway hyper-reactivity independently of adaptive immunity. *Nat Immunol* 2011;12:631-638.  
[PUBMED](#) | [CROSSREF](#)
23. Hardman CS, Panova V, McKenzie AN. IL-33 citrine reporter mice reveal the temporal and spatial expression of IL-33 during allergic lung inflammation. *Eur J Immunol* 2013;43:488-498.  
[PUBMED](#) | [CROSSREF](#)
24. Tjota MY, Williams JW, Lu T, Clay BS, Byrd T, Hrusch CL, Decker DC, de Araujo CA, Bryce PJ, Sperling AI. IL-33-dependent induction of allergic lung inflammation by FcγRIII signaling. *J Clin Invest* 2013;123:2287-2297.  
[PUBMED](#) | [CROSSREF](#)
25. Luzina IG, Pickering EM, Kopach P, Kang PH, Lockett V, Todd NW, Papadimitriou JC, McKenzie AN, Atamas SP. Full-length IL-33 promotes inflammation but not Th2 response *in vivo* in an ST2-independent fashion. *J Immunol* 2012;189:403-410.  
[PUBMED](#) | [CROSSREF](#)
26. Mizutani N, Nabe T, Yoshino S. IL-17A promotes the exacerbation of IL-33-induced airway hyperresponsiveness by enhancing neutrophilic inflammation via CXCR2 signaling in mice. *J Immunol* 2014;192:1372-1384.  
[PUBMED](#) | [CROSSREF](#)
27. Lan F, Yuan B, Liu T, Luo X, Huang P, Liu Y, Dai L, Yin H. Interleukin-33 facilitates neutrophil recruitment and bacterial clearance in *S. aureus*-caused peritonitis. *Mol Immunol* 2016;72:74-80.  
[PUBMED](#) | [CROSSREF](#)
28. Shaw JL, Fakhri S, Citardi MJ, Porter PC, Corry DB, Kheradmand F, Liu YJ, Luong A. IL-33-responsive innate lymphoid cells are an important source of IL-13 in chronic rhinosinusitis with nasal polyps. *Am J Respir Crit Care Med* 2013;188:432-439.  
[PUBMED](#) | [CROSSREF](#)
29. Miljkovic D, Bassiouni A, Cooksley C, Ou J, Hauben E, Wormald PJ, Vreugde S. Association between group 2 innate lymphoid cells enrichment, nasal polyps and allergy in chronic rhinosinusitis. *Allergy* 2014;69:1154-1161.  
[PUBMED](#) | [CROSSREF](#)
30. Lam M, Hull L, McLachlan R, Snidvongs K, Chin D, Pratt E, Kalish L, Sacks R, Earls P, Sewell W, et al. Clinical severity and epithelial endotypes in chronic rhinosinusitis. *Int Forum Allergy Rhinol* 2013;3:121-128.  
[PUBMED](#) | [CROSSREF](#)
31. Reh DD, Wang Y, Ramanathan M Jr, Lane AP. Treatment-recalcitrant chronic rhinosinusitis with polyps is associated with altered epithelial cell expression of interleukin-33. *Am J Rhinol Allergy* 2010;24:105-109.  
[PUBMED](#) | [CROSSREF](#)
32. Baba S, Kondo K, Kanaya K, Suzukawa K, Ushio M, Urata S, Asakage T, Kakigi A, Suzukawa M, Ohta K, et al. Expression of IL-33 and its receptor ST2 in chronic rhinosinusitis with nasal polyps. *Laryngoscope* 2014;124:E115-E122.  
[PUBMED](#) | [CROSSREF](#)
33. Kim DK, Jin HR, Eun KM, Mo JH, Cho SH, Oh S, Cho D, Kim DW. The role of interleukin-33 in chronic rhinosinusitis. *Thorax* 2017;72:635-645.  
[PUBMED](#) | [CROSSREF](#)
34. Godin MS, Waldman SR, Johnson CM Jr. Nasal augmentation using Gore-Tex. A 10-year experience. *Arch Facial Plast Surg* 1999;1:118-121.  
[PUBMED](#) | [CROSSREF](#)

35. Iwakura Y, Ishigame H, Saijo S, Nakae S. Functional specialization of interleukin-17 family members. *Immunity* 2011;34:149-162.  
[PUBMED](#) | [CROSSREF](#)
36. Fort MM, Cheung J, Yen D, Li J, Zurawski SM, Lo S, Menon S, Clifford T, Hunte B, Lesley R, et al. IL-25 induces IL-4, IL-5, and IL-13 and Th2-associated pathologies *in vivo*. *Immunity* 2001;15:985-995.  
