

Review article

<http://dx.doi.org/10.6065/apem.2013.18.3.101>
Ann Pediatr Endocrinol Metab 2013;18:101-105

Genetic syndromes associated with overgrowth in childhood

Jung Min Ko, MD, PhD

Department of Pediatrics,
Seoul National University College of
Medicine, Seoul, Korea

Overgrowth syndromes comprise a diverse group of conditions with unique clinical, behavioral and molecular genetic features. While considerable overlap in presentation sometimes exists, advances in identification of the precise etiology of specific overgrowth disorders continue to improve clinicians' ability to make an accurate diagnosis. Among them, this paper introduces two classic genetic overgrowth syndromes: Sotos syndrome and Beckwith-Wiedemann syndrome. Historically, the diagnosis was based entirely on clinical findings. However, it is now understood that Sotos syndrome is caused by a variety of molecular genetic alterations resulting in haploinsufficiency of the *NSD1* gene at chromosome 5q35 and that Beckwith-Wiedemann syndrome is caused by heterogeneous abnormalities in the imprinting of a number of growth regulatory genes within chromosome 11p15 in the majority of cases. Interestingly, the 11p15 imprinting region is also associated with Russell-Silver syndrome which is a typical growth retardation syndrome. Opposite epigenetic alterations in 11p15 result in opposite clinical features shown in Beckwith-Wiedemann syndrome and Russell-Silver syndrome. Although the exact functions of the causing genes have not yet been completely understood, these overgrowth syndromes can be good models to clarify the complex basis of human growth and help to develop better-directed therapies in the future.

Keywords: Macrosomia, Sotos syndrome, Beckwith-Wiedemann syndrome, genomic imprinting

Introduction

Somatic growth is dependent on an increase in both cell size and number. Human growth comes from a complex interplay of various factors including genetic backgrounds and environmental influences. However, complex mechanisms involved in the regulation of somatic growth remains to be completely understood particularly in the molecular level of genetic field. Overgrowth refers to a condition characterized by extreme physical size and stature including tall stature or generalized/localized overgrowth of tissues¹⁾. The condition originates during infancy, childhood or adolescence while epiphyseal growth plates remain open. Among various conditions showing overgrowth, genetic overgrowth syndrome refers to a nonhormonally mediated overgrowth condition which can accompany increased height and/or head circumference, various degrees of mental retardation, or physical dysmorphisms in children¹⁾. It is clearly distinguished from pituitary gigantism which is caused by excess of growth hormone. However, there are overlaps in clinical and molecular features between overgrowth syndromes, thus making a specific diagnosis is often difficult.

This paper reviews clinical characteristics and molecular basis of typical genetic overgrowth syndromes, focusing on Sotos syndrome (OMIM#117550) and Beckwith-Wiedemann syndrome (OMIM#130650).

Received: 20 August, 2013
Accepted: 26 August, 2013

Address for correspondence:

Jung Min Ko, MD, PhD
Department of Pediatrics,
Seoul National University Children's
Hospital, Seoul National University
College of Medicine, 101 Daehak-ro,
Jongno-gu, Seoul 110-744, Korea
Tel: +82-2-2072-3570
Fax: +82-2-743-3455
E-mail: jmko@snu.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ISSN: 2287-1012(Print)
ISSN: 2287-1292(Online)

Sotos syndrome

1. Clinical characteristics

Sotos syndrome (SS, OMIM#117550), also known as cerebral gigantism, is a prenatal and postnatal overgrowth syndrome characterized by excessive growth resulting in tall stature and macrocephaly, distinctive craniofacial features, and developmental delay. These three cardinal features are each present in over 90% of cases with SS^{2,3}. Since the first description by Sotos et al.⁴ in 1964, hundreds of cases have been reported to date, and the estimated incidence is 1/15,000–1/20,000¹.

The typical overgrowth pattern of SS starts prenatally, resulting in higher mean birth length and weight⁵. Pronounced postnatal growth is obvious in the first 6 years of life, consistently displaying height above the 97th percentile⁶. However, the final adult height is usually within the upper normal range due to accompanied bone age advancement⁶.

