



EDTA Treatment for Overcoming the Prozone Effect and for Predicting C1q Binding in HLA Antibody Testing

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The Luminex-based single antigen bead (SAB) assay is widely used to detect HLA antibody in transplant recipients. However, one limitation of the SAB assay is the prozone effect, which occurs mostly as a result of complement interference. We investigated the efficacy of EDTA treatment for overcoming the prozone effect and predicting C1q binding of HLA antibody. We subjected 27 non-treated (naïve) and EDTA-treated serum samples from highly sensitized patients to IgG-SAB assays, and we confirmed the prozone effect in 53% and 31% of class I and class II antibody tests, respectively, after EDTA treatment. When we conducted additional assays after dithiothreitol treatment and serum dilution, EDTA was the most efficacious in eliminating the prozone effect. Reducing the prozone effect by EDTA treatment strengthened the correlation between IgG mean fluorescence intensity (MFI) and C1q MFI values ($p=0.825$) as compared with the naïve sera ($p=0.068$). Although C1q positivity was dependent on the concentration of HLA antibody in EDTA-treated sera, the correlations varied individually. Overall, our results confirmed the efficacy of EDTA treatment for overcoming the prozone effect. EDTA treatment showed a positive effect on the correlation between IgG MFI and C1q MFI values.

Key Words: Prozone effect, EDTA, Single antigen bead assay, C1q binding, HLA antibody

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Accurate assessment of HLA antibody is crucial for successful transplant management [1]. The Luminex-based single antigen bead (SAB) assay is widely used for sensitive detection of low concentrations of antibody [2-5]. In the SAB assay, antibody amount is determined semi-quantitatively and is expressed as the mean fluorescence intensity (MFI) after reaction with anti-human IgG antibody reagent labeled with a fluorescent dye. MFI values correlate with transplant-related outcomes; thus, monitoring MFI values may be useful for risk assessment after transplantation [6, 7]. In addition, SAB assays for detecting C1q-binding HLA antibodies (C1q SAB) have been introduced to better discriminate clinically relevant antibodies [8].

However, clinical application of the SAB assay is limited because of technical issues, including assay variability, complement

interference, and the prozone effect [9]. Inhibition causing false-negative or falsely decreased MFI values has been observed in approximately 70% of highly sensitized patients [10]. Several studies have attempted to eliminate the prozone effect by treatment with EDTA or dithiothreitol (DTT), preheating, and/or dilution of serum [9, 11, 12]. There are multiple explanations for the prozone effect, including bead saturation [13, 14], steric hindrance from complement complex [11], and the presence of IgM HLA-specific antibodies [15]. Complement interference involves covalent binding and accumulation of C4 and C3 degradation products on the immune complex, which prevent fluorescent anti-IgG from binding to IgG. EDTA eliminates the prozone effect because, as an iron-chelating reagent, it can inhibit the formation of C1qrs complement complex and prevent com-

plement cascade activation [16]. DTT disrupts the pentameric structure of IgM and the disulfide bonds in complement molecules, thus reducing the prozone effect [15]. In this respect, SAB assays as a standard should include serum treatment to correct the prozone effect because MFI values are often used as a surrogate marker of antibody strength. Although a recent meeting report recommended EDTA and/or titration [1], there is currently no verified standard method, and different treatment methods are used across different laboratories.

We compared MFI values of SAB assays in serum samples treated with or without EDTA, DTT, or dilution. In addition, we evaluated the correlation between IgG-MFI and C1q-MFI values after EDTA treatment. To our knowledge, this is the first study to investigate the effects of EDTA on specific HLA loci in combination with C1q-MFI.

We collected leftover sera from 27 highly sensitized patients (median age=49 years, M:F=14:13). All samples contained strong HLA antibodies (MFI \geq 10,000), and the median panel-reactive antibody (%) was 95%. The patients had become sensitized by previous transplantation with or without transfusion or pregnancy (81.5%, N=22), previous pregnancy and transfusion (7.4%, N=2), previous pregnancy alone (7.4%, N=2), or previous transfusion alone (3.7%, N=1). The Institutional Review Board of Seoul St. Mary's Hospital, Seoul, Korea, approved this study (KC18SESI0323). LABScreen Single Antigen kits and

the LABScan 3D system (One Lambda, A Thermo Fisher Scientific Brand, Canoga Park, CA, USA) were used to detect HLA antibodies (11 samples for class I only, eight samples for class II only, and eight samples for both class I and class II). For EDTA treatment, a 0.5 M EDTA solution was diluted 1:20 (final concentration of 25 mM), added to the serum, vortexed, and centrifuged at room temperature for 10 minutes before testing [10]. We defined the prozone effect as doubled MFI values, with an MFI cutoff of 1,000, or 5,000 MFI increment after serum treatment compared with the results from naïve serum, considering recent recommendations and reports [1, 12, 17].

