

# The Effects of Cyclosporin A and FK-506 on the Cytokine Production of Lymphocytes in Atopic Dermatitis

Chun Wook Park, M.D., Kyung Ywal Lee, M.D.,  
Eun Hee Lee, M.D.\*, Cheol Heon Lee, M.D.

*Department of Dermatology, College of Medicine, Hallym University  
and Green Cross Reference Lab\*, Seoul, Korea*

**Background:** It has been demonstrated that patients with atopic dermatitis(AD) show an impaired capacity of their T cells to release of interleukin-2(IL-2) *in vitro* and elevated serum levels of soluble IL-2 receptor(sIL-2R). Both immunosuppressive agents, cyclosporin A(CsA) and FK-506 can block early events in T lymphocyte activation and FK-506 is 10- to 100-fold more potent in the inhibition of IL-2 and other lymphokines production.

**Objective:** We compared the effects of CsA and FK-506 on PHA-induced lymphokine and sIL-2R production and compared the effects of CsA and FK-506 on PHA-induced IL-4 production in a high IgE group and a low IgE group in patients with AD.

**Methods:** A total of 32 peripheral blood samples from 17 patients with AD and 15 control groups were tested. Lymphocytes were isolated from blood samples and were cultured with PHA(positive control), PHA and CsA(10 ng/ml), PHA and FK-506 (1 ng/ml), and without stimulation(negative control). The amount of cytokines such as IL-2, IL-3, IL-4 and sIL-2R were measured using an enzyme-linked immunosorbent assay(ELISA).

**Results:** CsA and FK-506 inhibited significantly the production of IL-2, IL-3, IL-4 in PHA-stimulated lymphocytes of both the AD patients and the control groups. FK-506(1 ng/ml) inhibited cytokines production more significantly than CsA(10 ng/ml). However, CsA and FK-506 did not significantly inhibit the production of sIL-2R. There were no significant differences in the inhibitory effect of CsA and FK-506 on IL-4 production upon PHA-stimulation between AD patients with a high IgE level and with a low IgE level.

**Conclusion:** Both FK-506 and CsA inhibit lymphokine production, but not the production of sIL-2R. FK-506 inhibited more significantly lymphokine production at a 10-fold lower concentration than CsA. Our data suggest that FK-506 could be more effective in the treatment of severe AD than CsA, and both these agents show their immunosuppressive activity through the suppression of lymphokine production, not via the suppression of sIL-2R production in AD. (Ann Dermatol 8:(2)98~106, 1996).

---

*Key Words :* Atopic dermatitis, Cyclosporin A, Cytokines, FK-506

Atopic dermatitis(AD) is a chronically relapsing skin disorder that is often associated with allergic rhinitis, asthma, or a family history of atopic disease<sup>1</sup>. The pathogenesis has not been defined,

but a variety of immunological parameters have been found to be altered in AD<sup>2</sup>.

High levels of serum IgE to environmental and food allergens have been demonstrated in most AD patients<sup>1,3,4</sup>. Recently, interleukin 4(IL-4) has been shown to induce enormously the production of IgE, suggesting the possible involvement of IL-4 in the pathogenesis of AD<sup>5,6</sup>. Also, it has been shown that patients with AD are characterized by an impaired capacity of their T cells to release in-

---

Received . October 31, 1995.

Accepted for publication January 15, 1995.

**Reprint request to :** This article was presented at the 47th Annual meeting of the Korean Dermatologic Association on October 14, 1995.

terleukin 2(IL-2) *in vitro*<sup>7</sup>. Also, elevated serum levels of soluble IL-2 receptor(sIL-2R) have been shown in AD patients<sup>7,8,9</sup>. Atopic asthma has been shown to be associated with activation of the IL-3, IL-4, and IL-5 in the bronchi<sup>10</sup>.

The immunosuppressive agents cyclosporin A (CsA) and FK-506 block early events in T lymphocyte activation<sup>11,12</sup>. Although both are fungal metabolites, CsA is a cyclic undecapeptide and FK-506 is a macrolide. Both these agents bind to distinct families of intracellular proteins (immunophilins) termed cyclophilins and FK-506-binding proteins (FKBPs). Then, both CsA-cyclophilins and FK-506-FKBPs complexes bind to calcineurin (phosphatase 2B) and inhibit its activity. Therefore these interfere with the production of IL-2 and other lymphokines<sup>13,14,15</sup>. Compared with CsA, FK-506 is 10- to 100-fold more potent and the two agents can act synergistically both *in vivo* and *in vitro*<sup>11,12,14</sup>.

In this study, we compared the effects of CsA and FK-506 on the expression of IL-2, IL-3, IL-4, and sIL-2R, respectively. Also, we compared the effects of these agents on the expression of IL-4 between AD patients with high and low serum IgE levels.

