

Microbiome of Hepatobiliary Diseases

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The microbiome, which has been defined as ‘the ecological community of commensal, symbiotic and pathogenic microorganisms that share our body space, may be distinguished from the microbiota as it includes the collective genomes. An increasing level of evidence reveals that the human microbiome plays a major role in health. For this reason, it is often referred to as the ‘forgotten organ.’ All surfaces of the human body that are exposed to the environment are colonized, including skin, respiratory system, urogenital tract and gastrointestinal (GI) tract, totaling at least 100 trillion microbial cells. The known roles of the GI microbiome include metabolic functions, synthesis functions, and immune roles. Recent studies indicate that the human gut microbiome plays a significant role in health and disease. Dysbiosis, defined as a pathological imbalance in a microbial community, is becoming increasingly appreciated as a ‘central environmental factor’ that is both associated with complex phenotypes and affected by host genetics, diet, and antibiotic use. More recently, a link has been established between the dysmetabolism of bile acids (BAs) in the gut and the gut-liver axis, and this relationship with the microbiome has been highlighted. This review summarizes the microbiome of the hepatobiliary system and how microbiome is related to diseases of the liver and biliary tract.

Key words: Microbiome; Liver; Gallbladder; Pancreas; Cancer

INTRODUCTION

Hepatobiliary and pancreatic cancers are associated with poor prognosis owing to their high level of tumor invasiveness, recurrence, hematogenous and lymphatic metastasis, resistance to first-line chemotherapy, and lack of effective target therapy [1,2]. Evidence in the literature suggests that hepatobiliary and pancreatic cancers develop through the accumulation of genetic and epigenetic alterations, which is influenced by host immune state, food, and environmental and microbial exposures [1-4].

The human microbiota is the collection of microorganisms exists in the human being, and the relationships with microorganisms and host can be considered to maintain a wide range of the spectrum, from mutualism to pathogen [5]. Abrupt changes in the microbiota of various human body areas associate with diverse localized or

systemic human diseases. The human gastrointestinal tract is one of the biggest storing spaces of microbes in the body and contains both commensal and pathogenic microbial species [6]. Research on intestinal microbiota has shown that inflammatory bowel disease is originated from the varied composition of microbial composition and abnormal and overflowing mucosal immune response [7]. Numerous pathogens can promote cancer through well-identified mechanisms [8]. Although most studies are confined to specific bacterial pathogens and viruses, the link between human cancer and bacterial microbiota has recently been studied actively by using next-generation sequencing technology for microbiome profiling [9]. There is an increasing interest in understanding the role of microbiome as a microenvironment for cancer development, particularly in the area of hepatobiliary and pancreatic cancers [10].

The liver, biliary tract, and pancreas are located in very close

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proximity, and these three structures link up with the gastrointestinal tract. Therefore, the gut microbiome can easily reach the liver through the portal vein [11]. The incidence of hepatobiliary cancer is higher in east and southeast Asian countries, such as Japan, Korea, and Thailand [4]. The incidence rate of cholangiocarcinoma in South Korea correlates with the prevalence of liver fluke (*Opisthorchi viverrini*) infection in the region [12].

In the rest of this review, we will describe the role of the microbiome in the hepatobiliary and pancreatic diseases, including non-alcoholic fatty liver disease (NAFLD), alcoholic liver disease, liver cirrhosis, hepatocellular carcinoma, and gallbladder cancer.

1. Microbiome of liver

The relationship between the gut and the liver is well understood [13]. The most prevalent type of hepatic disorder is NAFLD, and over 60 million Americans suffer from it [14]. The present understanding of the etiology of the spectrum of liver diseases is explained by proinflammatory changes in the host. Intestinal dysbiosis (anomalous or imbalanced gut microbial composition) and increased intestinal permeability lead to translocation of microorganisms and microbial products, including cell wall components and DNA, together referred to as microbial-associated molecular patterns (MAMPs) or pathogen-associated molecular patterns (PAMPs). These changes cause a basic spectrum of hepatic diseases with various bacterial species.

