

Expression of p63 in Various Hyperproliferative Skin Diseases

Seung Seog Han, M.D.¹, Sung Eun Chang, M.D.¹, Hae Jin Jung², Mi Woo Lee, M.D.¹,
Jee Ho Choi, M.D.¹, Kee Chan Moon, M.D.¹, Jai Kyoung Koh, M.D.¹

¹Department of Dermatology, Asan Medical Center, University of Ulsan College of Medicine, Seoul, Korea, ²Asan Institute for Life Science

The keratinocytes in human epidermis are replaced by a population of stem cells located in the basal layer of the epidermis and one candidate stem cell marker is the transcription factor p63. We studied the expression of p63, immunohistochemically, in various hyperproliferative skin diseases (10 poorly differentiated metastatic squamous cell carcinomas (SCCs), 10 non-metastatic primary cutaneous SCCs, 10 cases of Bowen's diseases, 10 actinic keratosis, and 10 melanomas) and also observed the change of p63 expression in psoriasis after cyclosporine treatment. p63 was normally expressed in basal layer cells. Poorly-differentiated metastatic SCC showed the highest expression in most of the tumor cells, while psoriasis, actinic keratosis, Bowen's disease and primary SCC showed an increased expression in the basal and suprabasal area compared to in normal epidermis. The cyclosporine treatment in psoriasis reduced the expression of p63 to a normal level. This data suggests that p63 expression may influence tumor cell differentiation and proliferation without a direct tumorigenesis effect in epithelial tissue. (Ann Dermatol (Seoul) 18(2) 64~69, 2006)

Key Words: Cutaneous neoplasm, p63, Psoriasis, Stem cell

INTRODUCTION

The keratinocytes in human epidermis are constantly turned over and replaced by a population of stem cells located in the basal layer of the epidermis. One class of candidate stem cell markers is the transcription factor p63. It has been shown that p63 is highly expressed in the basal cells of human epithelial tissue^{1,2}. It is also known that p63 is essential for ectodermal differentiation during embryogenesis³⁻⁵. Studies have shown that p63 is a nuclear transcription factor that triggers keratinocyte differentiation and is downregulated in terminally differentiated cells *in vitro*^{1,6,7}.

p63 may block the apoptosis-inducing activity of

p53 and could help to maintain the proliferative capacity of basal or progenitor cells¹⁻⁴. p63 may also play a role in the regulation of proliferation and differentiation in premalignant and malignant lesions of epithelial origin. So, p63 appears to play an oncogenic role in the development of human cancer.

Herein, we studied expression of p63, immunohistochemically, in various hyperproliferative skin diseases. We also observed the change of p63 expression after cyclosporine treatment and UV irradiation.

MATERIALS AND METHODS

Immunohistochemical staining, according to the streptavidin-biotin-peroxidase technique, using mouse monoclonal antibody 63P03(NeoMarkers, Fremont, USA) raised against all known p63 isoforms, was performed. The expression of p63 was evaluated in epidermal cells and skin appendages by semiquantitative analysis of p63 expression in the various hyperproliferative skin diseases (10 primary

Received January 16, 2006

Accepted for publication August 29, 2006

Reprint request to: Jee Ho Choi, M.D., Department of Dermatology, Asan Medical Center, University of Ulsan College of Medicine, 338-1, Pungnap-2dong, Songpa-gu, Seoul 138-736, Korea. Tel. 82-2-3010-3460, Fax: 82-2-486-7831, E-mail. jhchoy@www.amc.seoul.kr

non-metastatic cutaneous squamous cell carcinomas (SCCs), 10 poorly differentiated metastatic SCCs, 10 cases of Bowen's diseases, 10 actinic keratoses, and 10 basal cell carcinomas (BCCs)). We also evaluated the change of p63 expression in 10 psoriatic skin lesions, before and after cyclosporine treatment. Nuclear staining was considered as specific. The percentage of positive cells (-: 1+ : < 5%, 2+ : 5 ~ 25%, 3+ : 25 ~ 50%, 4+ : > 50%) and staining patterns (negative, basal, suprabasal, diffuse and peripheral) were assessed.

RESULTS

In normal control skin, p63 was expressed (1+ ~ 2+) in the nuclei of epidermal basal and suprabasal cells, in the cells of the germinative hair matrix and the external root sheath of hair follicles, in the myoepithelial cells and basal cells of the sweat glands, and also in the basal cells of the sebaceous glands. All terminally-differentiated cells were negative for p63.