## MATERIALS AND METHODS

### Blood samples

A total of 32 peripheral blood samples from 17 patients with severe AD and 15 control groups were tested. There were 8 male and 9 female patients, and the mean age was 10.6 (range 2-25) years. All patients fulfilled the diagnostic criteria for severe atopic dermatitis<sup>16</sup>, and systemic steroid therapy had been discontinued 2 months before and oral anti-histamine or topical steroid therapy 3 days before the start of the study.

We also evaluated the serum IgE level of AD patients and control groups with an enzyme-linked immunosorbent assay (ELISA).

### Reagents

CsA and FK-506 were obtained from Hanmi Pharmaceutical, Seoul, Korea and Fujisawa Pharmaceutical Co. Ltd., Osaka, Japan, respectively. Both drugs were dissolved to 1 mM in ethanol and added to the culture medium.

### Peripheral blood lymphocytes isolation

Lymphocytes were isolated by density centrifugation using fresh buffy coats from blood samples from the patients and control groups. Isolated lymphocytes were washed three times with RPMI-1640 and were adjusted to  $2 \times 10^6$  cells/ml with RPMI-1640.

### Medium

RPMI-1640, fetal calf serum (FCS) was purchased from Gibco (Grand Island, NY, U.S.A.). The complete medium comprised of RPMI-1640 supplemented with 10% heat-inactivated FCS, 2 mM L-glutamine, 50 U/mL penicillin and 50 ug/mL streptomycin.

### Lymphocyte culture

The cells were cultured in the complete medium in combination with phytohemagglutinin (PHA) (2% in total medium) and CsA (10 ng/ml) or FK-506 (1 ng/ml). As a positive control, the cells were cultured in the complete medium in combination with PHA (2% in total medium) (Gibco). As a negative control, the cells were cultured in the complete medium alone.

### Cytokines assays

The amount of cytokines such as IL-2, IL-3, IL-4, and sIL-2R were measured using an ELISA<sup>7,9</sup>. Briefly, 96 wells were precoated with a murine IgG monoclonal antibody (mAb) directed against human cytokines (IL-2, IL-3, IL-4: R&D Systems, Inc., Minneapolis, U.S.A., sIL-2R: Immunotech, Marseille, France).

At first, samples were added in the precoated wells and incubated for 2 hours at room temperature. Then each cytokine conjugate (polyclonal antibody against each cytokine, conjugated to enzyme) was added and incubated 2 hours at room temperature. Finally substrate solution was added and after 30 minutes, we determined the optical density of each well using a spectrophotometer set to 450 nm (sIL-2R: 405 nm). The inhibitory effects on the cytokine production of CsA or FK-506 were compared with the positive controls without both drugs.

### Statistical analyses

The results were expressed as mean values  $\pm$  SD. The statistical significance of differences was de-

terminated by the Student's Paired t-Test and the Standard Two-Sample t-Test.

## RESULTS

### 1. The effects of CsA or FK-506 on PHA-induced IL-2, IL-3, IL-4 and sIL-2R production (Table 1)(Fig. 1. A-D)

The amounts of IL-2, IL-3 and IL-4 production in PHA-stimulated lymphocytes added CsA(10 ng/ml) in vitro from 17 patients with AD were significantly decreased in comparison with the positive controls( $p < 0.005$ ). Also, the amounts of IL-2, IL-3 and IL-4 in PHA-stimulated lymphocytes added FK-506(1 ng/ml) were significantly decreased in comparison with the positive controls( $p < 0.001$ ). Furthermore, the production of IL-2, IL-3 and IL-4 in cultures with added FK-506 were more significantly decreased than the production of IL-2, IL-3 and IL-4 in cultures with added CsA.

There was no significant difference in sIL-2R production between the cultures with added CsA or FK-506 and the positive controls without both drugs( $p > 0.05$ ).

Our results from the 15 control group without AD showed a similar effect of CsA and FK-506 on PHA-induced IL-2, IL-3, IL-4 and sIL-2R production in comparison with AD patients.

### 2. The effects of CsA or FK-506 on PHA-induced IL-4 production between high IgE and low IgE (Table 2)(Fig. 2, A-B)

There was no significant difference in the inhibitory effect of CsA on IL-4 production of lymphocytes upon stimulation with PHA between AD patients with a high IgE level (more than 400 Iu/ml, n=9) and with a low IgE level (less than 400 Iu/ml, n=6) ( $p > 0.05$ ). Also, there was no significant difference in the inhibitory effect of FK-506 on IL-4 production between AD patients with a high IgE level and with a low IgE level ( $p > 0.05$ ).

phocytes upon stimulation with PHA between AD patients with a high IgE level (more than 400 Iu/ml, n=9) and with a low IgE level (less than 400 Iu/ml, n=6) ( $p > 0.05$ ). Also, there was no significant difference in the inhibitory effect of FK-506 on IL-4 production between AD patients with a high IgE level and with a low IgE level ( $p > 0.05$ ).