In a comparison of naïve and EDTA-treated samples, we investigated MFI data from 3,363 beads (1,843 class I and 1,520 class II) from 35 tests (19 class I and 16 class II) (Fig. 1). The prozone effect was identified in 53% (10 out of 19) of class I antibody tests and in 31% (5 out of 16) of class II antibody tests after EDTA treatment. Of all beads, 8.3% (26.5% of positive beads) were affected by complement inhibition, with the median MFI increasing from 2,921 (range, 0–13,890) to 21,043 (1,024–27,874) after EDTA treatment. Regarding HLA specificity, prozone effect incidences were as follows: HLA-A, 28.1%; HLA-B, 43.6%; HLA-C, 3.6%; HLA-DR, 2.7%; HLA-DQ, 18.0%; and HLA-DP, <0.1% of positive beads. These findings were consistent with a previous report wherein HLA-C beads were the least affected among class I beads and HLA-DQ beads were the

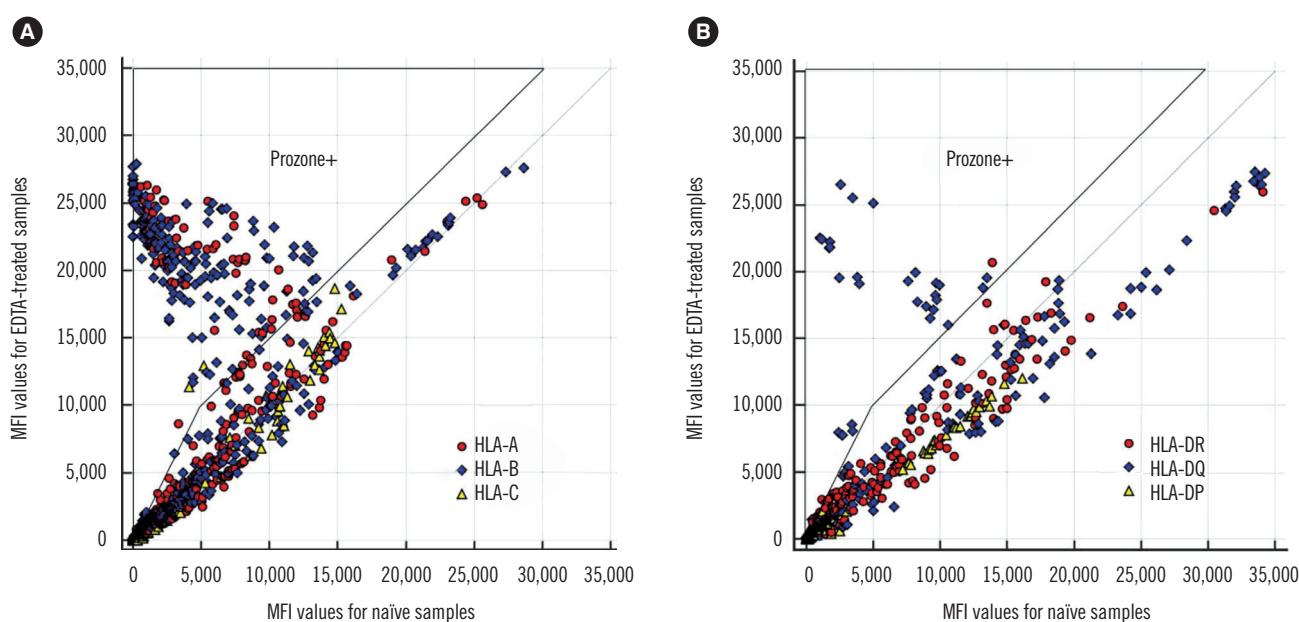


Fig. 1. Comparison of MFI values in naïve and EDTA-treated sera. (A) Class I IgG-SAB results in 19 sera (1,843 beads); (B) Class II IgG-SAB results in 16 sera (1,520 beads).

Abbreviations: MFI, mean fluorescence intensity; SAB, single antigen bead.

