

Review Article



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Conflicts of Interest

The authors declare no potential conflicts of interest.

Abbreviations

AID, activation-induced deaminase; BCR, B cell receptor; GC, germinal center; ICOS, inducible T cell costimulatory; ICOSL, inducible T cell costimulatory ligand; IFN, interferon; iNKT, invariant natural killer T; LPS, lipopolysaccharide; MHC, major histocompatibility complex; MZ, marginal

Expansion and Sub-Classification of T Cell-Dependent Antibody Responses to Encompass the Role of Innate-Like T Cells in Antibody Responses

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ABSTRACT

In addition to T cell-dependent (TD) Ab responses, T cells can also regulate T cell-independent (TI) B cell responses in the absence of a specific major histocompatibility complex (MHC) class II and antigenic peptide-based interaction between T and B cells. The elucidation of T cells capable of supporting TI Ab responses is important for understanding the cellular mechanism of different types of TI Ab responses. Natural killer T (NKT) cells represent 1 type of helper T cells involved in TI Ab responses and more candidate helper T cells responsible for TI Ab responses may also include $\gamma\delta$ T cells and recently reported B-1 helper CD4⁺ T cells. Marginal zone (MZ) B and B-1 cells, 2 major innate-like B cell subsets considered to function independently of T cells, interact with innate-like T cells. Whereas MZ B and NKT cells interact mutually for a rapid response to blood-borne infection, peritoneal memory phenotype CD49d^{high}CD4⁺ T cells support natural Ab secretion by B-1 cells. Here the role of innate-like T cells in the so-called TI Ab response is discussed. To accommodate the involvement of T cells in the TI Ab responses, we suggest an expanded classification of TD Ab responses that incorporate cognate and non-cognate B cell help by innate-like T cells.

Keywords: Antigen-antibody reactions; Innate lymphocytes; Natural killer T-cells; B-1 lymphocyte

INTRODUCTION

The activation of Ag-specific B cells via B cell receptor (BCR) is an essential requirement of the Ab response. However, a sufficient Ab response requires not only BCR-mediated signaling but also participation of CD4⁺ T cells or activation of innate immune receptors on B cells (1-3). Since protein Ag-specific B cells usually require major histocompatibility complex (MHC) class II-restricted Ag presentation and CD4⁺ T cell help, this kind of Ab response is called as T cell-dependent (TD) Ab response (4). On the other hand, Ab responses to sugar or lipid Ags can occur in the absence of CD4⁺ T cells as a T cell-independent (TI) Ab response since those Ags with repetitive epitopes engage multiple BCRs or are frequently associated with pathogen-associated molecular patterns (5). However, the TI Ab responses can be boosted by T cells (6). These T cell-promoted TI responses make it difficult to clearly distinguish between the TD and TI Ab responses. In this review, we would like to discuss the participation of innate or innate-like T cells in the TI Ab responses and the interaction between innate-like T cells and B cells.

zone; NKT, natural killer T; SLAM, signaling lymphocytic activation molecule; TCR, T cell receptor; TD, T cell-dependent; Tfh, follicular helper T; Th, T helper; TI, T cell-independent; TLR, toll-like receptor

Author Contributions

Conceptualization: Kim TJ; Data curation: Kim TJ, Park C; Formal analysis: Park C; Funding acquisition: Kim TJ; Investigation: Kim TJ, Park C; Methodology: Park C; Project administration: Kim TJ; Supervision: Kim TJ; Validation: Kim TJ; Writing - original draft: Kim TJ, Park C; Writing - review & editing: Kim TJ, Park C.

INNATE OR INNATE-LIKE LYMPHOCYTES

Innate T and B lymphocytes have recently attracted much attention. Although different opinions exist regarding innate lymphocytes, in this review, we refer to innate lymphocytes as cells having T cell receptors (TCRs) or BCRs, thereby distinguishing innate lymphoid cells from innate lymphocytes (7). Differently from conventional lymphocytes, innate lymphocytes obtain a memory phenotype, such as rapid cytokine secretion or rapid Ab production, during development in the absence of previous Ag exposure. Their rapid and strong responses support the initial priming of conventional naïve T cells, as exemplified in natural killer T (NKT) cells that bridge innate and adaptive immunity by secreting interferon (IFN)- γ in response to IL-12 produced by dendritic cells (8). As bona fide conventional memory lymphocytes are absent at the time of birth, innate or innate-like lymphocytes must function as first-line defenders in neonates and possibly in adults as well (7). $\gamma\delta$ T, NKT, and B-1a cells are well-established innate lymphocytes that function as both effectors and regulators of immunity (3,9).

B-1 and marginal zone (MZ) B cells – 2 distinct types of innate or innate-like B cells

B-1 and MZ B cells participate cooperatively in the TI Ab response to blood-borne bacteria (10), but they function via different mechanisms. B-1a cells, which are CD5⁺ B-1 cells derived from fetal hematopoietic stem cells and are localized in serosal cavities, are mostly autoreactive and also cross-reactive to common pathogens (11). The stimuli for functional activation of B-1a cells are usually through pattern recognition receptors, such as toll-like receptor (TLR)-4, since BCRs of B-1a cells are chronically activated by autoantigens (12,13). Stimulation of B-1a cells with lipopolysaccharide (LPS) leads to their migration out of the serosal cavities into the spleen and consequent secretion of natural Abs and granulocyte macrophage colony-stimulating factor (14,15). B-1a cells readily migrate into the lung, intestine, or regional lymph nodes upon infection of these organs (16,17). Natural IgM Ab secretion by B-1a cells has been found to be constitutive and dependent on Blimp-1, but Blimp-1-independent IgM Ab secretion by B-1a cells has also been noted, suggesting that B-1a cells do not exist as a single population (18). The subdivision of B-1a cells into homeostatic and induced responding populations has been suggested (19).