## DISCUSSION

The pathogenesis of AD is still undefined<sup>2</sup>. The prominent immunologic abnormalities associated with AD are defective cell-mediated immunity and increased IgE production<sup>1,4</sup>. Defective cell-mediated immunity (CMI) is manifested by an increased susceptibility to severe skin infections with viruses and to chronic dermatophyte infections; a reduced capacity to manifest delayed-type hypersensitivity and a low PHA-stimulated lymphocyte transformation<sup>1,3,4</sup>. Several investigators have reported that AD patients have a decreased proportion of circulating CD3+ T cells and CD8+ suppressor T cells and, therefore, a selective increase in the ratio of CD4+ cells to CD8+ cells<sup>1,2</sup>.

In mice, CD4+ T cells have been classified: TH1 cells, which produce IL-2, IFN- $\gamma$ , TNF- $\beta$ ; TH2 cells, which produce IL-4, IL-5, IL-6, and IL-10<sup>17</sup>. Several investigators have reported that allergen-specific T cell clones from AD patients produced increased IL-4 and IL-5 in combination with no or low levels of IFN- $\gamma$ <sup>18,19</sup>. Also, it has been seen that atopic asthma is associated with activation of the IL-3, IL-4 and IL-5 in the bronchi<sup>10</sup>. Therefore, the predominant CD4 subset in AD is a TH2-like subset but not TH1-like subset that secrete IF- $\gamma$ . IL-4 is a potent inducer of IgE production, and IFN- $\gamma$  inhibits IL-4-induced

**Table 1.** Comparison of PHA-induced IL-2, IL-3, IL-4 and sIL-2R production between positive controls and lymphocytes added CsA or FK-506 in 17 patients with AD (Mean  $\pm$  SD)

	Pos. C. <sup>§</sup>	CsA	FK-506
IL-2(pg/ml)	999.67 $\pm$ 192.88	438.40 $\pm$ 135.12*	195.03 $\pm$ 115.96**
IL-3(pg/ml)	1349.07 $\pm$ 211.81	1161.39 $\pm$ 183.81*	688.53 $\pm$ 86.75**
IL-4(pg/ml)	140.46 $\pm$ 32.15	107.53 $\pm$ 25.30*	72.03 $\pm$ 16.97**
sIL-2R(pM)	226.88 $\pm$ 43.73	207.06 $\pm$ 37.07	230.06 $\pm$ 38.78

§: positive controls indicate PHA-stimulated lymphocytes.

\*, \*\*: statistically significant in difference from positive controls (Student's Paired t-Test, \* $p < 0.005$ , \*\* $p < 0.001$ )