A characteristic facial appearance consists of a high and broad forehead, sparse fronto-temporal hair, malar flushing, down-slanted palpebral fissures and a pointed chin⁷. The head circumference is increased above the 97th percentile in most SS patients, and it is thought to be the most consistent indicator of SS at any age⁵.

The majority of SS patients have some degree of developmental delay/learning disability. Achievement of developmental milestones such as walking and speech is commonly delayed. However, most patients have mild to moderate intellectual impairment, and the severity is very broad, ranging from intelligence quotient below 30 to above 100².

Besides, there are other commonly associated features including a history of neonatal jaundice and feeding difficulty, variable types of cardiac and renal anomalies, seizure, scoliosis, strabismus, attention deficit hyperactivity disorder, nonspecific

abnormal brain image findings such as ventriculomegaly and corpus callosum hypoplasia. Patients with overgrowth syndromes including SS have higher risks for the development of neoplasias, particularly in their childhood. In SS patients, the frequency of tumor development has been reported to be 2–7%^{8,9}, and the relative risk of certain malignancies including neural crest tumors, sacrococcygeal teratomas and some hematological malignancies is increased². However, routine screening of tumor development is not a standardized recommendation.

2. Molecular and genetic basis

In 2002, the nuclear receptor set domain containing protein 1 gene, *NSDI*, on chromosome 5q35 was identified as a causing gene of SS¹⁰. SS is caused by haploinsufficiency of *NSDI* in 60–90% of clinically diagnosed SS patients and can be transmitted in an autosomal dominant manner, although more than 95% of patients gain the disease from *de novo* mutation¹¹.

The *NSDI* gene consists of 23 exons and encodes multiple functional domains, including the SU(VAR)3-9, E(Z), tirthorax (SET), SET-associated domains, which mediate histone methyltransferase activity, five plant homeo-domains implicated in chromatin regulation, and two proline-tryptophan-tryptophan-proline domains that may mediate protein interactions³. *NSDI* is expressed in several tissues including the brain, kidney, skeletal muscle, spleen, and thymus¹². Although the exact role of the *NSDI* protein has not been identified, the presence of two different ligand binding domains suggests that *NSDI* enables the regulation of both negative and positive transcription¹³.

Several reports have demonstrated *NSDI* abnormalities in patients with Sotos syndrome. *NSDI* abnormalities include microdeletion of 5q35, encompassing the entire *NSDI* deletions

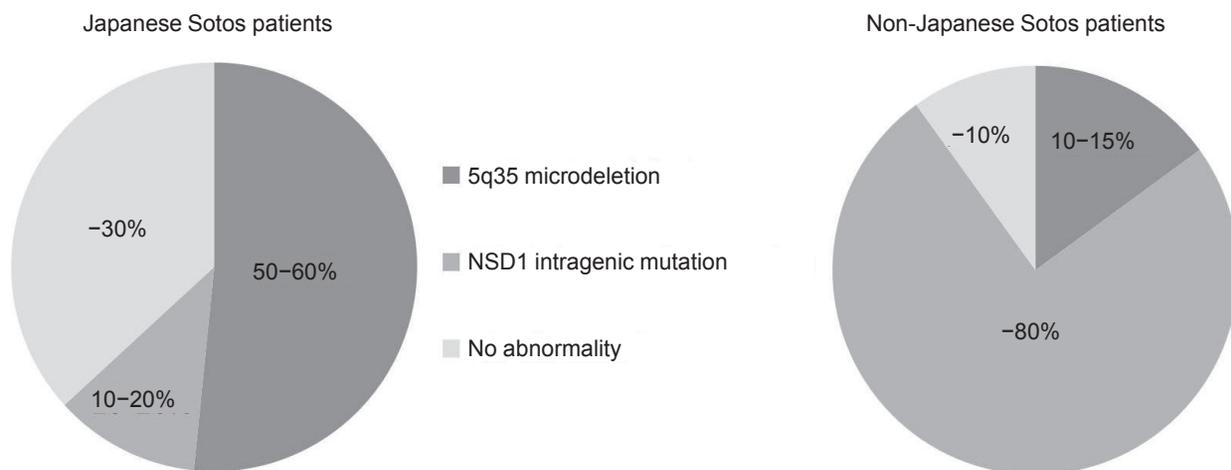


Fig. 1. Chromosome 5q35 microdeletions are more frequently found in Japanese patients with Sotos syndrome, whereas 5q35 microdeletions are uncommon in patients outside of Japan.