In contrast, MZ B cells are pre-activated B cells that are localized around the splenic marginal sinus. They can rapidly respond to blood-borne pathogens that are trapped by MZ macrophages around the sinus (2). The development of MZ B cells is important for proper responses against blood-borne pathogens. The appropriate selection of BCR specificities against common pathogens is essential for MZ B cell responses because the selection of autoreactive MZ B cells can be potentially harmful to the host (20). MZ B cells develop from bone marrow precursor cells through the stages of splenic transitional B and MZ precursor cells (21). The developmental decision from transitional B cells to MZ B or follicular B cells is determined by BCR specificities and Notch signaling. Although MZ B cells have some stem cell-like properties to repopulate themselves, new MZ B cells can develop from transitional B and MZ precursor cells (22).

In summary, B-1 and MZ B cells are the two main populations of B cells responsible for TI Ab responses, but they are regulated by different mechanisms. Here we would like to discuss the roles of innate-like T cells in TI Ab responses by B-1 and MZ B cells.

TD AB RESPONSE

The ultimate outcome of the classical TD Ab responses is the generation of high-affinity Abs via the interaction between germinal center (GC) B cells and follicular helper T (Tfh) cells. It is clear that Tfh cell differentiation is crucial for the GC reaction leading to the selection of high-affinity Ab responses against protein Ags (4). For the GC reaction, activated B and conventional CD4⁺ T cells interact closely in cognate or non-cognate manners, resulting in their respective differentiation into GC B and Tfh cells through Bcl-6 upregulation (23). Although the CD40–CD40L interaction is essential for the GC reaction, the GC reaction requires more T cell–B cell interactions via inducible T cell costimulatory (ICOS)–inducible T cell costimulatory ligand (ICOSL) and signaling lymphocytic activation molecule (SLAM)–SLAM interactions (24). Many T cell–B cell interactions fail to progress to the GC reaction, instead resulting in extrafollicular Ab responses (25). Targeted ablation of Tfh cells via CD4⁺ T cell-specific deletion of Bcl-6 could not inhibit Ab responses as effector CD4⁺ T cells, such as T helper (Th) 1 and Th2 cells, could promote extrafollicular TD Ab responses (26,27). Extrafollicular TD Ab responses lead to the generation of Ag-specific B cells with a low level of somatic hypermutation, but the affinity constants of Abs generated from these responses are far lower than those of Abs generated by the Tfh cell-dependent GC reaction and affinity maturation (27). Therefore, the TD Ab responses can occur with or without the GC reaction and the Tfh cell differentiation.

TI AB RESPONSE

TI Ab responses are conventionally divided into type-1 (TI-1) and type-2 (TI-2) responses. Whereas TI-1 Ab responses require the engagement of pattern recognition receptors, such as TLR-4 or TLR-9, in addition to BCR engagement, TI-2 responses require the engagement of at least 10 molecules of BCRs by highly repetitive Ags (28). The fact that TI Ab responses can occur in the absence of T cells does not mean that T cells do not play a role in these responses. The involvement of T cells in TI Ab responses, especially TI-2 responses, has been well established (6,29). Furthermore, T cell involvement in TI Ab responses is accompanied by class switching and a low level of somatic hypermutation, which are dependent on the expression of activation-induced deaminase (AID) (30). In fact, AID expression is upregulated during extrafollicular TD and TI Ab responses and is thus not unique to the GC reaction (25). Interestingly, B-1a cells are known to upregulate AID expression upon LPS stimulation (31,32). The identities of T cell subsets in the TI Ab responses are discussed in detail in the subsequent sections.

Because infectious agents contain both TD and TI Ags, both TD and TI Ab responses proceed concurrently upon infection in most cases. Thus, TD Ag-specific B cells can be initially activated by innate stimuli or the complement pathway and proliferate in a TI manner, thereby blurring the distinction between TD and TI Ab responses (33). Considering the extremely rare chance of cognate interactions between conventional Ag-specific B and T cells, initial TI B cell responses may facilitate TD Ab responses and enhance the possibility of cognate interactions between rare Ag-specific B and T cells via the expansion of Ag-specific B cells (1). Interestingly, early IgM secretion by B-1a cells or initially stimulated B cells can enhance the overall Ab response via promotion of Ag presentation by dendritic cells (34) or positive feedback through FcμR (35). Therefore, TD Ab responses can be supported by early TI Ab responses that can be actually supported by T cells.

INVOLVEMENT OF INNATE OR INNATE-LIKE T CELLS IN TI AB RESPONSE

NKT cells in the MZ B cell immune response

NKT cells have direct and indirect effects on B cell Ab responses. First, NKT cells amplify inflammatory or cytokine responses. They rapidly secrete IL-4 or IFN- γ in response to IL-12 produced by dendritic cells during early infection and facilitate adaptive immune responses of conventional T and B cells (8,36). α -Galactosylceramide, an NKT cell agonist presented within CD1d, can be used as an adjuvant the TD Ab responses (37). Notably, NKT cells can form a cognate interaction with CD1d-expressing B cells. In particular, MZ B cells more highly express CD1d than other subsets of B cells, making them excellent Ag-presenting cells for NKT cells (38).

When a given B cell is specific to a lipid Ag that is presented within CD1d, an NKT cell can provide cognate help to the B cell (39,40). The CD1d-restricted cognate interaction between NKT and B cells leads to the development Bcl-6-expressing follicular helper NKT cells, but interestingly, long-lived plasma cells are not generated by this CD1d-mediated cognate interaction (41). This response has been described as the TD type 2 (TD-2) Ab response because this is different from the classical TD response and usually fails to generate high-affinity Ag-specific B cells (42).

When a given B cell specific to a protein Ag is activated by the specific Ag in combination with innate stimuli, such as LPS, the activated B cell can present endogenously generated NKT cell ligands as well as peptides (43). This suggests that NKT cells have the potential to support B cells with different types of BCR specificities in both TD and TI Ab responses independently of the cognate interaction via MHC class II. It should be noted that NKT cells can provide non-cognate help for both MZ B cell TI and follicular B cell TD Ab responses because NKT cells are recruited to the regional lymph nodes under inflammatory conditions and provide IL-4 (36,44). For high-affinity Ab TD responses, an exact MHC class II-mediated cognate interaction is crucial, and the CD1d-mediated cognate interaction cannot replace the requirement of the MHC class II-mediated cognate interaction. In summary, NKT cells can support the CD1d-dependent TI Ab response via a cognate NKT-B cell interaction or in a non-cognate manner and boost the classical TD Ab response by supplying cytokines and costimulatory molecules.