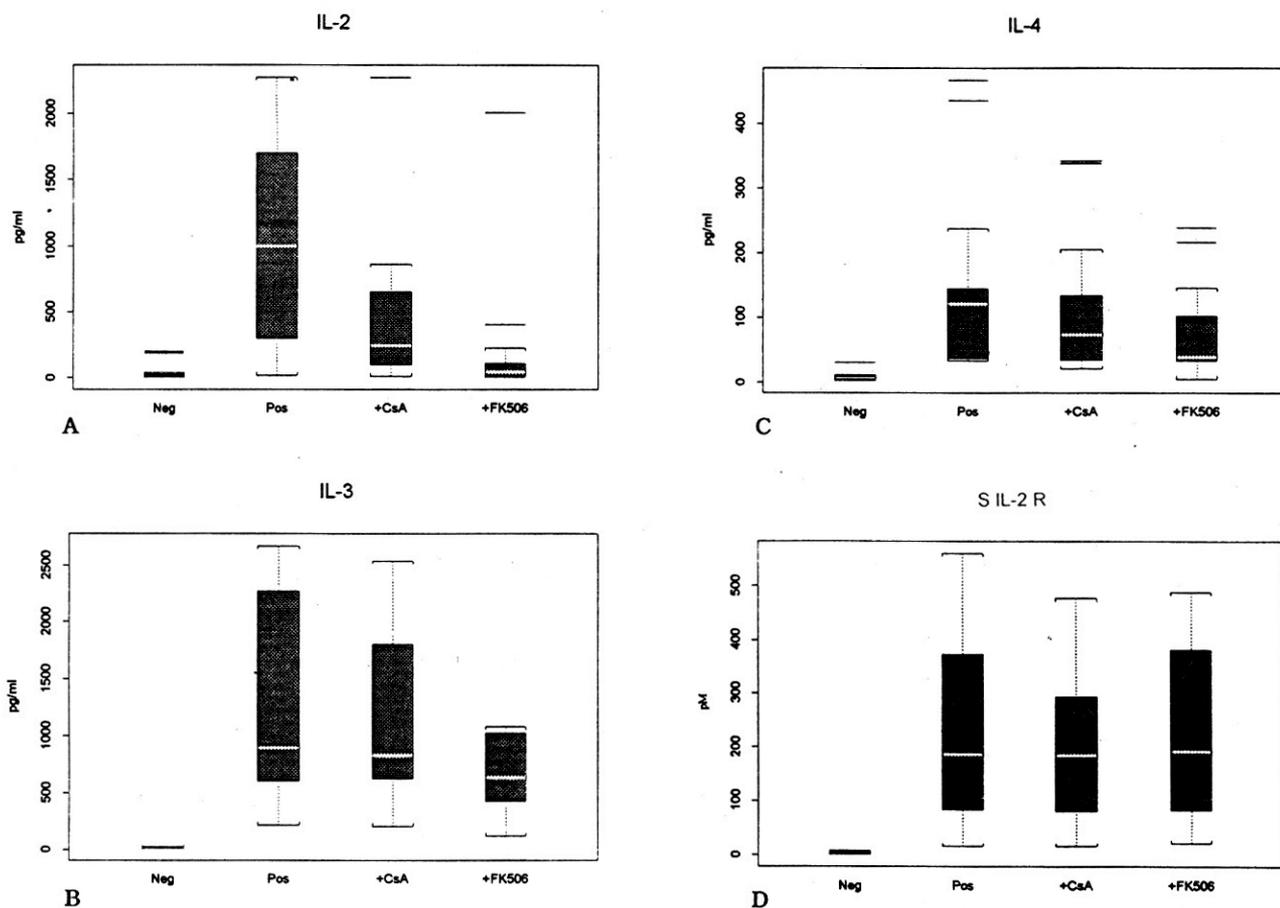


Fig. 1. Comparison of PHA-induced IL-2(A), IL-3(B), IL-4 and sIL-2R(D) production between positive controls and lymphocytes added CsA(10 ng/ml) or FK-506(1 ng/ml) in 17 patients with AD. Neg(negative controls) indicate non-stimulated lymphocytes. Pos(positive controls) indicate PHA-stimulated lymphocytes. Statistical evaluations were determined by Student's Paired t-Test (\*\* $p < 0.005$ , \*\* $p < 0.001$ , significant).

IgE synthesis in vitro. Therefore, the increased IgE production in AD may be influenced not only by IL-4 production but also by the simultaneous suppression of  $IFN-\gamma$ <sup>4,17</sup>. AD patients release less IL-1, TNF and IL-2, possibly due to the selective inhibitory effect on CD4+ T cells of prostaglandin

E overproduction by AD monocytes<sup>7</sup>.

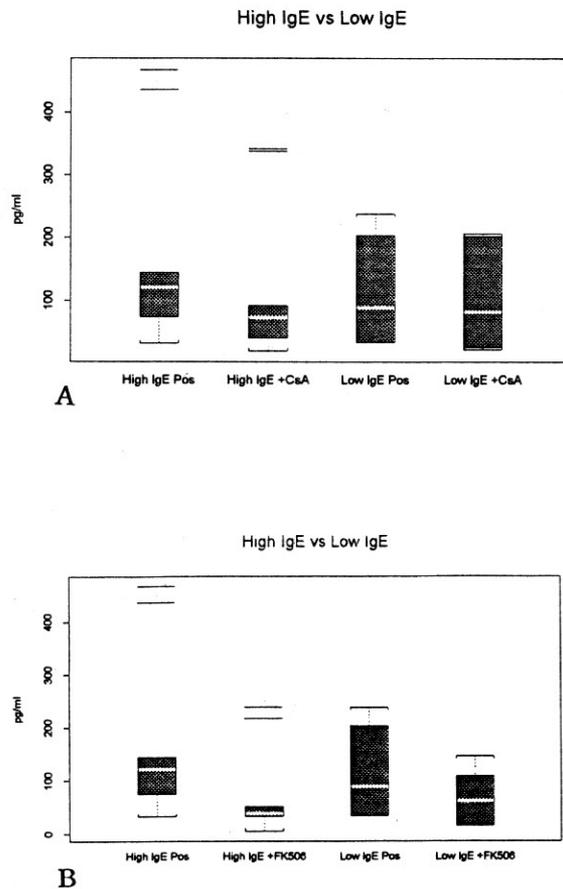
CsA, a cyclic undecapeptide, has been found to be effective in various dermatoses, including AD<sup>20-23</sup>. However, its use is restricted to severely affected patients because of drug-induced hypertension and nephrotoxicity<sup>23</sup>. FK-506, a macrolide antibi-

Table 2. The effects of CsA or FK-506 on IL-4 production between a high serum IgE(n=9) and low serum IgE(n=6)(Mean  $\pm$  SD)

	Pos. C.	CsA	FK-506
High IgE <sup>a</sup>	114.58 $\pm$ 37.72	103.37 $\pm$ 36.05 <sup>+</sup>	68.04 $\pm$ 23.41 <sup>+</sup>
Low IgE <sup>b</sup>	169.55 $\pm$ 54.91	119.99 $\pm$ 42.28	76.86 $\pm$ 28.92

a: more than 400 IU/ml, b: less than 400 IU/ml

+ : statistically not significant in difference between AD patients with a high serum IgE level and low serum IgE level (Standard Two-Sample t-Test, \* $P > 0.05$ )



**Fig. 2.** The effects of CsA(A) or FK-506(B) on PHA-induced IL-4 production between AD patients with a high serum IgE level (more than 400 IU/ml, n=9) and with a low serum IgE level (less than 400 IU/ml, n=6). Statistical evaluations were determined by Standard Two-Sample t-Test ( $p > 0.05$ , not significant).

otic, has been shown to be 10- to 100-fold more potent than CsA in its inhibitory action on T lymphocyte responses<sup>11,12,14</sup>. The immunosuppressive agents CsA and FK-506 have been shown to interfere with the transcriptional activation of early T cell activation genes such as IL-2, IL-3 and IL-4 in activated human peripheral blood T cells<sup>12,24-30</sup>.