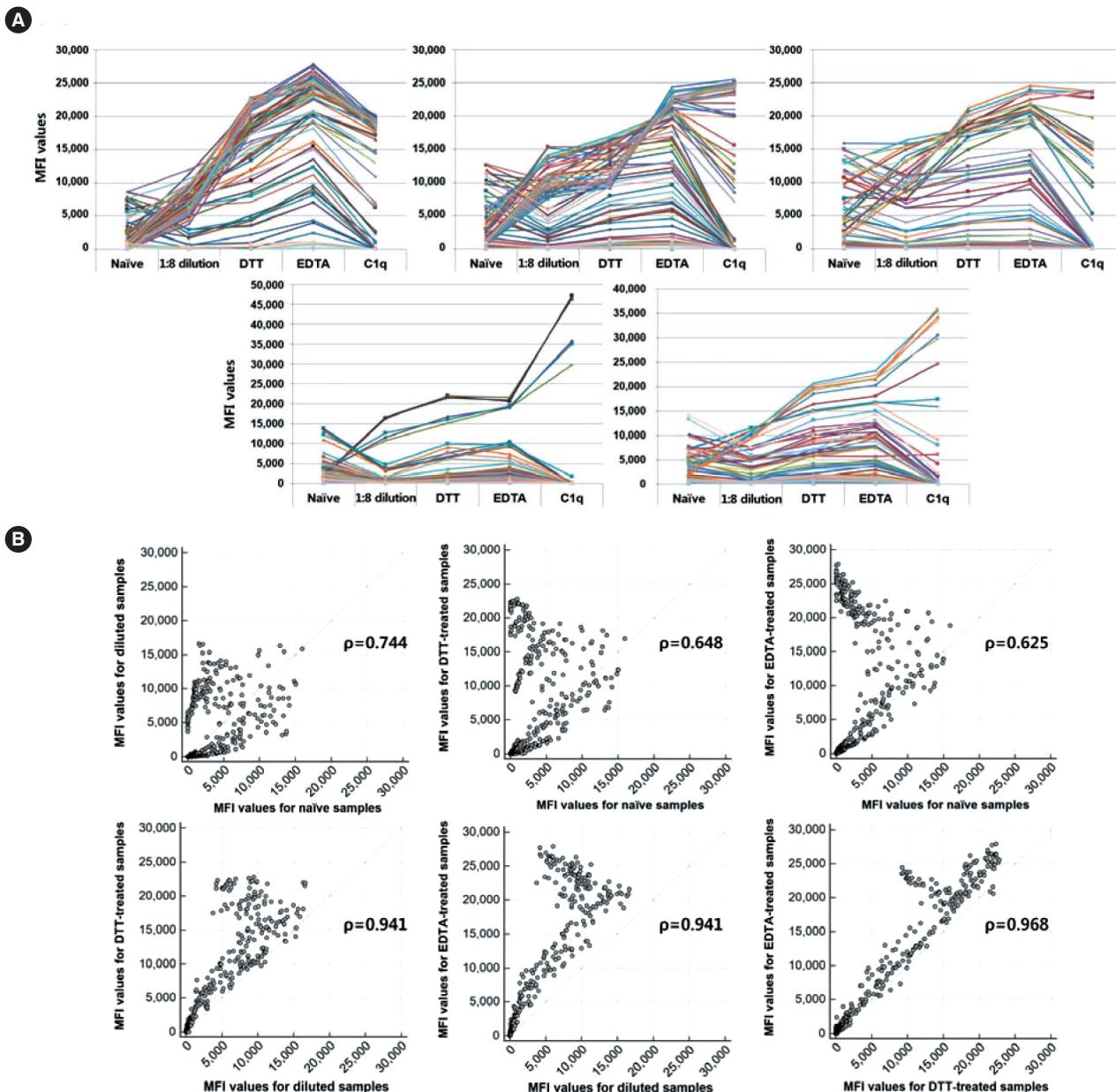


Fig. 2. MFI values for naïve samples and samples treated with 1:8 dilution, DTT, and EDTA, and C1q assay using sera from five sensitized patients. (A) MFI values for all samples with various treatments. (B) Correlations between MFI values for naïve, diluted, DTT-treated, and EDTA-treated samples.

Abbreviations: MFI, mean fluorescence intensity; DTT, dithiothreitol.

most affected among class II beads [18].

We selected five sera and additionally tested class I IgG after 1:8 dilution in phosphate-buffered saline and treatment with 5 mM DTT at 37°C for 30 minutes [19]. When we compared the three serum treatment methods, EDTA treatment was the most efficacious for overcoming prozone effect (Fig. 2). This finding was consistent with previous studies that showed the superior efficacy of EDTA for overcoming the prozone effect [12, 16].

DTT treatment and dilution resulted in small increases in MFI values in some of the beads, indicating a residual prozone effect, presumably because only IgM antibodies were removed (Supplemental Data Table S1).