$\gamma\delta$ T cells and autoreactive GCs

The GC reaction can be induced by $\gamma\delta$ T cells in $\alpha\beta$ T cell-deficient mice (45,46). $\gamma\delta$ T cells have been found in some GCs of humans, sheep, and TCR $\alpha^{-/-}$ mice (47). The V γ 9V δ 2 T cells, the most common human $\gamma\delta$ T cells in human peripheral blood, are activated by butyrophilin 3A1 that binds to phosphorylated metabolites derived from microbes and transformed cells (48) and secrete inflammatory cytokines and enter into GCs themselves, becoming $\gamma\delta$ Tfh cells (49). However, $\gamma\delta$ T cell-supported GCs are fewer than those in the classical TD Ab response. The GC formation is reduced in athymic *nu/nu* mice, but recovers to normal levels after adoptive transfer of conventional $\alpha\beta$ T cells (50). Remarkably, this $\gamma\delta$ T cell-driven GC response was induced by repeated parasitic infections and resulted in an enhancement of autoreactive B cells instead of pathogen-specific B cells (51). This GC reaction appears to be unique in that the $\gamma\delta$ T cells provide help for autoreactive B cells in a non-cognate fashion. This implicates the importance of $\gamma\delta$ T cells in the pathogenesis of autoimmune diseases

such as systemic lupus erythematosus and B cell dysfunction in acquired immune deficiency, as spontaneously developed GCs harbor autoreactive B cells with somatic hypermutations (52). At present, the cellular and molecular mechanisms underlying this interaction are not well understood. It would be interesting to address the identities of B cell-helping $\gamma\delta$ T cells and whether innate B cells are involved in the collaboration with $\gamma\delta$ T cells.

B-1 helper T cells in B-1a cell immune response

B-1 cells are divided into CD5⁺CD11b⁺ B-1a and CD5⁻CD11b⁺ B-1b cell types, which develop from fetal and adult hematopoietic stem cells, respectively (11). B-1a cells are thought to produce natural Abs in a TI manner, as innate stimuli or cytokines, such as IL-5, induce Ab production (53). Many carbohydrate and lipid Ags are believed to be recognized by B-1a cells, as noted in a report on B-1a cells expressing receptors for blood group A carbohydrates (54). Several B-1b cell Ags have been reported (55), and reportedly, B-1b cells form a TI memory against *Borrelia hermsii* (56). The involvement of T cells in B-1 cell Ab responses is not well investigated, but an active interaction between B-1 and CD4⁺ T cells is plausible because B-1 cells are excellent Ag-presenting cells for T cells (57). The combination of IL-4, IL-5, and the CD40–CD40L interaction was suggested to be a mechanism underlying CD4⁺ T cell help for B-1a cells (58).

NKT cells were thought to be good candidates as helpers of B-1a cells, according to a previous finding that NKT cells are helpers of B cells expressing BCRs for blood group A carbohydrates (59). However, in the case of response to α (1,3) Gal epitopes, the requirement of conventional CD4⁺ T cells in addition to NKT cells was demonstrated (60). Therefore, both conventional CD4⁺ T and NKT cells are plausible candidate helpers for B-1 cell Ab responses. Previously, we attempted to identify B-1a cell subpopulations for effector Ab-secreting function and/or repopulation with stem cell-like property and observed that B-1a cells conjugated to CD4⁺ T cells were superior in terms of IgM Ab production (61). The serosal CD4⁺ T cells contained a unique memory phenotype T cells that expressed a high level of CD49d (integrin $\alpha 4$) and developed spontaneously before 2 weeks of age. Upon stimulation with phorbol myristate acetate and ionomycin, these cells rapidly secreted Th1-type cytokines, such as IFN- γ , tumor necrosis factor- α , and IL-2. The capability of these cells to provide B-1a cell help was clearly revealed in the experiments with co-adoptive transfer of B-1a cells and serosal CD49d^{high}CD4⁺ T cells into lymphocyte-deficient mice and co-culture of these two types of cells. The CD49d^{high}CD4⁺ T cells expressed high levels of integrin $\alpha 4\beta 1$ and $\alpha 6\beta 1$, suggesting their capability to enter peripheral inflammatory sites and migrate via interaction with laminins (62).

We assume that the serosal CD49d^{high}CD4⁺ T cells are B-1 helper T cells capable of boosting B-1 cell secretion of natural Abs. A similar CD49d^{high}CD4⁺ T cell population was also noted in humans (63). At this point, the mechanism by which these CD49d^{high}CD4⁺ T cells assist B-1a cells is unknown. Different cellular and molecular mechanisms are possible, including: 1) a bystander interaction through costimulatory molecules, such as the pairs CD40–CD40L and ICOS–ICOSL independent of TCR–MHC class II; 2) TCR recognition of idiotypic peptides derived from immunoglobulin heavy chain forming an Ab idiotype–anti-idiotype network (64); or 3) MHC class II-dependent TCR recognition of non-peptide Ags, such as glycosylated MHC class II (65). In any case, the functional mechanism of B-1 cell help by these innate-like CD4⁺ T cells needs to be investigated in the future.