In this study, we investigated the effects of both drugs on the expression of cytokines such as IL-2, IL-3, IL-4 and sIL-2R in vitro in patients with AD. We showed that CsA and FK-506 inhibited significantly the production of each IL-2, IL-3, IL-4 in PHA stimulated lymphocytes in vitro in patients with AD. Furthermore, we showed that FK-

506 (1 ng/ml) inhibited more significantly cytokines production at a 10-fold lower concentration than CsA (10 ng/ml). Our results suggested that CsA and FK-506 selectively suppress the activation of CD4+ cells by inhibiting the expression of early T cell - activating genes encoding the cytokines such as IL-2, IL-3 and IL-4. Because AD patients appear to have decreased numbers of CD8+ T cells, the use of both drugs that suppress the activation of CD4+ cells may restore the T cell functional balance in AD and may result in a beneficial clinical effect<sup>22</sup>. Increased serum levels of sIL-2R have been shown in AD patients and has been shown a significant correlation with IgE levels and body surface involvement<sup>7,8,9</sup>. As well as being an early sign of T cell activation, sIL-2R may have an immunoregulatory role with inhibitory effects on T cell activation by binding IL-2<sup>9</sup>. Increased sIL-2R levels may reflect activation of immunopathogenic mechanisms contributing to the exacerbation of AD. Therefore, sIL-2R levels may be a helpful tool for assessing responses to new treatments. We showed that CsA and FK-506 could not significantly inhibit sIL-2R production in PHA-stimulated lymphocytes in vitro in AD patients. This result may indicate that sIL-2R production in PHA-stimulated lymphocytes is probably via post-transcriptional activation pathway<sup>24,31,32</sup>, as has been reported for CD28-activated cells<sup>33</sup>.

As IL-4 has been shown to induce enormously the production of IgE in the pathogenesis of AD<sup>5,6</sup>, we wanted to investigate the differences in the effects of CsA and FK-506 on IL-4 production between a high IgE and a low IgE group in AD patients. In our study, CsA and FK-506 significantly inhibited IL-4 production in PHA-stimulated lymphocytes as reported in recent studies<sup>26-28</sup>. However, there was no significant difference in the inhibitory effect of both drugs on IL-4 production between AD patients with a high IgE level and with a low IgE level. These results suggest that the extent of the inhibitory effect of both drugs on IL-4 production is not related to the serum IgE level.

There has been controversy over the mean IgE values of AD patients in Korea as reported with 293.4 IU/ml by Cheon<sup>34</sup> or 185 IU/ml by Kang<sup>35</sup>. In this study, we selected clinically severe AD patients and their serum IgE levels were relatively

high. Therefore, serum IgE levels were classified into a high and low groups on the basis of 400 IU/ml.

CsA and FK-506 could be suspected to affect IgE production through interfering IL-4 production in AD patients. Therefore, in this study, we wanted to investigate the effects of CsA and FK-506 on the expression of IgE level in supernatant with a paper radioimmunosorbent test (PRIST) method. A minimal unit of IgE measure with a PRIST method is 5 IU and most of the amount of IgE isolated from supernatant in this study were below 0.25 IU. Therefore, we could not evaluate the effect of CsA and FK-506 on the expression of IgE level in vitro. More sensitive IgE measuring techniques such as an overnight PRIST using RAST anti-IgE trace method than the usual PRIST will be necessary for the evaluation of IgE level in supernatant.

CsA has been used for the prevention and treatment of organ transplant rejection, including the kidney, liver, and heart<sup>36,37,38</sup>. Also, it has been shown to reduce the severity of graft-versus-host reactions in patients undergoing bone marrow transplants<sup>39</sup>. Recently, its reports have shown beneficial results in autoimmune medical disorders including ulcerative colitis<sup>40</sup>, Crohn's disease<sup>41</sup>, primary biliary cirrhosis<sup>42</sup>, uveitis<sup>43</sup>, pulmonary sarcoidosis<sup>44</sup>, type I diabetes mellitus<sup>45</sup>, aplastic anemia<sup>46</sup>, myasthenia gravis<sup>47</sup>, Goodpasture's syndrome<sup>47</sup>, rheumatoid arthritis<sup>47</sup>, multiple sclerosis<sup>48</sup>, Grave's disease<sup>49</sup>. Also, it has shown a potential usefulness for dermatologic conditions of presumed autoimmune T cell mediated pathogenesis. These include: psoriasis<sup>50,51</sup>, pemphigus and pemphigoid<sup>52</sup>, dermatomyositis and polymyositis<sup>53,54</sup>, systemic lupus erythematosus<sup>55</sup>, cutaneous T cell lymphoma<sup>56</sup>, Behçet's disease<sup>57</sup>, alopecia<sup>58</sup>, ichthyosis<sup>59</sup>, atopic dermatitis<sup>20,21,22</sup>.