2. Microbiome of specific liver diseases

1) Nonalcoholic fatty liver disease (NAFLD)

NAFLD can be defined as a spectrum of liver diseases that can be generally classified into two categories: nonalcoholic fatty liver, the simple form of NAFLD, and nonalcoholic steatohepatitis (NASH), the aggressive form of NAFLD [15]. NASH is typically related to type 2 diabetes mellitus, heart and vascular risk factors, and obesity [16,17]. However, NAFLD has also been commonly found in non-obese patients, supporting that genetic parameters also contribute to disease development [18-21].

Some studies have focused on the effect of the gut microbiota in NAFLD, but the cause-effect relationship has not been verified [22]. Individuals with NAFLD have a higher incidence of microbial dysbiosis [23]. Using 16S amplicon sequencing, the bacterial genera *Bacteroides* and *Ruminococcus* were profoundly increased, and *Prevotella* was decreased in persons with NASH compared with those without NASH [23]. Whole genome metagenomics in patients with NAFLD showed an increased prevalence of *Escherichia coli* and *Bacteroides vulgatus* in patients with advanced fibrosis [24].

2) Alcoholic liver disease (ALD)

Similar to NAFLD, the benign form of ALD is characterized by the accumulation of fat inside the liver (fatty liver or steatosis), whereas the progressive form is marked by inflammation and liver injury (alcoholic steatohepatitis (ASH)).

Our knowledge of contributions of the gut microbiota in ALD is increased. As in NAFLD, SIBO has been demonstrated as an important hallmark of alcohol-associated liver disease in humans [25] and mouse models [26,27]. Intestinal dysbiosis in individuals who abuse alcohol is characterized by marked enrichment of Enterobacteriaceae (family) and reduction in abundance of Bacteroidetes and *Lactobacillus* [26,28-30]. It has also been demonstrated that alcohol-induced dysbiosis is only partially reversible by alcohol withdrawal or probiotic supplement [31,32]. Interestingly, patients dependent on alcohol also displayed reduced fungal diversity and *Candida* overgrowth, presenting the first evidence of the role of the gut mycobiome in the pathogenesis of liver diseases [33].

3) Cirrhosis

Alterations in the gut microbiota, including dysbiosis and SIBO, have been associated with cirrhosis and its complications [34-37]. Gut microbiome alterations were observed in patients with alcohol-associated and hepatitis-associated cirrhosis in a Chinese cohort [38], with an invasion of the lower intestinal tract by microorganisms associated with the oral cavity, such as *Veillonella* and *Streptococcus*. Concordant with these findings, Chen and colleagues also found an over-representation of genera, including *Veillonella*, *Megasphaera*, *Dialister*, *Atopobium* and *Prevotella* in the duodenum of patients with cirrhosis. The genera *Neisseria* and *Gemella* were discriminative between HBV-related and PBC-related cirrhosis [37]. In 2017, Bajaj and colleagues observed a significantly high incidence of fungal dysbiosis in patients with cirrhosis and showed that the Bacteroidetes: *Ascomycota* ratio could independently predict hospitalization in these patients [39].

4) Hepatocellular carcinoma (HCC)

The etiology of HCC follows a so-called multiple step pathway, whereby liver steatosis, followed by oxidative and ER stress, together with intestinal dysbiosis and inflammation, contributes to the final cause of cancer. The gut microbiota definitely changes in composition in human bodies with HCC. Clostridium species have been found to be enriched in obesity-induced mouse models of HCC [40,41], but clinical studies of patients with HCC detected an overgrowth of intestinal *Escherichia coli* [42]. Mouse models and human studies have reported migration of *Helicobacter* species into HCC

tumor tissues [43-46].

3. Microbiome and gallbladder cancer

Gallbladder cancer (GBC) is a relatively uncommon primary malignancy. However, it is the most common malignant neoplasm of the biliary system and a lethal cancer with fatal outcome [47-49]. Globally, GBC rates exhibit marked regional variability, reaching epidemic levels for some regions and ethnicities, especially in countries in Asia such as India, Korea, Japan, and in South America [50]. The basis for this difference likely related to differences in environmental exposures interacting with genetic factors. The previous epidemiologic studies have revealed several risk factors associated with GBC, including gallstones, chronic cholecystitis, and infection, especially *Salmonella* [51].