SUGGESTION OF A NEW SUB-CLASSIFICATION OF TD AB RESPONSES

In this review, we discussed the role of innate-like T cells in TI Ab responses, which hampers the clear distinction between TD and TI Ab responses in *in vivo* humoral responses. The classical TD Ab response generates high-affinity Ab through the GC reaction and requires absolute involvement of T_{fh} cell differentiation. However, the TD Ab response also requires preceding multiple interactions between activated B and T cells, many of which lead to extrafollicular Ab responses. Considering multiple T-B interactions, we propose to discriminate the T-B interactions leading to the GCs or extrafollicular Ab responses as TD-1 or TD-2 Ab responses, respectively (**Fig. 1**). We suppose that the GC-generating TD-1 response is always accompanied with the extrafollicular TD-2 responses and that some Ab responses are predominated by extrafollicular TD-2 responses without the GC reaction. Thus, our definition of the TD-2 response is different from the TD-2 Ab response that had been previously suggested to refer to NKT cell help to B cells via CD1d-mediated cognate interaction (53). The newly suggested TD-2 response is extrafollicular and can involve both conventional CD4⁺ T and NKT cells. To expand the concept of the TD Ab responses, we suggest a new definition of a TD-3 Ab response, in which NKT or MHC class II-restricted T cells provide help for B cells in a non-cognate manner (**Table 1**). The difference between TD-2 and TD-3 responses

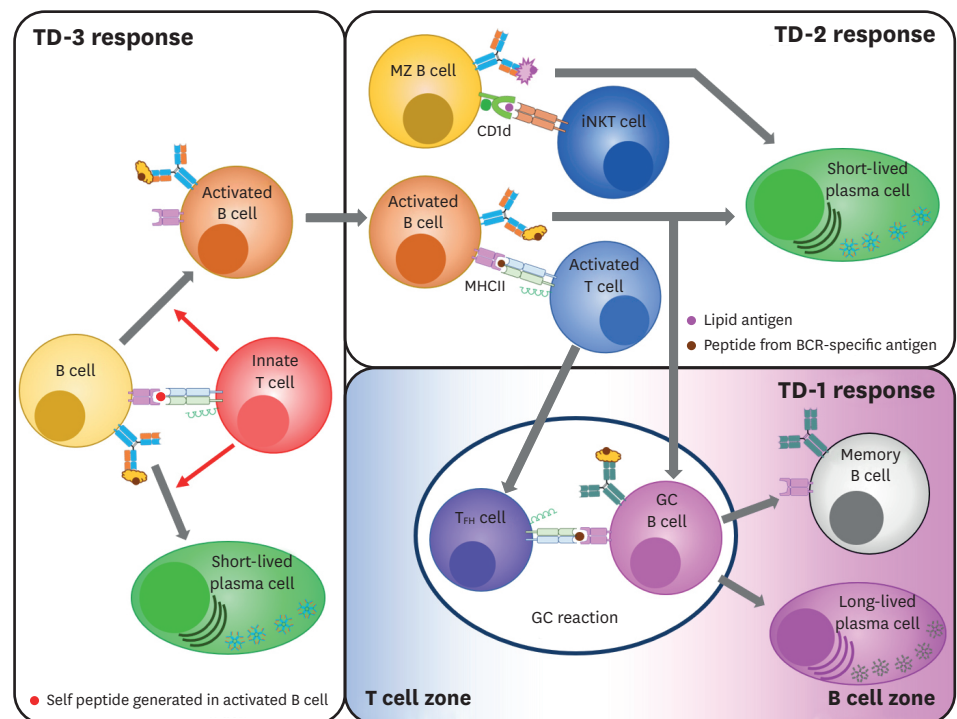


Figure 1. The sub-classification of the TD Ab responses. In the left panel (depicted as TD-3 response), a B cell is activated by an innate stimulus (not depicted, a TI response) and/or an innate-like T cell through non-cognate interaction involving TCRs and Ag-presenting molecules, such as MHC or CD1d (not depicted), irrespective of the BCR specificity (TD-3 response). The activated B cell can proliferate and advance further to interact with other T cells, eventually resulting in a cognate interaction involving TCRs and peptides derived from BCR-bound proteins. This interaction may lead to an extrafollicular Ab response (TD-2 response, right upper) or eventual GC formation (TD-1 response, right lower). iNKT cell can provide cognate help for MZ B cells, leading to extrafollicular MZ B cell response as shown in right upper corner.