and mutations within the *NSD1* gene (intragenic mutations). To date, over 300 different mutations associated with Sotos syndrome have been identified, including gross deletions, small indels, point mutations, and splice site mutations (Human Gene Mutation Database, <http://www.hgmd.org/>). Notably, 5q35 microdeletions are more frequently (approximately 50%) found in Japanese patients with Sotos syndrome¹⁰, whereas 5q35 microdeletions are uncommon (< 15%) in patients in other areas of the world³(Fig. 1). Recently, the first report of SS in Korea was published¹⁴, and this study documented that 53% of patients had a 5q35 microdeletion, a result that is very similar to that in Japan. Moreover, *NSD1* abnormalities have been delineated in up to 90% of non-Japanese patients with Sotos syndrome, whereas approximately 30% of Japanese and Korean patients do not have *NSD1* abnormalities^{14,15}. 5q35 microdeletions can be detected by fluorescence *in situ* hybridization (FISH), multiplex ligation dependent probe amplification (MLPA), or array comparative genomic hybridization methods (array CGH), and *NSD1* intragenic mutations can be identified by the direct sequencing method using the patient's genomic DNA.

However, the exact mechanism of such an interracial difference has not been clearly explained to date. Patients with a microdeletion tend to have certain congenital heart and/or urogenital anomalies, more severe mental retardation, and shorter stature than those with *NSD1* intragenic mutations¹⁶.

Beckwith-Wiedemann syndrome

1. Clinical characteristics

Beckwith-Wiedemann syndrome (BWS, OMIM#130650) is the most common genetic overgrowth syndrome, with an estimated incidence of 1/13,700¹⁷. However, the actual incidence may be higher than the estimated incidence, considering the broad clinical spectrum of BWS¹⁸.

This syndrome is characterized by prenatal and postnatal overgrowth, polyhydramnios, abdominal wall defects including omphalocele, macroglossia, and visceromegaly of abdominal organs, hemihyperplasia, neonatal hyperinsulinemic hypoglycemia, ear creases and pits, cleft palate, and predisposition to embryonal tumors¹⁹. These clinical features are quite variable,

and there are no absolute criteria for a clinical diagnosis to date. Although intelligence is usually normal, mild to moderate developmental delay can be found in patients with a history of hypoglycemia and airway problems¹⁹.

Because of the well documented association between BWS and increased risk of embryonal tumor development, early diagnosis of BWS is important in the clinical setting. Wilms tumor and hepatoblastoma are most commonly accompanied, and various malignancies including neuroblastoma, rhabdomyosarcoma, adrenocortical carcinoma, and benign tumors have also been reported in BWS patients^{20,21}. The published lifetime incidence of embryonal tumors in BWS patients varies between 4 and 21% (mean, 7.5%), and most of the tumors are present in the first 8 years of life¹⁹. Therefore, tumor surveillance is particularly targeted at detecting Wilms tumor and hepatoblastoma, and abdominal ultrasonography during infancy and early childhood is highly recommended to all BWS patients²². Tumor surveillance may reduce treatment-related morbidities if tumors can be detected at early stages.

Various protocols for tumor surveillance have been proposed^{20,23,24}. Although there are no established consensus guidelines for tumor surveillance in BWS, most previous studies suggest abdominal ultrasonography every 3–6 months to the age of 8 years and measuring serum alpha fetoprotein every 2–3 months until 4 years^{20,23,24}.

2. Molecular and genetic basis

The heterogeneous molecular basis for BWS has been studied since the mapping of chromosome 11p15 as the causative locus for BWS by Waziri et al.²⁵ in 1983. However, due to the genetic complexity in BWS, the molecular diagnosis of BWS is still difficult, and the diagnosis of BWS mainly relies on clinical features.