Although the role of microbiota in gallbladder carcinogenesis is still not well known, the previous epidemiologic studies have revealed that the risk of GBC increases with chronic infection by *Salmonella* species [52]. A meta-analysis from 11 different epidemiologic studies revealed that overall odd ratio of GBC in chronic *Salmonella typhi* carrier patients is over 4. Importantly, recent experimental studies revealed morphologic evidence and molecular mechanism for *Salmonella*-induced GB cancer or premalignant lesion [53,54]. *Salmonella* infection of gallbladder organoids induces loss of polarity, nuclear atypia with prominent nucleoli, and discohesiveness with loss of epithelial marker, E-cadherin [53]. This malignant transformation is also observed in mouse embryonic fibroblasts, and Akt and MAP kinase pathways, which are well-known cancer pathways, are activated during *Salmonella* infection [53]. Chronic cholecystitis is commonly observed in the gallbladder of mice with gallstones. However, atypical hyperplasia that is a premalignant condition, is only associated with chronic *Salmonella* infection regardless of the presence of gallstones [54]. The biologic effect of *Helicobacter bilis* has been investigated on a cell line of human bile duct cancer and showed activation of transcript factors, such as NF- κ B, E2F and CRE that stimulate the production of VEGF and lead to enhancement of angiogenesis [55]. These epidemiologic and experimental studies support the role of infection on GBC carcinogenesis.

CONCLUSION AND PERSPECTIVES

Various research activities for microbiome suggests that unknown pathophysiology of diseases of the hepatobiliary system can be solved and explained by the microbiome research in some part. The field is slowly moving from observation to real clinical practice. Also, cutting-edge techniques in this field will widen basic under-

standing of hepatobiliary diseases and hopefully improve the cure rate of these fatal diseases in the near future.

REFERENCES

1. Bruix, J., G.J. Gores, and V. Mazzaferro, Hepatocellular carcinoma: clinical frontiers and perspectives. *Gut*, 2014. 63(5): p. 844-55.
2. Ho, J., et al., Translational genomics in pancreatic ductal adenocarcinoma: A review with re-analysis of TCGA dataset. *Semin Cancer Biol*, 2018.
3. Salem, A.A. and G.G. Mackenzie, Pancreatic cancer: A critical review of dietary risk. *Nutr Res*, 2018. 52: p. 1-13.
4. Shibata, T., Y. Arai, and Y. Totoki, Molecular genomic landscapes of hepatobiliary cancer. *Cancer Sci*, 2018. 109(5): p. 1282-1291.
5. Hooper, L.V. and J.I. Gordon, Commensal host-bacterial relationships in the gut. *Science*, 2001. 292(5519): p. 1115-8.
6. Kostic, A.D., R.J. Xavier, and D. Gevers, The microbiome in inflammatory bowel disease: current status and the future ahead. *Gastroenterology*, 2014. 146(6): p. 1489-99.
7. Hollister, E.B., C. Gao, and J. Versalovic, Compositional and functional features of the gastrointestinal microbiome and their effects on human health. *Gastroenterology*, 2014. 146(6): p. 1449-58.
8. Moore, P.S. and Y. Chang, Why do viruses cause cancer? Highlights of the first century of human tumour virology. *Nat Rev Cancer*, 2010. 10(12): p. 878-89.
9. Schwabe, R.F. and C. Jobin, The microbiome and cancer. *Nat Rev Cancer*, 2013. 13(11): p. 800-12.
10. Abreu, M.T. and R.M. Peek, Jr., Gastrointestinal malignancy and the microbiome. *Gastroenterology*, 2014. 146(6): p. 1534-1546 e3.
11. Szabo, G., Gut-liver axis in alcoholic liver disease. *Gastroenterology*, 2015. 148(1): p. 30-6.
12. Kamsa-ard, S., et al., Risk Factors for Cholangiocarcinoma in Thailand: A Systematic Review and Meta-Analysis. *Asian Pac J Cancer Prev*, 2018. 19(3): p. 605-614.
13. Schnabl, B. and D.A. Brenner, Interactions between the intestinal microbiome and liver diseases. *Gastroenterology*, 2014. 146(6): p. 1513-24.
14. Younossi, Z.M., et al., The economic and clinical burden of nonalcoholic fatty liver disease in the United States and Europe. *Hepatology*, 2016. 64(5): p. 1577-1586.
15. Spengler, E.K. and R. Loomba, Recommendations for Diagnosis, Referral for Liver Biopsy, and Treatment of Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. *Mayo Clin Proc*, 2015. 90(9): p. 1233-46.
16. Loomba, R., et al., Association between diabetes, family history