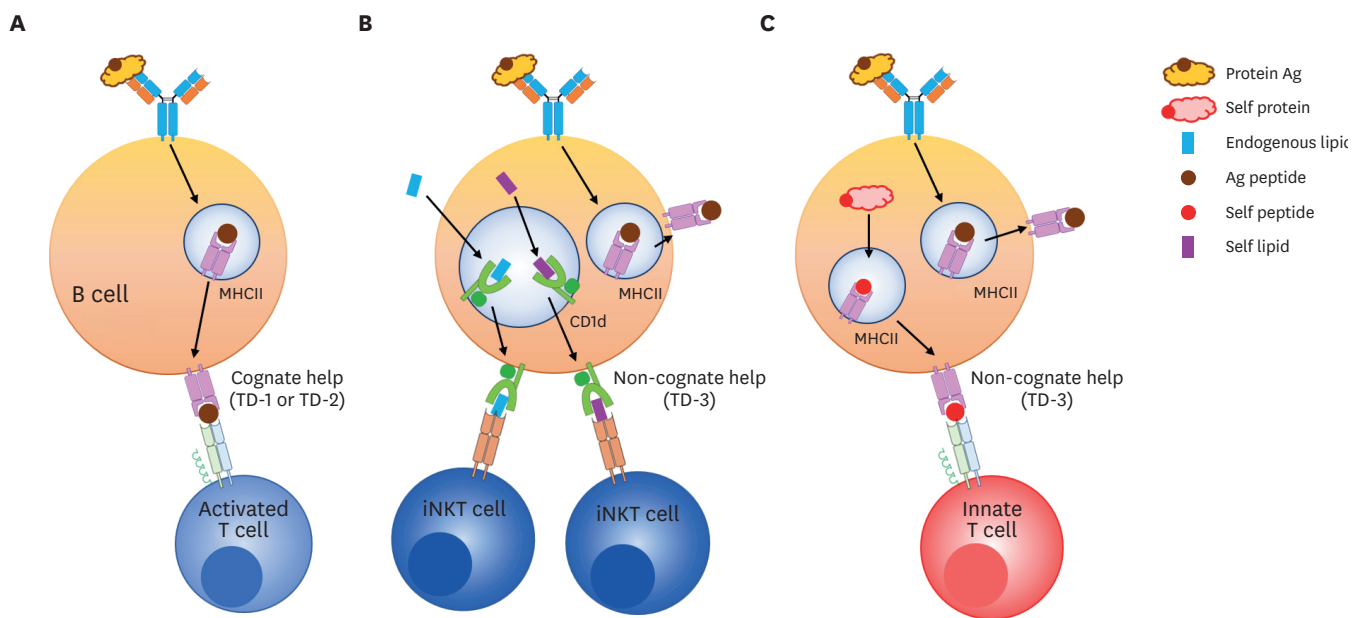


Figure 2. Cognate and non-cognate B cell help by MHC class II- or CD1d-restricted T cells. (A) B cell specific to a protein or lipid (not depicted) Ag presents the antigenic peptide within MHC class II or lipid moiety within CD1d (not depicted) and is helped by activated T cell or NKT cell, respectively (TD-1 or TD-2 responses). (B) Even though a B cell specific to a protein Ag presents a peptide from the protein Ag within MHC class II, the B cell can solicit help from a iNKT cell via the interaction between exogenous or self-lipid Ag within CD1d and TCR (TD-3 response). This help is non-cognate with respect to BCR. (C) The non-cognate B cell help can be supported by a bystander MHC class II-restricted helper T cell, which represents another TD-3 response. This bystander helper T cell can be activated T cell specific to other peptides or innate-like T cell that acquire a memory phenotype during development. The source of peptides for bystander helper T cells may be endogenous or derived from endocytosed proteins.

Table 1. The classification of TD Ab responses

Type of Ab response	T-B interaction	Cognate interaction	Ag presenting molecule	GC reaction	Affinity maturation
TD-1	+	+	MHC class II	+	+++
TD-2	+	+	MHC class II, CD1d	–	+
TD-3	+	–	MHC class II, CD1d	–	+

is whether BCR delivers the antigenic peptide or non-peptide Ag for the cognate interaction between TCR and MHC class II, CD1d, or other Ag-presenting molecules (**Fig. 2**). Therefore, critical points distinguishing TD-1, TD-2, and TD-3 responses are whether the response leads to the GC reaction (TD-1) or extrafollicular response (TD-2 and TD-3) and whether B cell presents peptide or lipid derived from the exact BCR-bound Ag to solicit T cell help (TD-1 and TD-2) or B cell is helped by bystander T cell (TD-3). The TD-3 response is less likely to occur between resting B and T cells, but occurs between activated B and T cells upon infection or other pathogenic stimuli. The non-cognate or bystander interaction between T and B cells is important for the overall Ab responses as previously reported (57,66–68). Another example of non-cognate B cell help via MHC class II is when CD4⁺ T cell recognizes idiotypic peptides from immunoglobulin heavy chain variable domain present within the MHC class II molecule (64). If NKT cell help to B cells specific for protein Ags occurs via CD1d, this CD1d-mediated interaction is non-cognate with respect to BCR specificity, and this Ab response should belong to a TD-3 response. The B cell help by $\gamma\delta$ T cells is regarded as a non-cognate TD-3 response, but appears to be exceptional in that this response can produce autoreactive GCs (47,51). It is interesting to address whether innate-like serosal CD49d^{high}CD4⁺ T cells provide B-1a cells in a non-cognate and MHC class II-dependent manner.

CONCLUDING REMARK

In vivo B cell Ab responses are extremely complex as they involve combinations of different types of TD and TI Ab responses. In addition to the classical cognate T–B interaction, the presence of non-cognate T–B interaction and extrafollicular Ab responses blurred the clear distinction between the TD and TI Ab response. Therefore, we suggested a new classification of the TD Ab responses to incorporate gray zone TD responses in this review. Especially, the innate-like T cells are thought to be important in boosting the overall Ab responses by providing cognate and non-cognate B cell help. The importance and types of innate-like helper T cells in all types of Ab responses needs to be investigated in the future.

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REFERENCES

1. Baumgarth N. A two-phase model of B-cell activation. *Immunol Rev* 2000;176:171-180.
[PUBMED](#) | [CROSSREF](#)
2. Lopes-Carvalho T, Foote J, Kearney JF. Marginal zone B cells in lymphocyte activation and regulation. *Curr Opin Immunol* 2005;17:244-250.
[PUBMED](#) | [CROSSREF](#)
3. Haas KM. B-1 lymphocytes in mice and nonhuman primates. *Ann N Y Acad Sci* 2015;1362:98-109.
[PUBMED](#) | [CROSSREF](#)
4. Vinuesa CG, Linterman MA, Yu D, MacLennan IC. Follicular helper T cells. *Annu Rev Immunol* 2016;34:335-368.
[PUBMED](#) | [CROSSREF](#)
5. Mond JJ, Lees A, Snapper CM. T cell-independent antigens type 2. *Annu Rev Immunol* 1995;13:655-692.
[PUBMED](#) | [CROSSREF](#)
6. Dellabona P, Casorati G. An unexpected requirement for CD4⁺ T cells in anti-glycolipid antibody responses. *Immunol Cell Biol* 2011;89:499-501.
[PUBMED](#) | [CROSSREF](#)
7. Vermijlen D, Prinz I. Ontogeny of innate T lymphocytes - some innate lymphocytes are more innate than others. *Front Immunol* 2014;5:486.
[PUBMED](#) | [CROSSREF](#)
8. Brigl M, Bry L, Kent SC, Gumperz JE, Brenner MB. Mechanism of CD1d-restricted natural killer T cell activation during microbial infection. *Nat Immunol* 2003;4:1230-1237.
[PUBMED](#) | [CROSSREF](#)
9. Jameson J, Witherden D, Havran WL. T-cell effector mechanisms: gammadelta and CD1d-restricted subsets. *Curr Opin Immunol* 2003;15:349-353.
[PUBMED](#) | [CROSSREF](#)
10. Martin F, Oliver AM, Kearney JF. Marginal zone and B1 B cells unite in the early response against T-independent blood-borne particulate antigens. *Immunity* 2001;14:617-629.
[PUBMED](#) | [CROSSREF](#)
11. Hardy RR. B-1 B cells: development, selection, natural autoantibody and leukemia. *Curr Opin Immunol* 2006;18:547-555.
[PUBMED](#) | [CROSSREF](#)
12. Holodick NE, Tumang JR, Rothstein TL. Continual signaling is responsible for constitutive ERK phosphorylation in B-1a cells. *Mol Immunol* 2009;46:3029-3036.
[PUBMED](#) | [CROSSREF](#)