[PUBMED](#) | [CROSSREF](#)
37. Sharkhuu T, Matthaehi KI, Forbes E, Mahalingam S, Hogan SP, Hansbro PM, Foster PS. Mechanism of interleukin-25 (IL-17E)-induced pulmonary inflammation and airways hyper-reactivity. *Clin Exp Allergy* 2006;36:1575-1583.  
[PUBMED](#) | [CROSSREF](#)
38. Corrigan CJ, Wang W, Meng Q, Fang C, Eid G, Caballero MR, Lv Z, An Y, Wang YH, Liu YJ, et al. Allergen-induced expression of IL-25 and IL-25 receptor in atopic asthmatic airways and late-phase cutaneous responses. *J Allergy Clin Immunol* 2011;128:116-124.  
[PUBMED](#) | [CROSSREF](#)
39. Spits H, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, Koyasu S, Locksley RM, McKenzie AN, Mebius RE, et al. Innate lymphoid cells--a proposal for uniform nomenclature. *Nat Rev Immunol* 2013;13:145-149.  
[PUBMED](#) | [CROSSREF](#)
40. Yokota Y, Mansouri A, Mori S, Sugawara S, Adachi S, Nishikawa S, Gruss P. Development of peripheral lymphoid organs and natural killer cells depends on the helix-loop-helix inhibitor Id2. *Nature* 1999;397:702-706.  
[PUBMED](#) | [CROSSREF](#)
41. Robinette ML, Colonna M. Immune modules shared by innate lymphoid cells and T cells. *J Allergy Clin Immunol* 2016;138:1243-1251.  
[PUBMED](#) | [CROSSREF](#)
42. Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TK, Bucks C, Kane CM, Fallon PG, Pannell R, et al. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature* 2010;464:1367-1370.  
[PUBMED](#) | [CROSSREF](#)
43. Moro K, Yamada T, Tanabe M, Takeuchi T, Ikawa T, Kawamoto H, Furusawa J, Ohtani M, Fujii H, Koyasu S. Innate production of T(H)2 cytokines by adipose tissue-associated c-Kit(+)Sca-1(+) lymphoid cells. *Nature* 2010;463:540-544.  
[PUBMED](#) | [CROSSREF](#)
44. Mjösberg JM, Trifari S, Crellin NK, Peters CP, van Drunen CM, Piet B, Fokkens WJ, Cupedo T, Spits H. Human IL-25- and IL-33-responsive type 2 innate lymphoid cells are defined by expression of CCR2 and CD161. *Nat Immunol* 2011;12:1055-1062.  
[PUBMED](#) | [CROSSREF](#)
45. Mjösberg J, Bernink J, Golebski K, Karrich JJ, Peters CP, Blom B, te Velde AA, Fokkens WJ, van Drunen CM, Spits H. The transcription factor GATA3 is essential for the function of human type 2 innate lymphoid cells. *Immunity* 2012;37:649-659.  
[PUBMED](#) | [CROSSREF](#)
46. Shikotra A, Choy DF, Ohri CM, Doran E, Butler C, Hargadon B, Shelley M, Abbas AR, Austin CD, Jackman J, et al. Increased expression of immunoreactive thymic stromal lymphopoietin in patients with severe asthma. *J Allergy Clin Immunol* 2012;129:104-111.e1-9.  
[PUBMED](#) | [CROSSREF](#)
47. Morita H, Moro K, Koyasu S. Innate lymphoid cells in allergic and nonallergic inflammation. *J Allergy Clin Immunol* 2016;138:1253-1264.  
[PUBMED](#) | [CROSSREF](#)
48. Kim HY, Chang YJ, Subramanian S, Lee HH, Albacker LA, Matangkasombut P, Savage PB, McKenzie AN, Smith DE, Rottman JB, et al. Innate lymphoid cells responding to IL-33 mediate airway hyperreactivity independently of adaptive immunity. *J Allergy Clin Immunol* 2012;129:216-227.e1-6.  
[PUBMED](#) | [CROSSREF](#)
49. Ho J, Bailey M, Zaunders J, Mrad N, Sacks R, Sewell W, Harvey RJ. Cellular comparison of sinus mucosa vs polyp tissue from a single sinus cavity in chronic rhinosinusitis. *Int Forum Allergy Rhinol* 2015;5:14-27.  
[PUBMED](#) | [CROSSREF](#)
50. Ho J, Bailey M, Zaunders J, Mrad N, Sacks R, Sewell W, Harvey RJ. Group 2 innate lymphoid cells (ILC2s) are increased in chronic rhinosinusitis with nasal polyps or eosinophilia. *Clin Exp Allergy* 2015;45:394-403.  