BWS is caused by dysregulation of imprinted growth regulatory genes on chromosome 11p15²⁶. In contrast to most autosomal genes, only one allele of either the paternal or maternal allele in each imprinted gene is expressed normally. It is a parental origin-specific manner. Abnormalities in the imprinting process of a number of growth regulatory genes within the two domains on 11p15 lead to dysregulation of prenatal and postnatal growth and cause BWS¹⁹.

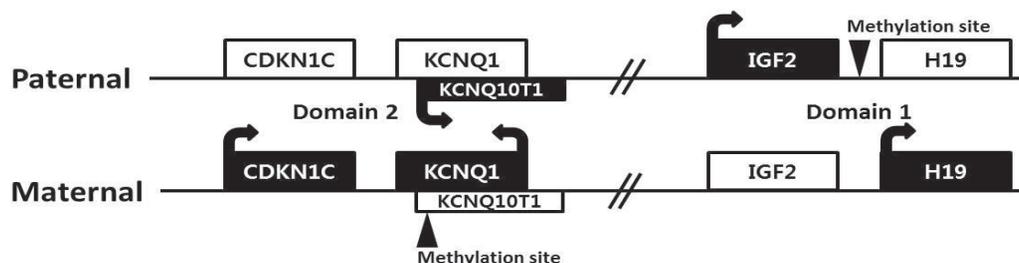


Fig. 2. Chromosome 11p15 region has two imprinting domains (domain 1 and 2) and four major genes (*IGF2*, *H19*, *CDKN1C*, and *KCNQ10T1*) associated with genomic imprinting.

There are four major genes (*IGF2*, *H19*, *CDKN1C*, and *KCNQ10T1*) associated with genomic imprinting in two imprinting domains (imprinting domain 1 and 2) on 11p15, and expression of these genes in each domain is cis-regulated by imprinting centers 1 and 2, respectively (Fig. 2). *IGF2* and *H19* are located in domain 1, and *CDKN1C* and *KCNQ10T1* are located in domain 2. Normally, *IGF2* is a paternally expressed gene and encodes insulin-like growth factor 2, which is the major growth factor in utero that promotes prenatal growth. *H19* is a maternally expressed gene and encodes an untranslated mRNA, which may function as a tumor suppressor and a growth restrictor^{27,28}. *CDKN1C* is a maternally expressed gene and encodes a cyclin-dependent kinase inhibitor that negatively regulates cell proliferation²⁹. *KCNQ10T1* is a paternally expressed gene and encodes untranslated mRNA, which may act as a negative regulator for *CDKN1C* expression³⁰. Therefore, paternally expressed genes (*IGF2* and *KCNQ10T1*) promote growth particularly in utero, and maternally expressed genes (*H19* and *CDKN1C*) restrict growth, opposing the effect of paternally expressed genes.

In BWS, alterations of these imprinting processes in the genes of domain 1 and 2 can be molecularly identified in approximately 70% of patients by various kinds of methylation studies¹⁹. Approximately 50% of patients show hypomethylation of domain 2 in the maternal allele, causing expression of maternal *KCNQ10T1*. Increased expression of *IGF2* occurs in approximately 25% of patients via paternal uniparental disomy (20%) or hypermethylation of domain 1 in the maternal allele (5%). The broad clinical spectrum of BWS is likely associated with the heterogeneity of these underlying genetic mechanisms. Furthermore, approximately 5% of patients have intragenic mutations in *CDKN1C* of maternal allele. However, more than 20% of BWS patients still remain with undetermined molecular causes¹⁹. Methylation-specific MLPA is currently the most robust method to confirm BWS, and it can detect methylation defects of domain 1 or 2 (55%), paternal uniparental disomy (20%), and microdeletion/microduplication involving domain 1 and/or 2 (rare). Cytogenetically detectable abnormalities using conventional karyotyping and FISH analysis are found only less than 1% in BWS patients. Sequence analysis of the *CDKN1C* gene can identify intragenic mutations (5%)³¹.