All primary cutaneous SCCs, actinic keratoses and psoriasis before cyclosporine treatment showed 2+

~ 3+ immunoreactivity. BCC, Bowen's disease, and poorly-differentiated metastatic SCC showed 3+ ~ 4+ positivity constantly. Terminally-differentiated squamous cell carcinoma showed less expression of p63 than an undifferentiated one. Terminally-differentiated squamous cell carcinoma also showed a peripheral staining pattern. Malignant melanoma showed only a slightly increased expression (Fig. 1, Table 1).

Psoriatic lesional skin after cyclosporine treatment (12 weeks) was similar to normal control skin. On the other hand, the psoriatic lesion before and after cyclosporine treatment (3 weeks, 6 weeks) showed 2+ ~ 3+ positivity (Fig. 2, Table 2).

DISCUSSION

The p53 gene family includes p53, p63 and p73 genes based on gene sequence homologies. p63 is comprised of at least six different protein isoforms that lead to two fundamentally different roles⁸⁻¹⁰. Three of the p63 isoforms (TAp63) encode proteins with roles similar to p53, including transactivation and induction of apoptosis⁸. The other three isoforms

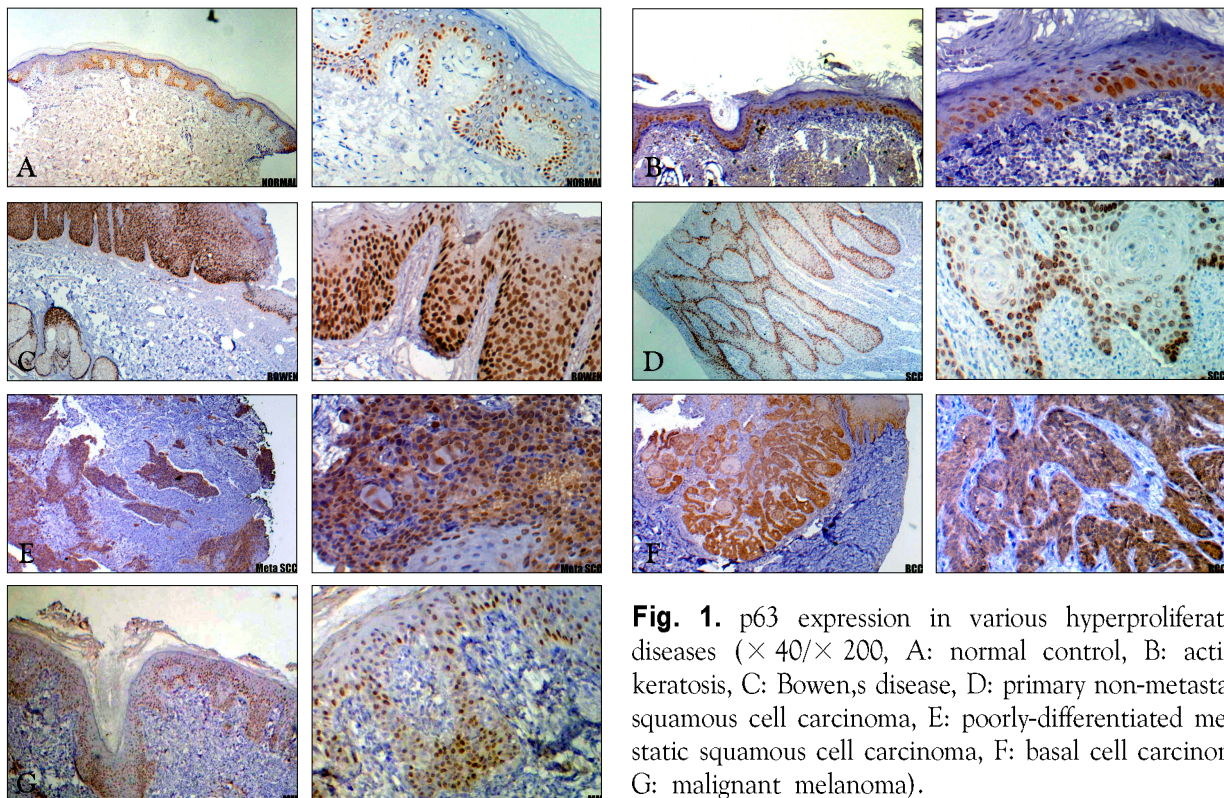
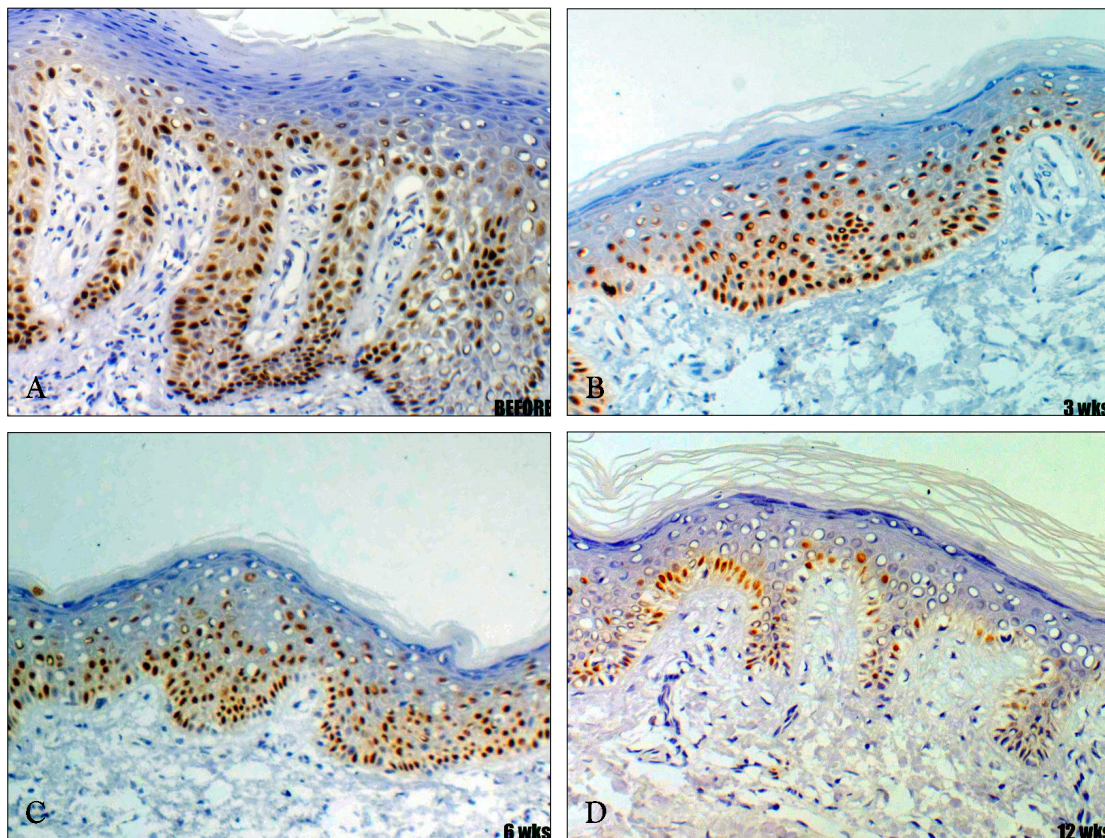


Fig. 1. p63 expression in various hyperproliferative diseases ($\times 40/\times 200$, A: normal control, B: actinic keratosis, C: Bowen's disease, D: primary non-metastatic squamous cell carcinoma, E: poorly-differentiated metastatic squamous cell carcinoma, F: basal cell carcinoma, G: malignant melanoma).

Table 1. p63 Staining Pattern and Degree in Various Hyperproliferative Diseases

	Normal control (N=10)	Actinic keratosis (N=10)	Bowen disease (N=10)	Squamous cell carcinoma (N=10)	Poorly diff. metastatic SCC (N=10)	Basal cell carcinoma (N=10)	Malignant melanoma (N=10)
Staining Pattern	SB, B	SB, B	SB, B or D	B, SB or P	D	D	SB, B
Staining Degree	1+ ~ 2+	3+	3+ ~ 4+	2+ ~ 3+	4+	4+	2+ ~ 3+

SB: Suprabasal, B: Basal, P: Peripheral, D: Diffuse

**Fig. 2.** p63 expression in psoriasis, before and after cyclosporine treatment ($\times 200$, A: before treatment, B: 3 wks after treatment, C: 6 wks after treatment, D: 12 wks after treatment).