[PUBMED](#) | [CROSSREF](#)
51. Townley RG, Agrawal S. CCR2 antagonists in the treatment of allergic responses involving TH2 cells, basophils, and eosinophils. *Ann Allergy Asthma Immunol* 2012;109:365-374.  
[PUBMED](#) | [CROSSREF](#)

52. Xue L, Salimi M, Panse I, Mjösberg JM, McKenzie AN, Spits H, Klenerman P, Ogg G. Prostaglandin D2 activates group 2 innate lymphoid cells through chemoattractant receptor-homologous molecule expressed on TH2 cells. *J Allergy Clin Immunol* 2014;133:1184-1194.  
[PUBMED](#) | [CROSSREF](#)
53. Chang JE, Doherty TA, Baum R, Broide D. Prostaglandin D2 regulates human type 2 innate lymphoid cell chemotaxis. *J Allergy Clin Immunol* 2014;133:899-901.e3.  
[PUBMED](#) | [CROSSREF](#)
54. Kato A, Chustz RT, Ogasawara T, Kulka M, Saito H, Schleimer RP, Matsumoto K. Dexamethasone and FK506 inhibit expression of distinct subsets of chemokines in human mast cells. *J Immunol* 2009;182:7233-7243.  
[PUBMED](#) | [CROSSREF](#)
55. Doherty TA, Khorram N, Lund S, Mehta AK, Croft M, Broide DH. Lung type 2 innate lymphoid cells express cysteinyl leukotriene receptor 1, which regulates TH2 cytokine production. *J Allergy Clin Immunol* 2013;132:205-213.  
[PUBMED](#) | [CROSSREF](#)
56. Takabayashi T, Kato A, Peters AT, Suh LA, Carter R, Norton J, Grammer LC, Tan BK, Chandra RK, Conley DB, et al. Glandular mast cells with distinct phenotype are highly elevated in chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol* 2012;130:410-420.e5.  
[PUBMED](#) | [CROSSREF](#)
57. Petersen BC, Budelsky AL, Baptist AP, Schaller MA, Lukacs NW. Interleukin-25 induces type 2 cytokine production in a steroid-resistant interleukin-17RB+ myeloid population that exacerbates asthmatic pathology. *Nat Med* 2012;18:751-758.  
[PUBMED](#) | [CROSSREF](#)
58. Kabata H, Moro K, Fukunaga K, Suzuki Y, Miyata J, Masaki K, Betsuyaku T, Koyasu S, Asano K. Thymic stromal lymphopoietin induces corticosteroid resistance in natural helper cells during airway inflammation. *Nat Commun* 2013;4:2675.  
[PUBMED](#) | [CROSSREF](#)
59. Kim HY, Lee HJ, Chang YJ, Pichavant M, Shore SA, Fitzgerald KA, Iwakura Y, Israel E, Bolger K, Paul J, et al. Interleukin-17-producing innate lymphoid cells and the NLRP3 inflammasome facilitate obesity-associated airway hyperreactivity. *Nat Med* 2014;20:54-61.  
[PUBMED](#) | [CROSSREF](#)
60. Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nat Rev Immunol* 2005;5:953-964.  
[PUBMED](#) | [CROSSREF](#)
61. Geissmann F, Gordon S, Hume DA, Mowat AM, Randolph GJ. Unravelling mononuclear phagocyte heterogeneity. *Nat Rev Immunol* 2010;10:453-460.  
[PUBMED](#) | [CROSSREF](#)
62. Murray PJ, Wynn TA. Obstacles and opportunities for understanding macrophage polarization. *J Leukoc Biol* 2011;89:557-563.  
[PUBMED](#) | [CROSSREF](#)
63. Meng J, Zhou P, Liu Y, Liu F, Yi X, Liu S, Holtappels G, Bachert C, Zhang N. The development of nasal polyp disease involves early nasal mucosal inflammation and remodelling. *PLoS One* 2013;8:e82373.  
[PUBMED](#) | [CROSSREF](#)
64. Yamaguchi M, Zacharia J, Laidlaw TM, Balestrieri B. PLA2G5 regulates transglutaminase activity of human IL-4-activated M2 macrophages through PGE2 generation. *J Leukoc Biol* 2016;100:131-141.  
[PUBMED](#) | [CROSSREF](#)
65. Yao Y, Wang ZC, Liu JX, Ma J, Chen CL, Deng YK, Liao B, Wang N, Wang H, Ning Q, et al. Increased expression of TIPE2 in alternatively activated macrophages is associated with eosinophilic inflammation and disease severity in chronic rhinosinusitis with nasal polyps. *Int Forum Allergy Rhinol* 2017;7:963-972.  