Interestingly, the 11p15 imprinting region is also associated with Russell-Silver syndrome (RSS, OMIM#180860), which is a typical growth retardation syndrome^{32,33}. In contrast to BWS, silencing paternally expressed genes (*IGF2* and *KCNQ10T1*) or enhancing maternally expressed genes (*H19* and *CDKN1C*) leads to RSS³⁴. Opposite epigenetic alterations in 11p15 result in opposite clinical features shown in BWS and RSS. Alterations in the imprinting process on 11p15 can be identified in approximately 50% of RSS patients^{32,33}.

Conclusions

Human growth ensues from a complex interplay of genetic traits and environmental influences. In particular, basic studies

have continued to clarify the molecular basis of growth disorders in the genetic field. Although the exact functions of the causing genes have not yet been completely understood, genetic overgrowth syndromes can be good models to clarify the complex basis of human growth and assist with the development of better-directed therapies in the future.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

We thank the patients and their families for participating in this study, which was supported by a grant no. NRF-2012R1A1A3001588 from the Korean Ministry of Science, ICT and Future Planning, and a grant of the Korean Society of Pediatric Endocrinology.

References

1. Neylon OM, Werther GA, Sabin MA. Overgrowth syndromes. *Curr Opin Pediatr* 2012;24:505-11.
2. Tatton-Brown K, Rahman N. Sotos syndrome. *Eur J Hum Genet* 2007;15:264-71.
3. Tatton-Brown K, Douglas J, Coleman K, Baujat G, Cole TR, Das S, et al. Genotype-phenotype associations in Sotos syndrome: an analysis of 266 individuals with NSD1 aberrations. *Am J Hum Genet* 2005;77:193-204.
4. Sotos JF, Dodge PR, Muirhead D, Crawford JD, Talbot NB. Cerebral gigantism in childhood: a syndrome of excessively rapid growth and acromegalic features and a nonprogressive neurologic disorder. *N Engl J Med* 1964;271:109-16.
5. Cole TR, Hughes HE. Sotos syndrome: a study of the diagnostic criteria and natural history. *J Med Genet* 1994;31:20-32.
6. Agwu JC, Shaw NJ, Kirk J, Chapman S, Ravine D, Cole TR. Growth in Sotos syndrome. *Arch Dis Child* 1999;80:339-42.
7. Allanson JE, Cole TR. Sotos syndrome: evolution of facial phenotype subjective and objective assessment. *Am J Med Genet* 1996;65:13-20.
8. Wit JM, Beemer FA, Barth PG, Oorthuys JW, Dijkstra PF, Van den Brande JL, et al. Cerebral gigantism (Sotos syndrome). Compiled data of 22 cases. Analysis of clinical features, growth and plasma somatomedin. *Eur J Pediatr* 1985;144:131-40.
9. Hersh JH, Cole TR, Bloom AS, Bertolone SJ, Hughes HE. Risk of malignancy in Sotos syndrome. *J Pediatr* 1992;120(4 Pt 1):572-4.
10. Kurotaki N, Imaizumi K, Harada N, Masuno M, Kondoh T, Nagai T, et al. Haploinsufficiency of NSD1 causes Sotos syndrome. *Nat Genet* 2002;30:365-6.