We subjected 10 samples with the prozone effect ($N=5$ for class I and $N=5$ for class II HLA antibodies) to C1qScreen assays (One Lambda). We selected beads that were positive in EDTA-treated sera to calculate Spearman's correlation coeffi-

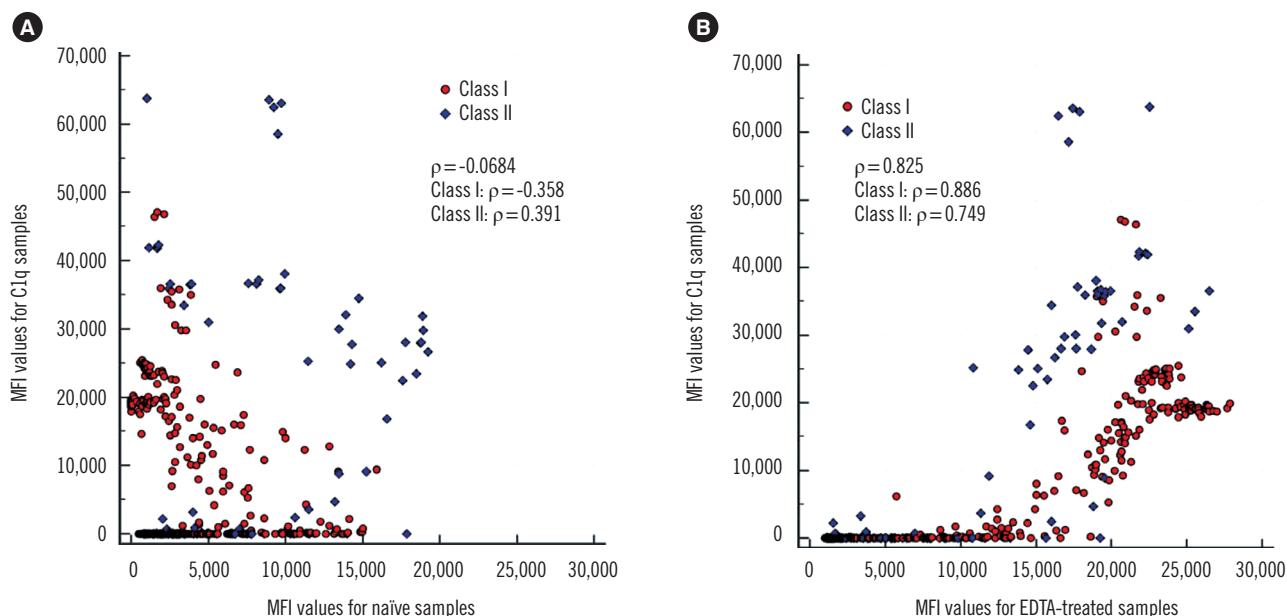


Fig. 3. Correlation of MFI values from C1q- and IgG-SAB assays using (A) naïve sera and (B) EDTA-treated sera.

Abbreviations: MFI, mean fluorescence intensity; SAB, single antigen bead.

cient (ρ). MFI values were summarized as median (range). For statistical analysis, we used MedCalc Statistical Software version 18.9 (MedCalc Software, Ostend, Belgium).

When naïve sera were used, the correlation between IgG-MFI values and C1q-MFI values was weak ($\rho=-0.0684$; class I: -0.358, class II: 0.391). After EDTA treatment, the correlation between IgG-MFI values and C1q-MFI values became stronger ($\rho=0.825$; class I: 0.886, class II: 0.749) (Fig. 3). Beads that displayed low IgG-MFI values in naïve sera, but high C1q-MFI values, showed high IgG-MFI values after EDTA treatment. In naïve sera, C1q-SAB beads with $\text{MFI} \geq 1,000$ (19.7% of all beads) had a median IgG-MFI value of 2,842 (range, 0–19,256). In EDTA-treated sera, C1q-positive beads had a median IgG-MFI value of 20,900 (1,539–27,874). These results supported previous findings that C1q binding ability depends on the concentration of HLA antibody [12]. Tambur and Wiebe [9] reported that MFI cutoffs that predicted C1q positivity varied from 6,237 to 14,154 using non-treated samples. However, not all beads that had high IgG-MFI values in EDTA-treated sera showed C1q positivity, and some high-level IgG antibodies did not bind to C1q. When we plotted IgG-MFI against C1q-MFI values from the data in 10 patients separately, we found a wide range of correlation coefficients (0.283–0.977). Therefore, it remains to be elucidated whether there is an interdependence between C1q positivity and IgG-MFI values.