[PUBMED](#) | [CROSSREF](#)
66. Krysko O, Holtappels G, Zhang N, Kubica M, Deswarte K, Derycke L, Claeys S, Hammad H, Brusselle GG, Vandenabeele P, et al. Alternatively activated macrophages and impaired phagocytosis of *S. aureus* in chronic rhinosinusitis. *Allergy* 2011;66:396-403.  
[PUBMED](#) | [CROSSREF](#)
67. Takabayashi T, Kato A, Peters AT, Hulse KE, Suh LA, Carter R, Norton J, Grammer LC, Tan BK, Chandra RK, et al. Increased expression of factor XIII-A in patients with chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol* 2013;132:584-592.e4.  
[PUBMED](#) | [CROSSREF](#)
68. Van Zele T, Claeys S, Gevaert P, Van Maele G, Holtappels G, Van Cauwenberge P, Bachert C. Differentiation of chronic sinus diseases by measurement of inflammatory mediators. *Allergy* 2006;61:1280-1289.  
[PUBMED](#) | [CROSSREF](#)

69. Cuartero MI, Ballesteros I, Moraga A, Nombela F, Vivancos J, Hamilton JA, Corbí AL, Lizasoain I, Moro MA. N2 neutrophils, novel players in brain inflammation after stroke: modulation by the PPAR $\gamma$  agonist rosiglitazone. *Stroke* 2013;44:3498-3508.  
[PUBMED](#) | [CROSSREF](#)
70. Ma Y, Yabluchanskiy A, Iyer RP, Cannon PL, Flynn ER, Jung M, Henry J, Cates CA, Deleon-Pennell KY, Lindsey ML. Temporal neutrophil polarization following myocardial infarction. *Cardiovasc Res* 2016;110:51-61.  
[PUBMED](#) | [CROSSREF](#)
71. Pothoven KL, Norton JE, Suh LA, Carter RG, Harris KE, Biyasheva A, Welch K, Shintani-Smith S, Conley DB, Liu MC, et al. Neutrophils are a major source of the epithelial barrier disrupting cytokine oncostatin M in patients with mucosal airways disease. *J Allergy Clin Immunol* 2017;139:1966-1978.e9.  
[PUBMED](#) | [CROSSREF](#)
72. Pothoven KL, Norton JE, Hulse KE, Suh LA, Carter RG, Rocci E, Harris KE, Shintani-Smith S, Conley DB, Chandra RK, et al. Oncostatin M promotes mucosal epithelial barrier dysfunction, and its expression is increased in patients with eosinophilic mucosal disease. *J Allergy Clin Immunol* 2015;136:737-746.e4.  
[PUBMED](#) | [CROSSREF](#)
73. Fokkens WJ, Lund VJ, Mullol J, Bachert C, Alobid I, Baroody F, Cohen N, Cervin A, Douglas R, Gevaert P, et al. European position paper on rhinosinusitis and nasal polyps 2012. *Rhinol Suppl* 2012;23:3 p preceding table of contents, 1-298.  
[PUBMED](#)
74. Weathington NM, van Houwelingen AH, Noerager BD, Jackson PL, Kraneveld AD, Galin FS, Folkerts G, Nijkamp FP, Blalock JE. A novel peptide CXCR ligand derived from extracellular matrix degradation during airway inflammation. *Nat Med* 2006;12:317-323.  
[PUBMED](#) | [CROSSREF](#)
75. Snelgrove RJ, Jackson PL, Hardison MT, Noerager BD, Kinloch A, Gaggari A, Shastry S, Rowe SM, Shim YM, Hussell T, et al. A critical role for LTA4H in limiting chronic pulmonary neutrophilic inflammation. *Science* 2010;330:90-94.  
[PUBMED](#) | [CROSSREF](#)
76. Wen W, Liu W, Zhang L, Bai J, Fan Y, Xia W, Luo Q, Zheng J, Wang H, Li Z, et al. Increased neutrophilia in nasal polyps reduces the response to oral corticosteroid therapy. *J Allergy Clin Immunol* 2012;129:1522-1528.e5.  
[PUBMED](#) | [CROSSREF](#)
77. Cao PP, Li HB, Wang BF, Wang SB, You XJ, Cui YH, Wang DY, Desrosiers M, Liu Z. Distinct immunopathologic characteristics of various types of chronic rhinosinusitis in adult Chinese. *J Allergy Clin Immunol* 2009;124:478-484, 484.e1-2.  
[PUBMED](#) | [CROSSREF](#)