- of diabetes, and risk of nonalcoholic steatohepatitis and fibrosis. *Hepatology*, 2012. 56(3): p. 943-51.
17. Doycheva, I., et al., Non-invasive screening of diabetics in primary care for NAFLD and advanced fibrosis by MRI and MRE. *Aliment Pharmacol Ther*, 2016. 43(1): p. 83-95.
 18. Loomba, R., et al., Heritability of Hepatic Fibrosis and Steatosis Based on a Prospective Twin Study. *Gastroenterology*, 2015. 149(7): p. 1784-93.
 19. Cui, J., et al., Shared genetic effects between hepatic steatosis and fibrosis: A prospective twin study. *Hepatology*, 2016. 64(5): p. 1547-1558.
 20. Caussy, C., et al., Nonalcoholic fatty liver disease with cirrhosis increases familial risk for advanced fibrosis. *J Clin Invest*, 2017. 127(7): p. 2697-2704.
 21. Gao, B. and R. Bataller, Alcoholic liver disease: pathogenesis and new therapeutic targets. *Gastroenterology*, 2011. 141(5): p. 1572-85.
 22. Wieland, A., et al., Systematic review: microbial dysbiosis and non-alcoholic fatty liver disease. *Aliment Pharmacol Ther*, 2015. 42(9): p. 1051-63.
 23. Boursier, J., et al., The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology*, 2016. 63(3): p. 764-75.
 24. Loomba, R., et al., Gut Microbiome-Based Metagenomic Signature for Non-invasive Detection of Advanced Fibrosis in Human Non-alcoholic Fatty Liver Disease. *Cell Metab*, 2017. 25(5): p. 1054-1062 e5.
 25. Mouzaki, M., et al., Bile Acids and Dysbiosis in Non-Alcoholic Fatty Liver Disease. *PLoS One*, 2016. 11(5): p. e0151829.
 26. Yan, A.W., et al., Enteric dysbiosis associated with a mouse model of alcoholic liver disease. *Hepatology*, 2011. 53(1): p. 96-105.
 27. Ferrere, G., et al., Fecal microbiota manipulation prevents dysbiosis and alcohol-induced liver injury in mice. *J Hepatol*, 2017. 66(4): p. 806-815.
 28. Mutlu, E.A., et al., Colonic microbiome is altered in alcoholism. *Am J Physiol Gastrointest Liver Physiol*, 2012. 302(9): p. G966-78.
 29. Tuomisto, S., et al., Changes in gut bacterial populations and their translocation into liver and ascites in alcoholic liver cirrhotics. *BMC Gastroenterol*, 2014. 14: p. 40.
 30. Chen, Y., et al., Characterization of fecal microbial communities in patients with liver cirrhosis. *Hepatology*, 2011. 54(2): p. 562-72.
 31. Kirpich, I.A., et al., Probiotics restore bowel flora and improve liver enzymes in human alcohol-induced liver injury: a pilot study. *Alcohol*, 2008. 42(8): p. 675-82.
 32. Leclercq, S., et al., Intestinal permeability, gut-bacterial dysbiosis, and behavioral markers of alcohol-dependence severity. *Proc Natl Acad Sci U S A*, 2014. 111(42): p. E4485-93.
 33. Yang, A.M., et al., Intestinal fungi contribute to development of alcoholic liver disease. *J Clin Invest*, 2017. 127(7): p. 2829-2841.
 34. Bajaj, J.S., et al., Gut Microbiota Alterations can predict Hospitalizations in Cirrhosis Independent of Diabetes Mellitus. *Sci Rep*, 2015. 5: p. 18559.
 35. Jun, D.W., et al., Association between small intestinal bacterial overgrowth and peripheral bacterial DNA in cirrhotic patients. *Dig Dis Sci*, 2010. 55(5): p. 1465-71.
 36. Yao, J., et al., Nutrition status and small intestinal bacterial overgrowth in patients with virus-related cirrhosis. *Asia Pac J Clin Nutr*, 2016. 25(2): p. 283-91.
 37. Chen, Y., et al., Dysbiosis of small intestinal microbiota in liver cirrhosis and its association with etiology. *Sci Rep*, 2016. 6: p. 34055.
 38. Qin, N., et al., Alterations of the human gut microbiome in liver cirrhosis. *Nature*, 2014. 513(7516): p. 59-64.
 39. Bajaj, J.S., et al., Fungal dysbiosis in cirrhosis. *Gut*, 2018. 67(6): p. 1146-1154.
 40. Yoshimoto, S., et al., Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature*, 2013. 499(7456): p. 97-101.
 41. Xie, G., et al., Distinctly altered gut microbiota in the progression of liver disease. *Oncotarget*, 2016. 7(15): p. 19355-66.
 42. Grat, M., et al., Relevance of Pre-Transplant alpha-fetoprotein Dynamics in Liver Transplantation for Hepatocellular Cancer. *Ann Transplant*, 2016. 21: p. 115-24.
 43. Fox, J.G., et al., Gut microbes define liver cancer risk in mice exposed to chemical and viral transgenic hepatocarcinogens. *Gut*, 2010. 59(1): p. 88-97.
 44. Rogers, A.B., Distance burning: how gut microbes promote extraintestinal cancers. *Gut Microbes*, 2011. 2(1): p. 52-7.
 45. Huang, Y., et al., Identification of helicobacter species in human liver samples from patients with primary hepatocellular carcinoma. *J Clin Pathol*, 2004. 57(12): p. 1273-7.
 46. Kruttgen, A., et al., Study on the association of Helicobacter species with viral hepatitis-induced hepatocellular carcinoma. *Gut Microbes*, 2012. 3(3): p. 228-33.
 47. Ferlay, J., et al., Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, 2015. 136(5): p. E359-86.
 48. Siegel, R., et al., Cancer statistics, 2014. *CA Cancer J Clin*, 2014. 64(1): p. 9-29.
 49. Jung, K.W., et al., Cancer statistics in Korea: incidence, mortality, survival, and prevalence in 2011. *Cancer Res Treat*, 2014. 46(2): p.