Some limitations of the present study were that we could not

test serial dilutions of samples, and we did not confirm the HLA-specific antibodies against intact HLA antigen on the lymphocyte. Because high-level antibodies against denatured HLA protein might also bind to C1q [20], future studies on interference by denatured HLA are needed. In addition, we selected samples with high panel-reactive antibody and strong HLA antibodies. Thus, our results may not represent the frequency of the prozone effect in SAB assays in general.

In conclusion, we confirmed the efficacy of EDTA treatment for eliminating the prozone effect. Further, EDTA treatment had a positive effect on the correlation between IgG MFI and C1q MFI values. Our study highlights the importance of method standardization to correct the prozone effect.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article are reported.

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Supplemental Data Table S1. MFI values of Class I 97 beads in SAB assays among naïve, dilution, DTT, and EDTA treatments using sera from five sensitized patients

Allele specificity	Sample 1			Sample 2			Sample 3			Sample 4			Sample 5			
	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA
A*01:01	398	225	173	476	3,427	705*	1,008	3,979	10,420	6,042	10,426	11,662	360	1	110	242
A*02:01	2,603	10,939	18,806	23,628	5,955	7,808	10,322	15,526	0	0	0	0	1,697	16,341	21,951	20,609
A*02:03	3,884	8,599	15,035	18,954	7,753	4,861	8,316	12,439	4	0	0	0	1,522	16,639	21,891	21,618
A*02:06	4,748	13,304	17,832	21,865	8,604	6,309	8,706	13,494	0	0	0	0	2,119	16,546	21,507	20,909
A*03:01	649	117	396	542	7,757	2,256	4,526	9,643	11,483	7,545	13,046	14,650	2	0	0	16
A*11:01	96	3	91	98	7,408	1,657	4,317	9,099	12,637	8,527	14,907	16,611	4,095	613*	1,318	2,395
A*11:02	0	0	0	0	3,338	1,842	5,070	8,654	11,378	10,133	16,527	18,606	3,385	643*	1,545	2,959
A*23:01	348	108	172	409	0	0	0	0	11,931	5,041	10,814	12,272	1,687	118*	40*	1,121
A*24:02	843	68	353	764	0	0	0	0	10,198	10,411	15,444	16,329	0	0	0	0
A*24:03	1,356	158*	623*	1,284	0	0	0	0	9,820	8,341	12,669	12,886	285	0	0	113
A*25:01	13	0	0	21	2,621	8,598	13,329	20,257	8,709	2,998	7,971	9,610	13,776	3,726	7,606	10,382
A*26:01	141	0	170	274	2,457	7,278	14,464	20,788	5,032	1,589	4,617	5,674	13,153	3,202	6,439	9,260
A*29:01	446	107	177	493	1,475	8,516	18,162	24,266	630	119	645	961	1,122	61*	313*	619*
A*29:02	707	217	314	728	1,081	8,372	18,021	24,572	804	155	752	1,136	1,218	70*	297*	675*
A*30:01	603	132	450	559	1,009	9,208	22,240	26,329	476	60	444	709	0	0	0	16
A*30:02	49	0	42	46	2,284	11,327	21,799	25,200	383	58	407	632	0	0	0	1
A*31:01	0	0	0	0	0	6,551	21,886	25,952	584	70	504	801	653	23	255	325
A*32:01	187	20	112	137	2,224	8,820	13,867	20,658	5,580	1,423	4,539	6,169	698	68	208	391
A*33:01	4	0	4	0	1,260	8,679	19,647	25,417	2,504	1,169	2,922	3,475	2,843	328*	934*	1,587
A*33:03	0	0	0	0	0	808	8,317	19,464	846	210	856	1,203	3,929	488*	1,216	2,135
A*34:01	182	10	165	196	3,135	9,654	18,301	23,785	802	238	935	1,288	3,991	464*	1,163	2,296
A*34:02	773	143	563	678	584	7,355	18,737	26,201	1,705	620*	1,708	2,269	5,113	623*	1,586	3,144
A*36:01	53	0	23	74	2,025	171*	137*	2,403	2,647	2,894	3,950	4,595	229	0	51	153
A*43:01	254	54	332	482	2,842	6,596	11,931	19,174	6,301	2,123	5,634	6,948	6,681	1,086	2,293	3,954
A*66:01	42	0	64	138	1,112	7,450	18,541	24,466	6,614	2,004	5,744	6,854	13,653	3,408	6,786	9,822
A*66:02	10,671	6,154	6,517	9,830	1,599	8,757	18,816	24,792	1,243	349*	1,413	1,744	2,242	200*	560*	1,257
A*68:01	5,855	7,548	18,125	21,625	7,262	2,927	3,424	7,039	765	160	686	1,048	3,560	10,696	15,125	19,141
A*68:02	7,701	7,400	16,826	20,575	7,242	2,965	3,659	7,066	4,640	1,696	4,248	5,880	3,100	11,594	16,755	19,037
A*69:01	7,544	9,064	16,814	19,805	6,595	2,938	4,529	8,103	557	95	479	764	3,880	12,663	16,017	19,436
A*74:01	957	194	745	691	711	7,711	19,304	25,306	532	106	474	790	588	70	250	370