13. Morris DL, Rothstein TL. Abnormal transcription factor induction through the surface immunoglobulin M receptor of B-1 lymphocytes. *J Exp Med* 1993;177:857-861.
[PUBMED](#) | [CROSSREF](#)
14. Moon H, Lee JG, Shin SH, Kim TJ. LPS-induced migration of peritoneal B-1 cells is associated with upregulation of CXCR4 and increased migratory sensitivity to CXCL12. *J Korean Med Sci* 2012;27:27-35.
[PUBMED](#) | [CROSSREF](#)
15. Rauch PJ, Chudnovskiy A, Robbins CS, Weber GF, Etzrodt M, Hilgendorf I, Tiglaio E, Figueiredo JL, Iwamoto Y, Theurl I, et al. Innate response activator B cells protect against microbial sepsis. *Science* 2012;335:597-601.
[PUBMED](#) | [CROSSREF](#)
16. Weber GF, Chousterman BG, Hilgendorf I, Robbins CS, Theurl I, Gerhard LM, Iwamoto Y, Quach TD, Ali M, Chen JW, et al. Pleural innate response activator B cells protect against pneumonia via a GM-CSF-IgM axis. *J Exp Med* 2014;211:1243-1256.
[PUBMED](#) | [CROSSREF](#)
17. Waffarn EE, Hastey CJ, Dixit N, Soo Choi Y, Cherry S, Kalinke U, Simon SI, Baumgarth N. Infection-induced type I interferons activate CD11b on B-1 cells for subsequent lymph node accumulation. *Nat Commun* 2015;6:8991.
[PUBMED](#) | [CROSSREF](#)
18. Savage HP, Yenson VM, Sawhney SS, Mousseau BJ, Lund FE, Baumgarth N. Blimp-1-dependent and -independent natural antibody production by B-1 and B-1-derived plasma cells. *J Exp Med* 2017;214:2777-2794.
[PUBMED](#) | [CROSSREF](#)
19. Baumgarth N, Waffarn EE, Nguyen TT. Natural and induced B-1 cell immunity to infections raises questions of nature versus nurture. *Ann N Y Acad Sci* 2015;1362:188-199.
[PUBMED](#) | [CROSSREF](#)
20. Park C, Kho IS, In Yang J, Kim MJ, Park S, Cha HS, Lee J, Kim TJ. Positive selection of type II collagen-reactive CD80^{high} marginal zone B cells in DBA/1 mice. *Clin Immunol* 2017;178:64-73.
[PUBMED](#) | [CROSSREF](#)
21. Pillai S, Cariappa A. The follicular versus marginal zone B lymphocyte cell fate decision. *Nat Rev Immunol* 2009;9:767-777.
[PUBMED](#) | [CROSSREF](#)
22. Srivastava B, Quinn WJ 3rd, Hazard K, Erikson J, Allman D. Characterization of marginal zone B cell precursors. *J Exp Med* 2005;202:1225-1234.
[PUBMED](#) | [CROSSREF](#)
23. Kerfoot SM, Yaari G, Patel JR, Johnson KL, Gonzalez DG, Kleinstein SH, Haberman AM. Germinal center B cell and T follicular helper cell development initiates in the interfollicular zone. *Immunity* 2011;34:947-960.
[PUBMED](#) | [CROSSREF](#)
24. Cannons JL, Qi H, Lu KT, Dutta M, Gomez-Rodriguez J, Cheng J, Wakeland EK, Germain RN, Schwartzberg PL. Optimal germinal center responses require a multistage T cell: B cell adhesion process involving integrins, SLAM-associated protein, and CD84. *Immunity* 2010;32:253-265.
[PUBMED](#) | [CROSSREF](#)
25. Odegard JM, Marks BR, DiPlacido LD, Poholek AC, Kono DH, Dong C, Flavell RA, Craft J. ICOS-dependent extrafollicular helper T cells elicit IgG production via IL-21 in systemic autoimmunity. *J Exp Med* 2008;205:2873-2886.
[PUBMED](#) | [CROSSREF](#)
26. Miyauchi K, Sugimoto-Ishige A, Harada Y, Adachi Y, Usami Y, Kaji T, Inoue K, Hasegawa H, Watanabe T, Hijikata A, et al. Protective neutralizing influenza antibody response in the absence of T follicular helper cells. *Nat Immunol* 2016;17:1447-1458.
[PUBMED](#) | [CROSSREF](#)
27. MacLennan IC, Toellner KM, Cunningham AF, Serre K, Sze DM, Zúñiga E, Cook MC, Vinuesa CG. Extrafollicular antibody responses. *Immunol Rev* 2003;194:8-18.
[PUBMED](#) | [CROSSREF](#)
28. Vos Q, Lees A, Wu ZQ, Snapper CM, Mond JJ. B-cell activation by T-cell-independent type 2 antigens as an integral part of the humoral immune response to pathogenic microorganisms. *Immunol Rev* 2000;176:154-170.
[PUBMED](#) | [CROSSREF](#)
29. Mongini PK, Paul WE, Metcalf ES. T cell regulation of immunoglobulin class expression in the antibody response to trinitrophenyl-ficoll. Evidence for T cell enhancement of the immunoglobulin class switch. *J Exp Med* 1982;155:884-902.
[PUBMED](#) | [CROSSREF](#)