($\Delta Np63$) have an inhibitory effects on p53 activity. Therefore, p63 could act either as a tumor suppressor gene, or an oncogene¹¹.

It has been shown that p63 is highly expressed in the basal cells of human epithelial tissue^{1,2}. p63 expression has also been identified in many normal tissues including urothelium, bronchial epithelium

and the myoepithelial layers of breast, prostate and submucosal glands¹²⁻¹⁵. p63 expression has also been established in several neoplasms, including squamous, urothelial, endometrial, and papillary thyroid carcinomas and thymomas¹⁶⁻²³.

It has been demonstrated that p63 is essential for ectodermal differentiation during embryogenesis³⁻⁵.

Table 2. p63 Staining Pattern in Psoriasis, before and after Cyclosporine Treatment

	Psoriasis with cyclosporin treatment			
	Before Tx (N=10)	3 wks (N=10)	6 wks (N=10)	12 wks (N=10)
Staining pattern	SB, B	SB, B	SB, B	B
Staining degree	2+ ~ 3+	2+ ~ 3+	2+	1+

SB: suprabasal, B: basal, P: peripheral, D: diffuse

The expression of p63 in normal human epidermis, cutaneous appendages and skin carcinomas has been assessed and it has been suggested that analysis of p63 expression may help in the differential diagnosis of primary cutaneous tumors compared to metastatic cutaneous tumors^{24,26}. In this study, the expression of p63 in malignant melanoma did not increase, which implied the specificity of p63 with stratified epithelium.

The p63 staining of metastatic SCCs and SCCs was highly increased when compared with that of other non-malignant specimens. A diffuse p63 staining pattern was evident in metastatic SCCs. A suprabasal p63 staining pattern was found in keratinocytes within the entire range of noninvasive lesions. Poorly-differentiated squamous cell carcinoma showed a diffuse increased pattern, while well-differentiated squamous cell carcinoma showed peripheral staining. These finding imply that p63 is involved in keratinocyte differentiation. p63 was constitutively expressed in basal cells, its overexpression in squamous dysplasia and neoplasia may reflect immaturity of the tumor cell lineage, and it was observed that p63 staining was more uniform and homogeneous in less-differentiated tumor areas. Studies have shown that p63 is a nuclear transcription factor that triggers keratinocyte differentiation and is downregulated in terminally differentiated cells *in vitro*^{1,6,23}. p63 expression was mainly noted in the peripheral cells of tumor nests in well-differentiated tumor areas^{27,28}. p63 proteins could be used as an immunohistochemical marker for the differential diagnosis of poorly-differentiated and undifferentiated squamous cell carcinoma²⁷.

p63 expression was greater in metastatic SCCs when compared with that of normal skin and actinic keratosis. A diffuse high level of p63 expression in metastatic SCCs suggested that the degree of the p63-positive stems are correlated with the poor differentiation of malignant SCCs. p63 has been

identified in keratinocyte stem cells.¹ It is thought that stem cells are involved in the formation of malignant tumors³¹. p63 locus, chromosome 3q27-ter is frequently amplified in squamous cell carcinoma of the skin, lungs and esophagus³²⁻³⁴. Therefore, p63 appears to play an oncogenic role in the development of human cancer.

Basal cell carcinomas were shown to have high levels of p63 expression with minimal variability in their staining pattern. This correlates to the report that basal cell carcinomas show less variation in differentiation than squamous cell carcinomas^{24,29}.

In psoriasis, there was an increased expression of p63, but after 12 weeks of cyclosporine treatment, the expression of p63 was found to be normal. Furthermore all malignant lesions, except malignant melanoma, showed an increased expression. This data suggests that p63 expression influences tumor cell differentiation and may be associated with cancer development in epithelial tissue. But p63 positive cells did not always express Ki-67^{24,29}. Thus, p63 positive cells were not undergoing active mitosis. p63 might block the apoptosis-inducing activity of p53 and could help to maintain the proliferative capacity of basal or progenitor cells^{1,2,20,21}. Later stages of wound healing were associated with higher levels of ΔNp63 expression within basal keratinocytes, implying a crucial role in maintaining proliferative potential³⁰.

p63 was normally expressed in basal cells. Poorly-differentiated metastatic SCC showed the highest expression involving most of the tumor cells, while psoriasis, actinic keratosis, Bowen's disease and primary SCC showed slightly increased expression in the basal and suprabasal area in comparison to normal epidermis. The cyclosporine treatment in psoriasis reduced the expression of p63 to a normal level. This data suggests that p63 expression may be associated with differentiation and progression in skin epithelial tumors. High expression of p63 in

benign skin lesions such as psoriasis suggest that in comparison to p53, p63 is not involved in early carcinogenesis of epithelial skin cancer.