(Continued to the next page)

Supplemental Data Table S1. Continued

Allele specificity	Sample 1			Sample 2			Sample 3			Sample 4			Sample 5			
	Näive	Dilution	DTT	EDTA	Näive	Dilution	DTT	EDTA	Näive	Dilution	DTT	EDTA	Näive	Dilution	DTT	EDTA
A*80:01	151	68	171	111	4,090	554*	2,183	4,338	7,236	2,646	6,618	7,082	311	0	98	229
B*07:02	9,802	14,960	18,646	20,929	0	0	0	0	1,328	440*	1,512	1,732	5,017	704*	2,209	2,861
B*08:01	1,062	575*	1,164	1,045	0	0	0	157	7,770	7,883	13,732	15,575	165	0	0	114
B*13:01	14,831	11,111	11,800	13,324	3,772	7,724	14,183	20,658	5,306	13,521	16,342	19,563	0	0	0	0
B*13:02	5,319	14,007	17,940	20,492	148	5,741	18,143	24,893	2,060	15,269	14,032	22,063	0	0	0	5
B*14:01	55	0	41	65	576	6,382	18,850	22,545	2,838	11,222	13,861	18,703	41	0	0	46
B*14:02	33	0	27	41	2,548	8,384	17,389	20,007	6,100	7,684	11,902	13,172	0	0	0	5
B*15:01	10,700	7,384	8,632	10,285	0	4,994	21,178	26,986	840	9,061	9,847	23,570	0	0	0	0
B*15:02	951	364	801	843	0	4,384	22,332	26,493	707	7,701	9,197	23,540	70	0	0	51
B*15:03	9,326	6,185	7,788	8,936	0	6,011	22,373	26,368	941	9,375	10,022	22,923	0	0	0	16
B*15:10	4,564	2,754	3,785	4,573	0	5,728	21,771	26,463	588	7,657	9,291	23,637	138	0	27	99
B*15:11	883	488	907	914	361	6,912	22,533	25,753	1,093	10,825	11,751	21,860	21	0	0	20
B*15:12	4,939	2,607	4,193	5,043	0	4,221	21,784	21,682	730	8,521	9,278	24,426	4,507	629*	1,840	2,763
B*15:13	204	107	225	253	493	6,676	21,822	24,442	1,688	11,071	12,119	22,035	539	21	50	374
B*15:16	0	0	0	0	0	5,151	21,097	25,808	839	9,927	10,173	22,304	2,683	88*	1,032	1,769
B*18:01	125	0	66	111	240	6,039	22,309	27,874	2,112	13,795	13,362	22,671	56	0	0	51
B*27:05	2,056	9,546	20,644	23,873	5,636	6,447	7,970	12,309	0	0	0	0	12,246	4,750	9,918	9,649
B*27:08	1,785	8,840	21,170	24,630	2,643	8,058	11,848	16,220	0	0	0	0	10,921	3,586	9,008	7,311
B*35:01	2,023	966*	1,901	1,985	81	5,997	20,824	25,787	1,219	9,341	10,418	22,104	49	0	16	49
B*37:01	6,879	11,854	19,147	22,436	0	4,901	17,881	23,405	3,998	12,256	14,624	19,462	1,960	203*	637*	1,166
B*38:01	13,296	8,629	11,114	12,638	53	5,393	18,697	23,241	3,149	13,688	15,843	20,680	5,350	859*	2,458	3,525
B*39:01	11,980	6,800	10,371	11,456	0	3,737	17,517	22,511	2,679	12,535	15,527	20,745	3,455	429*	1,472	2,376
B*40:01	12,864	16,318	18,128	20,678	0	0	0	0	4,590	14,123	16,949	20,683	2,717	337*	1,288	1,477
B*40:02	10,001	15,581	18,104	20,900	0	0	0	0	3,656	13,658	15,800	21,309	1,781	179*	677*	1,037
B*40:06	15,922	15,838	16,995	18,856	0	0	0	0	5,938	13,029	16,199	19,842	871	112	499	530
B*41:01	14,994	10,794	12,400	13,959	0	0	0	110	2,989	15,355	15,297	21,138	682	95	254	412
B*42:01	4,482	13,146	19,135	21,774	786	150	0	855	81	1	109	122	2,051	190*	677*	1,150
B*44:02	3,434	1,075	2,809	2,964	8,619	11,974	16,266	18,990	2,815	13,869	13,323	21,298	5,605	858*	2,027	3,360
B*45:01	4,655	1,706	3,945	4,278	1,256	8,695	20,114	23,775	922	10,458	9,773	22,811	4,564	718*	1,900	2,836