11. de Boer L, Kant SG, Karperien M, van Beers L, Tjon J, Vink GR, et al. Genotype-phenotype correlation in patients suspected of having Sotos syndrome. *Horm Res* 2004;62:197-207.
12. Kurotaki N, Harada N, Yoshiura K, Sugano S, Niikawa N, Matsumoto N. Molecular characterization of NSD1, a human homologue of the mouse Nsd1 gene. *Gene* 2001;279:197-204.
13. Huang N, vom Baur E, Garnier JM, Lerouge T, Vonesch JL, Lutz Y, et al. Two distinct nuclear receptor interaction domains in NSD1, a novel SET protein that exhibits characteristics of both corepressors and coactivators. *EMBO J* 1998;17:3398-412.
14. Sohn YB, Lee CG, Ko JM, Yang JA, Yun JN, Jung EJ, et al. Clinical and genetic spectrum of 18 unrelated Korean patients with Sotos syndrome: frequent 5q35 microdeletion and identification of four novel NSD1 mutations. *J Hum Genet* 2013;58:73-7.
15. Niikawa N. Molecular basis of Sotos syndrome. *Horm Res* 2004;62 Suppl 3:60-5.
16. Nagai T, Matsumoto N, Kurotaki N, Harada N, Niikawa N, Ogata T, et al. Sotos syndrome and haploinsufficiency of NSD1: clinical features of intragenic mutations and submicroscopic deletions. *J Med Genet* 2003;40:285-9.
17. Engstrom W, Lindham S, Schofield P. Wiedemann-Beckwith syndrome. *Eur J Pediatr* 1988;147:450-7.
18. Greer KJ, Kirkpatrick SJ, Weksberg R, Pauli RM. Beckwith-Wiedemann syndrome in adults: observations from one family and recommendations for care. *Am J Med Genet A* 2008;146A:1707-12.
19. Choufani S, Shuman C, Weksberg R. Beckwith-Wiedemann syndrome. *Am J Med Genet C Semin Med Genet* 2010;154C:343-54.
20. Tan TY, Amor DJ. Tumour surveillance in Beckwith-Wiedemann syndrome and hemihyperplasia: a critical review of the evidence and suggested guidelines for local practice. *J Paediatr Child Health* 2006;42:486-90.
21. Cohen MM Jr. Beckwith-Wiedemann syndrome: historical, clinicopathological, and etiopathogenetic perspectives. *Pediatr Dev Pathol* 2005;8:287-304.
22. DeBaun MR, Tucker MA. Risk of cancer during the first four years of life in children from The Beckwith-Wiedemann Syndrome Registry. *J Pediatr* 1998;132(3 Pt 1):398-400.
23. Clericuzio CL, Martin RA. Diagnostic criteria and tumor screening for individuals with isolated hemihyperplasia. *Genet Med* 2009;11:220-2.
24. Zarate YA, Mena R, Martin LJ, Steele P, Tinkle BT, Hopkin RJ. Experience with hemihyperplasia and Beckwith-Wiedemann syndrome surveillance protocol. *Am J Med Genet A* 2009;149A:1691-7.
25. Waziri M, Patil SR, Hanson JW, Bartley JA. Abnormality of chromosome 11 in patients with features of Beckwith-Wiedemann syndrome. *J Pediatr* 1983;102:873-6.
26. Weksberg R, Shuman C, Smith AC. Beckwith-Wiedemann syndrome. *Am J Med Genet C Semin Med Genet* 2005;137C:12-23.
27. Hao Y, Crenshaw T, Moulton T, Newcomb E, Tycko B. Tumour-suppressor activity of H19 RNA. *Nature* 1993;365:764-7.
28. Guo L, Choufani S, Ferreira J, Smith A, Chitayat D, Shuman C, et al. Altered gene expression and methylation of the human chromosome 11 imprinted region in small for gestational age (SGA) placentae. *Dev Biol* 2008;320:79-91.
29. Tsugu A, Sakai K, Dirks PB, Jung S, Weksberg R, Fei YL, et al. Expression of p57(KIP2) potently blocks the growth of human astrocytomas and induces cell senescence. *Am J Pathol* 2000;157:919-32.
30. Cerrato F, Vernucci M, Pedone PV, Chiariotti L, Sebastio G, Bruni CB, et al. The 5' end of the KCNQ10T1 gene is hypomethylated in the Beckwith-Wiedemann syndrome. *Hum Genet* 2002;111:105-7.
31. Weksberg R, Shuman C, Beckwith JB. Beckwith-Wiedemann syndrome. *Eur J Hum Genet* 2010;18:8-14.
32. Eggermann T, Begemann M, Binder G, Spengler S. Silver-Russell syndrome: genetic basis and molecular genetic testing. *Orphanet J Rare Dis* 2010;5:19.
33. Eggermann T. Russell-Silver syndrome. *Am J Med Genet C Semin Med Genet* 2010;154C:355-64.
34. Eggermann T. Silver-Russell and Beckwith-Wiedemann syndromes: opposite (epi)mutations in 11p15 result in opposite clinical pictures. *Horm Res* 2009;71 Suppl 2:30-5.