REFERENCES

1. Pellegrini G, Dellambra E, Golisano O, Martinelli E, Fantozzi I, Bondanza S, et al. p63 identifies keratinocyte stem cells. *Proc Natl Acad Sci USA* 2001;98:3156-3161.
2. Mills AA, Zheng B, Wang XJ, Vogel H, Roop DR, Bradley A. p63 is a p53 homologue required for limb and epidermal morphogenesis. *Nature* 1999;398:708-713.
3. Yang A, Schweitzer R, Sun D, Kaghad M, Walker N, Bronson RT, et al. p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. *Nature* 1999;398:714-718.
4. Parsa R, Yang A, McKeon F, Green H. Association of p63 with proliferative potential in normal and neoplastic human keratinocytes. *J Invest Dermatol* 1999;113:1099-1105.
5. Celli J, Duijff P, Hamel BC, Bamshad M, Kramer B, Smits AP, et al. Heterozygous germline mutations in the p53 homolog p63 are the cause of EEC syndrome. *Cell* 1999;99:143-153.
6. Levrero M, De Laurenzi V, Costanzo A, Gong J, Wang JY, Melino G. The p53/p63/p73 family of transcription factors: overlapping and distinct functions. *J Cell Sci* 2000;113:1661-1670.
7. Reis-Filho JS, Torio B, Albergaria A, Schmitt FC. p63 expression in normal skin and usual cutaneous carcinomas. *J Cutan Pathol* 2002;29:517-523.
8. Yang A, Kaghad M, Wang Y, Gillett E, Fleming MD, Dotsch V, et al. p63, a p53 homolog at 3q27-29, encodes multiple products with transactivating, death-inducing, and dominant-negative activities. *Mol Cell* 1998;2:305-316.
9. Yang A, McKeon F. p63 and P73: P53 mimics, menaces and more. *Nat Rev Mol Cell Biol* 2000;1:199-207.
10. Yang A, Kaghad M, Caput D, McKeon F. On the shoulders of giants: p63, p73 and the rise of p53. *Trends Genet* 2002;18:90-95.
11. Jost CA, Marin MC, Kaelin WG Jr. p73 is a simian [correction of human] p53-related protein that can induce apoptosis. *Nature* 1997;389:191-194.
12. Di Como CJ, Urist MJ, Babayan I, Drobnjak M, Hedvat CV, Teruya-Feldstein J, et al. p63 expression profiles in human normal and tumor tissues. *Clin Cancer Res* 2002;8:494-501.
13. Signoretti S, Waltregny D, Dilks J, Isaac B, Lin D, Garraway L, et al. p63 is a prostate basal cell marker and is required for prostate development. *Am J Pathol* 2000;157:1769-1775.
14. Barbareschi M, Pecciarini L, Cangi MG, Macri E, Rizzo A, Viale G, et al. p63, a p53 homologue, is a selective nuclear marker of myoepithelial cells of the human breast. *Am J Surg Pathol* 2001;25:1054-1060.
15. Wang BY, Gil J, Kaufman D, Gan L, Kohtz DS, Burstein DE. p63 in pulmonary epithelium, pulmonary squamous neoplasms, and other pulmonary tumors. *Hum Pathol* 2002;33:921-926.
16. Nylander K, Coates PJ, Hall PA. Characterization of the expression pattern of p63 alpha and delta Np63 alpha in benign and malignant oral epithelial lesions. *Int J Cancer* 2000;87:368-372.
17. Quade BJ, Yang A, Wang Y, Sun D, Park J, Sheets EE, et al. Expression of the p53 homologue p63 in early cervical neoplasia. *Gynecol Oncol* 2001;80:24-29.
18. Wang TY, Chen BF, Yang YC, Chen H, Wang Y, Cviko A, et al. Histologic and immunophenotypic classification of cervical carcinomas by expression of the p53 homologue p63: a study of 250 cases. *Hum Pathol* 2001;32:479-486.
19. Glickman JN, Yang A, Shahsafaei A, McKeon F, Odze RD. Expression of p53-related protein p63 in the gastrointestinal tract and in esophageal metaplastic and neoplastic disorders. *Hum Pathol* 2001;32:1157-1165.
20. Park BJ, Lee SJ, Kim JI, Lee SJ, Lee CH, Chang SG, et al. Frequent alteration of p63 expression in human primary bladder carcinomas. *Cancer Res* 2000;60:3370-3374.
21. O'Connell JT, Mutter GL, Cviko A, Nucci M, Quade BJ, Kozakewich HP, et al. Identification of a basal/reserve cell immunophenotype in benign and neoplastic endometrium: a study with the p53 homologue p63. *Gynecol Oncol* 2001;80:30-36.
22. Unger P, Ewart M, Wang BY, Gan L, Kohtz DS, Burstein DE. Expression of p63 in papillary thyroid carcinoma and in Hashimoto's thyroiditis: a pathobiologic link? *Hum Pathol* 2003;34:764-769.
23. Chilosi M, Zamo A, Brighenti A, Malpeli G, Montagna L, Piccoli P, et al. Constitutive expression of DeltaN-p63alpha isoform in human thymus and thymic epithelial tumours. *Virchows Arch*