(Continued to the next page)

Supplemental Data Table S1. Continued

Allele specificity	Sample 1				Sample 2				Sample 3				Sample 4				Sample 5				
	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	
B*46:01	0	0	13	4	0	<u>5.575</u>	<u>21,683</u>	<u>25,195</u>	2,974	<u>13,798</u>	<u>14,405</u>	<u>20,933</u>	0	0	0	0	29	0	91	47	
B*47:01	14,042	8,646	11,219	12,849	8,460	6,391	7,081	11,184	12,828	10,237	15,749	16,913	2,598	226*	616*	1,453	9,721	5,604	8,219	9,699	
B*48:01	13,405	15,434	17,495	19,415	4,926	8,614	15,223	19,252	4,208	<u>13,779</u>	<u>16,608</u>	<u>20,212</u>	2,409	246*	958*	1,265	3,242	10,932	19,226	21,672	
B*49:01	11,637	8,707	10,536	11,785	947	<u>9,445</u>	<u>22,805</u>	<u>25,531</u>	790	<u>8,046</u>	<u>9,610</u>	<u>23,851</u>	2,887	305*	784*	1,859	7,776	5,671	8,384	9,727	
B*50:01	13,252	9,989	11,198	12,334	1,005	<u>9,794</u>	<u>22,058</u>	<u>24,262</u>	853	<u>8,969</u>	<u>9,926</u>	<u>23,771</u>	303	33	145	212	4,349	7,383	13,193	15,020	
B*51:01	0	0	0	0	0	<u>5,899</u>	<u>22,166</u>	<u>26,356</u>	997	<u>9,389</u>	<u>10,351</u>	<u>22,787</u>	1,583	166*	430*	1,049	0	0	6	0	
B*51:02	362	204	491	563	0	<u>5,381</u>	<u>21,346</u>	<u>26,417</u>	848	<u>8,343</u>	<u>9,911</u>	<u>22,950</u>	1,494	123*	250*	963*	58	0	91	74	
B*52:01	816	271	830	802	1,016	<u>8,997</u>	<u>22,059</u>	<u>24,913</u>	1,452	<u>11,205</u>	<u>12,232</u>	<u>21,790</u>	707	0	46	448	654	394	821	1,178	
B*53:01	2,355	1,048	1,995	2,021	103	<u>6,268</u>	<u>21,245</u>	<u>25,311</u>	1,252	<u>10,543</u>	<u>11,129</u>	<u>21,817</u>	1,897	138*	518*	1,182	609	202	990	841	
B*54:01	11,317	9,452	13,552	14,919	4,535	<u>9,756</u>	<u>15,842</u>	<u>18,960</u>	6,514	2,285	5,769	7,683	348	0	115	237	3,698	1,515	3,810	4,359	
B*55:01	15,036	10,604	12,340	13,916	7,616	10,561	<u>14,611</u>	<u>18,182</u>	7,357	3,866	7,147	8,586	792	92	315	494	3,293	1,264	3,160	3,510	
B*56:01	6,685	<u>13,129</u>	<u>17,589</u>	<u>19,757</u>	1,156	<u>9,873</u>	<u>22,155</u>	<u>24,834</u>	2,209	<u>12,824</u>	<u>14,210</u>	<u>20,445</u>	600	29	259	396	4,025	2,843	6,037	7,519	
B*57:01	0	0	0	0	0	<u>5,184</u>	<u>21,056</u>	<u>25,586</u>	1,329	<u>10,491</u>	<u>11,108</u>	<u>22,157</u>	4,547	1,429	6,256	5,615	228	65	450	468	
B*57:03	0	0	0	0	0	<u>4,219</u>	<u>21,159</u>	<u>26,756</u>	895	<u>9,647</u>	<u>10,172</u>	<u>22,670</u>	3,033	1,512	<u>7,629</u>	<u>6,398</u>	511	200	967	1,024	
B*58:01	0	0	0	0	720	<u>8,737</u>	<u>22,653</u>	<u>25,153</u>	1,267	<u>10,971</u>	<u>11,613</u>	<u>22,728</u>	4,886	1,428	6,018	5,547	576	168	897	968	
B*59:01	7,153	3,989	6,374	6,632	5,066	7,269	<u>11,018</u>	<u>15,030</u>	6,357	11,237	<u>15,459</u>	<u>17,676</u>	1,361	79*	330*	911*	529	188	721	656	
B*67:01	2,010	<u>10,178</u>	<u>20,079</u>	<u>23,290</u>	655	<u>7,025</u>	<u>17,373</u>	<u>20,863</u>	97	0	97	126	2,357	246*	967*	1,465	6,473	4,760	7,275	8,857	
B*73:01	2,132	<u>9,505</u>	<u>20,132</u>	<u>23,358</u>	672	166	397	1,148	119	0	94	162	2,221	180*	936*	1,235	5,957	9,239	14,733	16,464	
B*78:01	11	0	0	0	0	<u>5,583</u>	<u>21,542</u>	<u>26,469</u>	1,049	<u>10,026</u>	<u>10,639</u>	<u>22,900</u>	209	21	102	158	358	159	491	475	
B*81:01	2,868	<u>11,109</u>	<u>20,021</u>	<u>23,651</u>	0	0	0	0	1,368	438*	1,434	1,683	5,007	695*	2,084	2,805	2,621	<u>9,525</u>	<u>20,293</u>	<u>22,345</u>	
B*82:01	11,236	13,365	<u>16,253</u>	<u>18,447</u>	4,442	<u>9,965</u>	<u>17,360</u>	<u>20,610</u>	6,462	2,506	6,381	7,558	2,988	442*	1,287	1,908	898	245	772	832	
C*01:02	514	107	377	427	0	0	0	9	30	0	71	41	171	0	25	86	26	0	46	23	
C*02:02	1,351	505*	1,058	1,097	388	386	0	852	736	254	884	967	0	0	0	0	13,316	6,023	6,863	12,998	
C*03:02	0	0	0	0	0	0	0	0	77	11,526	5,983	11,811	13,030	0	0	0	0	705	514	1,475	1,233
C*03:03	0	0	0	0	0	0	0	0	10,954	4,705	10,055	11,415	0	0	0	0	1,013	729*	2,022	1,522	
C*03:04	2	0	2	0	0	0	0	0	9,763	4,302	9,357	10,691	0	0	0	0	571	442	<u>1,290</u>	935	
C*04:01	61	0	82	87	355	50	0	625	222	42	407	535	0	0	0	1	0	8	0	0	
C*05:01	0	0	52	11	0	0	0	0	724	143	789	854	0	0	0	0	18	0	75	14	
C*06:02	60	0	187	67	0	0	0	192	738	142	767	868	0	0	194	0	52	0	130	78	
C*07:02	38	0	130	46	0	0	0	0	173	55	468	564	1,118	45*	113*	574*	1,071	521*	1,834	1,248	