30. Xu Z, Pone EJ, Al-Qahtani A, Park SR, Zan H, Casali P. Regulation of *aicda* expression and AID activity: relevance to somatic hypermutation and class switch DNA recombination. *Crit Rev Immunol* 2007;27:367-397.
[PUBMED](#) | [CROSSREF](#)
31. Yamamoto N, Kerfoot SM, Hutchinson AT, Dela Cruz CS, Nakazawa N, Szczepanik M, Majewska-Szczepanik M, Nazimek K, Ohana N, Bryniarski K, et al. Expression of activation-induced cytidine deaminase enhances the clearance of pneumococcal pneumonia: evidence of a subpopulation of protective anti-pneumococcal B1a cells. *Immunology* 2016;147:97-113.
[PUBMED](#) | [CROSSREF](#)
32. Kaku H, Holodick NE, Tumang JR, Rothstein TL. CD25+ B-1a cells express *Aicda*. *Front Immunol* 2017;8:672.
[PUBMED](#) | [CROSSREF](#)
33. Vinuesa CG, Chang PP. Innate B cell helpers reveal novel types of antibody responses. *Nat Immunol* 2013;14:119-126.
[PUBMED](#) | [CROSSREF](#)
34. Ochsenbein AF, Fehr T, Lutz C, Suter M, Brombacher F, Hengartner H, Zinkernagel RM. Control of early viral and bacterial distribution and disease by natural antibodies. *Science* 1999;286:2156-2159.
[PUBMED](#) | [CROSSREF](#)
35. Ouchida R, Lu Q, Liu J, Li Y, Chu Y, Tsubata T, Wang JY. FcγR interacts and cooperates with the B cell receptor to promote B cell survival. *J Immunol* 2015;194:3096-3101.
[PUBMED](#) | [CROSSREF](#)
36. Gaya M, Barral P, Burbage M, Aggarwal S, Montaner B, Warren Navia A, Aid M, Tsui C, Maldonado P, Nair U, Ghneim K, et al. Initiation of antiviral B cell immunity relies on innate signals from spatially positioned NKT cells. *Cell* 2018;172:517-533.e20.
37. Doherty DG, Melo AM, Moreno-Olivera A, Solomos AC. Activation and regulation of B cell responses by invariant natural killer T cells. *Front Immunol* 2018;9:1360.
[PUBMED](#) | [CROSSREF](#)
38. Bialecki E, Paget C, Fontaine J, Capron M, Trottein F, Faveeuw C. Role of marginal zone B lymphocytes in invariant NKT cell activation. *J Immunol* 2009;182:6105-6113.
[PUBMED](#) | [CROSSREF](#)
39. Barral P, Eckl-Dorna J, Harwood NE, De Santo C, Salio M, Illarionov P, Besra GS, Cerundolo V, Batista FD. B cell receptor-mediated uptake of CD1d-restricted antigen augments antibody responses by recruiting invariant NKT cell help *in vivo*. *Proc Natl Acad Sci U S A* 2008;105:8345-8350.
[PUBMED](#) | [CROSSREF](#)
40. Galli G, Nuti S, Tavarini S, Galli-Stampino L, De Lalla C, Casorati G, Dellabona P, Abrignani S. CD1d-restricted help to B cells by human invariant natural killer T lymphocytes. *J Exp Med* 2003;197:1051-1057.
[PUBMED](#) | [CROSSREF](#)
41. Chang PP, Barral P, Fitch J, Pratama A, Ma CS, Kallies A, Hogan JJ, Cerundolo V, Tangye SG, Bittman R, et al. Identification of Bcl-6-dependent follicular helper NKT cells that provide cognate help for B cell responses. *Nat Immunol* 2011;13:35-43.
[PUBMED](#) | [CROSSREF](#)
42. Chaudhry MS, Karadimitris A. Role and regulation of CD1d in normal and pathological B cells. *J Immunol* 2014;193:4761-4768.
[PUBMED](#) | [CROSSREF](#)
43. Van Kaer L, Parekh VV, Wu L. The response of CD1d-restricted invariant NKT cells to microbial pathogens and their products. *Front Immunol* 2015;6:226.
[PUBMED](#) | [CROSSREF](#)
44. Kwon DI, Lee YJ. Lineage differentiation program of invariant natural killer T cells. *Immune Netw* 2017;17:365-377.
[PUBMED](#) | [CROSSREF](#)
45. Wen L, Pao W, Wong FS, Peng Q, Craft J, Zheng B, Kelsoe G, Dianda L, Owen MJ, Hayday AC. Germinal center formation, immunoglobulin class switching, and autoantibody production driven by “non alpha/beta” T cells. *J Exp Med* 1996;183:2271-2282.
[PUBMED](#) | [CROSSREF](#)
46. Mizoguchi A, Mizoguchi E, de Jong YP, Takedatsu H, Preffer FI, Terhorst C, Bhan AK. Role of the CD5 molecule on TCR gamma delta T cell-mediated immune functions: development of germinal centers and chronic intestinal inflammation. *Int Immunol* 2003;15:97-108.
[PUBMED](#) | [CROSSREF](#)
47. Dianda L, Gulbranson-Judge A, Pao W, Hayday AC, MacLennan IC, Owen MJ. Germinal center formation in mice lacking alpha beta T cells. *Eur J Immunol* 1996;26:1603-1607.
[PUBMED](#) | [CROSSREF](#)