- 2003;443:175-183.
24. Tsujita-Kyutoku M, Kiuchi K, Danbara N, Yuri T, Senzaki H, Tsubura A. p63 expression in normal human epidermis and epidermal appendages and their tumors. *J Cutan Pathol* 2003;30:11-17.
 25. Qureshi HS, Ormsby AH, Lee MW, Zarbo RJ, Ma CK. The diagnostic utility of p63, CK5/6, CK 7, and CK 20 in distinguishing primary cutaneous adnexal neoplasms from metastatic carcinomas. *J Cutan Pathol* 2004;31:145-152.
 26. Ivan D, Hafeez Diwan A, Prieto VG. Expression of p63 in primary cutaneous adnexal neoplasms and adenocarcinoma metastatic to the skin. *Mod Pathol* 2005;18:137-142.
 27. Chen YK, Hsue SS, Lin LM. Correlation between inducible nitric oxide synthase and p53 expression for DMBA-induced hamster buccal-pouch carcinomas. *Oral Dis* 2003;9:227-234.
 28. Sniezek JC, Matheny KE, Burkey BB, Netterville JL, Pietenpol JA. Expression of p63 and 14-3-3sigma in normal and hyperdifferentiated mucosa of the upper aerodigestive tract. *Otolaryngol Head Neck Surg* 2002;126:598-601.
 29. Wrone DA, Yoo S, Chipps LK, Moy RL. The expression of p63 in actinic keratoses, seborrheic keratosis, and cutaneous squamous cell carcinomas. *Dermatol Surg* 2004;30:1299-1302.
 30. Noszczyk BH, Majewski ST. p63 expression during normal cutaneous wound healing in humans. *Plast Reconstr Surg* 2001;108:1242-1247.
 31. Miller SJ, Wei ZG, Wilson C, Dzubow L, Sun TT, Lavker RM. Mouse skin is particularly susceptible to tumor initiation during early anagen of the hair cycle: possible involvement of hair follicle stem cells. *J Invest Dermatol* 1993;101:591-594.
 32. Bockmuhl U, Schwendel A, Dietel M, Petersen I. Distinct patterns of chromosomal alterations in high- and low-grade head and neck squamous cell carcinomas. *Cancer Res* 1996;56:5325-5329.
 33. Bjorkqvist AM, Husgafvel-Pursiainen K, Anttila S, Karjalainen A, Tammilehto L, Mattson K, et al. DNA gains in 3q occur frequently in squamous cell carcinoma of the lung, but not in adenocarcinoma. *Genes Chromosomes Cancer* 1998;22:79-82.
 34. Taniere P, Martel-Planche G, Saurin JC, Lombard-Bohas C, Berger F, Scoazec JY, et al. TP53 mutations, amplification of p63 and expression of cell cycle proteins in squamous cell carcinoma of the oesophagus from a low incidence area in Western Europe. *Br J Cancer* 2001;85:721-726.