FK-506 is a new immunosuppressive agent that is more potent than CsA and can act synergistically with CsA<sup>11,12,14</sup>. It has been reported that FK-506 prevented allograft rejection in rats<sup>60</sup>, dogs<sup>61</sup>, monkeys, and baboons<sup>62</sup>. In humans, the immunosuppressive activity of FK-506 for liver, kidney, and pancreas transplantation has been reported<sup>63</sup>. Also, it has been reported that FK-506 was effective in treating patients with severe recalcitrant psoriasis<sup>64</sup>. Topical FK-506 has recently been shown to have a suppressive effect of contact der-

matitis in the guinea pig<sup>65</sup> and to have a therapeutic effect in AD patients<sup>66</sup>. Therefore, it may have potential clinical applications in human skin diseases.

In conclusion, we demonstrated that FK-506 more significantly inhibited cytokines such as IL-2, IL-3 and IL-4 production in vitro than CsA in AD patients. Furthermore, the inhibition of IL-4 production by FK-506 suggests that FK-506 can inhibit IgE production. Our data suggest that FK-506 could be more effective in the treatment of severe AD than CsA, and both these agents show their immunosuppressive activity through the suppression of lymphokine production, not via the suppression of sIL-2R production in AD.

## REFERENCES

1. Leung DYM, Rhodes AR, Geha RS: Atopic Dermatitis. In Fitzpatrick TB, Eisen AZ, Wolff-K, Freedberg IM, Austen KF(eds): Dermatology in General Medicine. McGraw-Hill Book Company, New York, 1993, pp1543-1564.
2. Cooper K: Mechanisms of atopic dermatitis. In Norris DA(ed): Immune Mechanisms in Cutaneous Disease. Marcel Dekker, Inc., New York, 1989, pp247-276.
3. Hanifin JM: Atopic dermatitis. *J Am Acad Dermatol* 6:1-13, 1982.
4. Hanifin JM: Immunologic aspects of atopic dermatitis. *Derm Clinics* 8:747-750, 1990.
5. Future M, Ohtsuki M, Ogata F, Ishibashi Y: Responsiveness to interleukin 4 and interleukin 2 of peripheral blood mononuclear cells in atopic dermatitis. *J Invest Dermatol* 96:468-472, 1991.
6. Renz H, Jugo K, Bradley KL, Domenico J, Gelfand EW, Leung DY: Enhanced IL-4 production and IL-4 receptor expression in atopic dermatitis and their modulation by interferon-gamma. *J Invest Dermatol* 99:403-408, 1992.
7. Kapp A, Neuner P, Krutmann J, Luger TA, Schopf E: Production of interleukin-2 by mononuclear cells in vitro in patients with atopic dermatitis and psoriasis. Comparison with serum interleukin-2 receptor levels. *Acta Derm Venereol(Stockh)* 71:403-406, 1991.
8. Czech W, Krutmann J, Schopf E, Kapp A: Serum eosinophil cationic protein (ECP) is a sensitive measure for disease activity in atopic