- 109-23.
50. Randi, G., S. Franceschi, and C. La Vecchia, Gallbladder cancer worldwide: geographical distribution and risk factors. *Int J Cancer*, 2006. 118(7): p. 1591-602.
 51. Boutros, C., et al., Gallbladder cancer: past, present and an uncertain future. *Surg Oncol*, 2012. 21(4): p. e183-91.
 52. Nagaraja, V. and G.D. Eslick, Systematic review with meta-analysis: the relationship between chronic *Salmonella typhi* carrier status and gall-bladder cancer. *Aliment Pharmacol Ther*, 2014. 39(8): p. 745-50.
 53. Scanu, T., et al., *Salmonella* Manipulation of Host Signaling Pathways Provokes Cellular Transformation Associated with Gallbladder Carcinoma. *Cell Host Microbe*, 2015. 17(6): p. 763-74.
 54. Gonzalez-Escobedo, G., K.M. La Perle, and J.S. Gunn, Histopathological analysis of *Salmonella* chronic carriage in the mouse hepatopancreatobiliary system. *PLoS One*, 2013. 8(12): p. e84058.
 55. Takayama, S., et al., Effect of *Helicobacter bilis* infection on human bile duct cancer cells. *Dig Dis Sci*, 2010. 55(7): p. 1905-10.