48. Vavassori S, Kumar A, Wan GS, Ramanjaneyulu GS, Cavallari M, El Daker S, Beddoe T, Theodossis A, Williams NK, Gostick E, et al. Butyrophilin 3A1 binds phosphorylated antigens and stimulates human $\gamma\delta$ T cells. *Nat Immunol* 2013;14:908-916.
[PUBMED](#) | [CROSSREF](#)
49. Tyler CJ, Doherty DG, Moser B, Eberl M. Human V γ 9/V δ 2 T cells: innate adaptors of the immune system. *Cell Immunol* 2015;296:10-21.
[PUBMED](#) | [CROSSREF](#)
50. Miller C, Stedra J, Kelsoe G, Cerny J. Facultative role of germinal centers and T cells in the somatic diversification of IgVH genes. *J Exp Med* 1995;181:1319-1331.
[PUBMED](#) | [CROSSREF](#)
51. Pao W, Wen L, Smith AL, Gulbranson-Judge A, Zheng B, Kelsoe G, MacLennan IC, Owen MJ, Hayday AC. Gamma delta T cell help of B cells is induced by repeated parasitic infection, in the absence of other T cells. *Curr Biol* 1996;6:1317-1325.
[PUBMED](#) | [CROSSREF](#)
52. Domeier PP, Chodisetti SB, Soni C, Schell SL, Elias MJ, Wong EB, Cooper TK, Kitamura D, Rahman ZS. IFN- γ receptor and STAT1 signaling in B cells are central to spontaneous germinal center formation and autoimmunity. *J Exp Med* 2016;213:715-732.
[PUBMED](#) | [CROSSREF](#)
53. Moon BG, Takaki S, Miyake K, Takatsu K. The role of IL-5 for mature B-1 cells in homeostatic proliferation, cell survival, and Ig production. *J Immunol* 2004;172:6020-6029.
[PUBMED](#) | [CROSSREF](#)
54. Irei T, Ohdan H, Zhou W, Ishiyama K, Tanaka Y, Ide K, Asahara T. The persistent elimination of B cells responding to blood group A carbohydrates by synthetic group A carbohydrates and B-1 cell differentiation blockade: novel concept in preventing antibody-mediated rejection in ABO-incompatible transplantation. *Blood* 2007;110:4567-4575.
[PUBMED](#) | [CROSSREF](#)
55. Cunningham AF, Flores-Langarica A, Bobat S, Dominguez Medina CC, Cook CN, Ross EA, Lopez-Macias C, Henderson IR. B1b cells recognize protective antigens after natural infection and vaccination. *Front Immunol* 2014;5:535.
[PUBMED](#) | [CROSSREF](#)
56. Alugupalli KR, Leong JM, Woodland RT, Muramatsu M, Honjo T, Gerstein RM. B1b lymphocytes confer T cell-independent long-lasting immunity. *Immunity* 2004;21:379-390.
[PUBMED](#) | [CROSSREF](#)
57. Margry B, Wieland WH, van Kooten PJ, van Eden W, Broere F. Peritoneal cavity B-1a cells promote peripheral CD4 $^{+}$ T-cell activation. *Eur J Immunol* 2013;43:2317-2326.
[PUBMED](#) | [CROSSREF](#)
58. Erickson LD, Foy TM, Waldschmidt TJ. Murine B1 B cells require IL-5 for optimal T cell-dependent activation. *J Immunol* 2001;166:1531-1539.
[PUBMED](#) | [CROSSREF](#)
59. Tazawa H, Irei T, Tanaka Y, Igarashi Y, Tashiro H, Ohdan H. Blockade of invariant TCR-CD1d interaction specifically inhibits antibody production against blood group A carbohydrates. *Blood* 2013;122:2582-2590.
[PUBMED](#) | [CROSSREF](#)
60. Christiansen D, Vaughan HA, Milland J, Dodge N, Mouhtouris E, Smyth MJ, Godfrey DI, Sandrin MS. Antibody responses to glycolipid-borne carbohydrates require CD4 $^{+}$ T cells but not CD1 or NKT cells. *Immunol Cell Biol* 2011;89:502-510.
[PUBMED](#) | [CROSSREF](#)
61. Moon H, Park C, Lee JG, Shin SH, Lee JH, Kho I, Kang K, Cha HS, Kim TJ. Early development in the peritoneal cavity of CD49d high Th1 memory phenotype CD4 $^{+}$ T cells with enhanced B cell helper activity. *J Immunol* 2015;195:564-575.
[PUBMED](#) | [CROSSREF](#)
62. Yang JI, Park C, Kho I, Lee S, Suh KS, Kim TJ. Serosal cavities contain two populations of innate-like integrin $\alpha 4^{high}$ CD4 $^{+}$ T cells, integrin $\alpha 4\beta 1^{+}\alpha 6\beta 1^{+}\alpha 4\beta 7$ and $\alpha 4\beta 1^{+}\alpha 6\beta 1\alpha 4\beta 7^{+}$ cells. *Immune Netw* 2017;17:392-401.
[PUBMED](#) | [CROSSREF](#)
63. Lee JG, Jang JY, Fang T, Xu Y, Yan JJ, Ryu JH, Jeon HJ, Koo TY, Kim DK, Oh KH, et al. Identification of human B-1 helper T cells with a Th1-like memory phenotype and high integrin CD49d expression. *Front Immunol* 2018;9:1617.
[PUBMED](#) | [CROSSREF](#)

64. Jacobsen JT, Lunde E, Sundvold-Gjerstad V, Munthe LA, Bogen B. The cellular mechanism by which complementary Id+ and anti-Id antibodies communicate: T cells integrated into idiotypic regulation. *Immunol Cell Biol* 2010;88:515-522.
[PUBMED](#) | [CROSSREF](#)
65. Ryan SO, Bonomo JA, Zhao F, Cobb BA. MHCII glycosylation modulates *Bacteroides fragilis* carbohydrate antigen presentation. *J Exp Med* 2011;208:1041-1053.
[PUBMED](#) | [CROSSREF](#)
66. Tangye SG, Brink R, Goodnow CC, et al. SnapShot: interactions between B cells and T cells. *Cell* 2015;162:926-966.e1.
67. Boyman O. Bystander activation of CD4+ T cells. *Eur J Immunol* 2010;40:936-939.
[PUBMED](#) | [CROSSREF](#)
68. Silver J, Zuo T, Chaudhary N, Kumari R, Tong P, Giguere S, Granato A, Donthula R, Devereaux C, Wesemann DR. Stochasticity enables BCR-independent germinal center initiation and antibody affinity maturation. *J Exp Med* 2018;215:77-90.
[PUBMED](#) | [CROSSREF](#)