- dermatitis. *Br J Dermatol* 126:351-355, 1992.
9. Colver GB, Symons JA, Duff GW: Soluble interleukin 2 receptor in atopic eczema. *Br Med J* 298:1426-1428, 1989.
  10. Robinson DS *et al*: Predominant TH2-like bronchoalveolar T-lymphocyte population on atopic asthma. *N Engl J Med* 326:298-304, 1991.
  11. Wiederrecht G, Lam E, Hung S, Martin M, Sigal N: The mechanism of action of FK-506 and cyclosporin A. *Ann N Y Acad Sci* 696:9-19, 1993.
  12. Thomson AW: FK-506 - How much potential? *Immunol Today* 10:6-9, 1989.
  13. Fruman DA, Klee CB, Bierer BE, Burakoff SJ: Calcineurin phosphatase activity in T lymphocytes is inhibited by FK 506 and cyclosporin A. *Proc Natl Acad Sci USA* 89:3686-3690, 1992.
  14. Bishop DK, Wenhua LI: Cyclosporin A and FK 506 mediate differential effects on T cell activation in vivo. *J Immunol* 148:1049-1054, 1992.
  15. Schreiber SL, Crabtree GR: The mechanism of action of cyclosporin A and FK 506. *Immunol Today* 13:136-142, 1992.
  16. Hanifin JM, Lobitz WC: Newer concepts of atopic dermatitis. *Arch Dermatol* 113:663-670, 1977.
  17. Cooper KD: Atopic dermatitis: recent trends in pathogenesis and therapy. *J Invest Dermatol* 102:128-137, 1994.
  18. Van der Heijden FL *et al*: High frequency of IL-4-producing CD4<sup>+</sup> allergen-specific T lymphocytes in atopic dermatitis lesional skin. *J Invest Dermatol* 97:389-394, 1991.
  19. Parronchi P *et al*: Allergen- and bacterial antigen-specific T cell clones established from atopic donors show a different profile of cytokine production. *Proc Natl Acad Sci USA* 88:4538-4542, 1991.
  20. van joost T, Stolz E, Heule F: Efficacy of low-dose cyclosporine in severe atopic skin disease [Letter]. *Arch Dermatol* 123:166-167, 1987.
  21. Munro CS, Higgins EM, Marks JM, Daly BM, Friedmann PS, Shuster S: Cyclosporin A in atopic dermatitis: therapeutic response is dissociated from effects on allergic reactions. *Br J Dermatol* 124:43-48, 1991.
  22. Taylor III RS, Cooper KD, Headington JT, Ho VC, Ellis CN, Voorhees JJ: Cyclosporine therapy for severe atopic dermatitis. *J Am Acad Dermatol* 21:580-583, 1989.
  23. Duncan JI: Differential inhibition of cutaneous T-cell-mediated reactions and epidermal cell proliferation by cyclosporin A, FK-506, and Rapamycin. *J Invest Dermatol* 102:84-88, 1994.
  24. Tocci MJ, Matkovich D, Collier K, Kwok P, Dumont F, Lin S, Degubicibus S, Siekierka JJ, Chin J, Hutchinson N: The immunosuppressant FK 506 selectively inhibits expression of early T cell activation genes. *J Immunol* 143:718-726, 1989.
  25. Kita H, Ohnishi T, Okubo Y, Weiler D, Abrams JS, Gleich GJ: Granulocyte/macrophage colony-stimulating factor and interleukin 3 release from human peripheral blood eosinophils and neutrophils. *J Exp Med* 174:745-748, 1991.
  26. Todd MD, Grusby MJ, Lederer JA, Lacy E, Lichtman AH, Glimcher LH: Transcription of the interleukin 4 gene is regulated by multiple promoter elements. *J Exp Med* 177:1663-1674, 1993.
  27. Yefenof E, Abboud G, Epszteyn S, Vitetta ES: Treatment of premalignancy: Prevention of lymphoma in radiation leukemia virus-inoculated mice by cyclosporin A and immunotoxin. *Proc Natl Acad Sci USA* 89:728-732, 1992.
  28. Motta I, Colle J-H, Shidani B, Truffa-Bachi P: Interleukin 2/interleukin 4-independent T helper cell generation during an in vitro antigenic stimulation of mouse spleen cells in the presence of cyclosporin A. *Eur J Immunol* 21:551-557, 1991.
  29. Waldmann TA: The IL-2/IL-2 receptor system: a target for rational immune intervention. *Immunol Today* 14:264-270, 1993.
  30. Sawada S, Suzuki G, Kawase Y, Takaku F: Novel immunosuppressive agent, FK 506: In vitro effects on the cloned T cell activation. *J Immunol* 139:1797-1803, 1987.
  31. Natazwka T, Umemiya-Okada T, Matsui T, Saida T, Nakao Y: FK 506 and cyclosporin A regulate proliferation and proto-oncogene expression in HTLV-1-associated myelopathy/Tropical-spastic-paraparesis-derived T cells. *Int J Cancer* 54:348-354, 1993.