(Continued to the next page)

Supplemental Data Table S1. Continued

Allele specificity	Sample 1			Sample 2			Sample 3			Sample 4			Sample 5							
	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA	Naïve	Dilution	DTT	EDTA				
C*08:01	115	0	151	124	0	0	0	0	17	0	39	36	0	0	0	567	125	732	544	
C*12:03	77	0	110	69	0	0	0	97	64	0	127	108	0	0	0	33	0	43	29	
C*14:02	52	0	61	59	0	0	0	24	57	0	104	132	0	0	0	0	0	13	0	
C*15:02	67	0	120	120	0	0	0	240	756	146	759	1,000	697	134	204	261	0	0	21	0
C*16:01	47	0	125	58	0	0	0	144	52	0	105	71	0	0	2	548	236	946	686	
C*17:01	207	25	359	299	890	297	823	1,184	868	297	1,379	1,371	1,067	59*	279*	538*	14,122	7,546	9,791	14,438
C*18:02	24	0	88	31	0	0	0	229	750	152	823	828	0	0	158	0	36	0	86	31

Beads exhibiting the prozone effect are bolded and underscored.

*MFI value of bead was $\geq 1,000$ in the naïve sample and decreased to $<1,000$ after serum treatment.
 Abbreviations: MFI, mean fluorescence intensity; SAB, single antigen bead; DTT, dithiothreitol.