32. Woo J, Sewell HF, Thomson AW: The influence of FK-506 and low-concentration cyclosporin on human lymphocyte activation antigen expression and blastogenesis: a flow cytometric analysis. *Scand J Immunol* 31:297-304, 1990.
33. June C, Ledbetter J, Gillespie M & Thompson C: T-cell proliferation involving the CD28 pathway is associated with cyclosporine-resistant interleukin 2 gene expression. *Mol Cell Biol* 7:4472-4481, 1987.
34. Cheon HW, Cho CK, Lee SN: IgE in atopic dermatitis. *Kor J Dermatol* 19:847-851, 1981.
35. Kang SB, Lee SB, Kim JW, Kim JK, Kim CW: Total serum IgE level in each age group of patients with atopic dermatitis. *Kor J Dermatol* 26:507-512, 1988.
36. Oyer PE, Stinson EB, Jamieson SW, et al: Cyclosporine in cardiac transplantation: A 2½ year follow-up. *Transplant Proc* 15:2546-2552, 1983.
37. Starzel TE, Inatsuki DH, Van Thiel JC, et al: Report of Colorado-Pittsburgh liver transplantation studies. *Transplant Proc* 15:2582-2585, 1983.
38. European Multicenter Trial Group: Cyclosporine in cadaveric renal transplantation. *Lancet* 1:986-989, 1983.
39. Harper JI, Kendra JR, Desai S, et al: Dermatological aspects of the use of cyclosporin A for prophylaxis of graft-versus-host disease. *Br J Dermatol* 110:469-474, 1984.
40. Gupta S, Keshavarzian A, Hodgson HJF: Cyclosporine in ulcerative colitis. *Lancet* 2:1277-1278, 1984.
41. Allison MC, Pounder RE: Cyclosporin for Crohn's disease. *Lancet* 1:902-903, 1984.
42. Routhier G, Epstein O, Janossy G, et al: Effects of cyclosporine A on suppressor and induced T lymphocytes in primary biliary cirrhosis. *Lancet* 2:1223-1226, 1980.
43. Nussenblatt RB, Palestine AG, Rook AH, et al: Treatment of intraocular inflammatory disease with cyclosporin A. *Lancet* 2:235-238, 1983.
44. Rebuck AS, Stiller CR, Braude AC, et al: Cyclosporin for pulmonary sarcoidosis. *Lancet* 1:1174, 1984.
45. Assan R, Feutren G, Debray-Sachs M, et al: Metabolic and immunologic effects of cyclosporine in recently diagnosed type I diabetes mellitus. *Lancet* 1:67-71, 1985.
46. Wisloff F, Godal HC: Cyclosporine in refractory severe aplastic anemia. *N Engl J Med* 312:1193, 1985.
47. Biren CA, Barr RJ: Dermatologic applications of cyclosporine. *Arch Dermatol* 122:1028-1032, 1986.
48. Mertin J, Knight SC, Rudge P, et al: Double blind controlled trial of immunosuppression in treatment of multiple sclerosis. *Lancet* 2:949-951, 1980.
49. McGregor AM, Bech L, Hall R: Cyclosporin A in management of Grave's disease. *J R Soc Med* 78:511-512, 1985.
50. Harper JI, Keat ACS, Staughton RCD: Cyclosporin for psoriasis. *Lancet* 2:981-982, 1984.
51. Van Hooff JP, Leunissen RML, Staak WVD: Cyclosporine and psoriasis. *Lancet* 1:335, 1985.
52. Thivolet J, Barthelmy H, Rigot-Mulier G, et al: Effects of cyclosporine on bullous pemphigoid and pemphigus. *Lancet* 1:334-335, 1985.
53. Van der Meer S, Inhof JW, Borleff JC: Cyclosporine for polymyositis. *Annals Rheum Dis* 45:612, 1986.
54. Ejstrup L: Severe dermatomyositis treated with cyclosporine A. *Annals Rheum Dis* 45:612-613, 1986.
55. Isenberg DA, Snaith ML, Marrow WJ, et al: Cyclosporine A for the treatment of systemic lupus erythematosus. *Int J Immunopharmacol* 3:163-169, 1981.
56. Totterman TH, Scheymius A, Killander A, et al: Treatment of therapy-resistant Sezary Syndrome with cyclosporin A: Suppression of pruritus, leukaemic T cell activation markers and tumour mass. *Scand J Haematol* 34:196-203, 1985.
57. Nussenblatt RB, Palestine AG, Chan C, et al: Effectiveness of cyclosporin therapy for Behçet's disease. *Arthritis Rheum* 28:671-679, 1985.
58. Gebhart W, Schmidt JB, Schemper M, et al: Cyclosporin-A-induced hair growth in human renal allograft recipients and alopecia areata. *Arch Dermatol Res* 278:238-240, 1986.
59. Velthuis PJ, Jesserun RFM: Improvement of

- ichthyosis by cyclosporine. *Lancet* 1:335, 1985.
60. Inamura N, Nakahara K, Kino T, Goto T, Aoki H, Yamaguchi I, Kohsaka M & Ochiai T: Prolongation of skin allograft survival in rats by a novel immunosuppressive agent, FK-506. *Transplantation* 45:206, 1988.
  61. Collier DSTJ, Thiru S & Calne RY: Kidney transplantation in the dog receiving FK-506. *Transplant Proc* 19(Suppl. 6):62, 1987.
  62. Todo S, Ueda Y, Demetris JA, Imventarza O, Nalesnik M, Venkataramanan R, Mako wka L & Starzl TE: Immunosuppression of canine, monkey and baboon allografts by FK-506: with special reference to synergism with other drugs and to tolerance induction. *Surgery* 104:239-249, 1988.
  63. Starzl TE, Todo S, Fung J, Demetris AJ, Venkataramman R, Jain A: FK-506 for liver, kidney, and pancreas transplantation. *Lancet* 28:1000-1004, 1989.
  64. Jegasothy BV, Ackerman CD, Todo S, et al: Tacrolimus(FK-506) - A new therapeutic agent for severe recalcitrant psoriasis. *Arch Dermatol* 128:781-785, 1992.
  65. Lauerma AI, Stein BD, Homey B, Lee CH, Bloom E, Maibach HI: Topical FK-506: suppression of allergic and irritant contact dermatitis in the guinea pig. *Arch Dermatol Res* 286:337-340, 1994.
  66. Nakagawa H, Etoh T, Ishibashi Y, higaki Y, Kawashima M, Torii H, Harada S: Tacrolimus ointment for atopic dermatitis. *Lancet* 344: 883, 1994.