

A Pilot Study of Intralymphatic Immunotherapy for House Dust Mite, Cat, and Dog Allergies

Sang Pyo Lee,¹ Seung Joon Choi,² Eugene Joe,³ Sang Min Lee,^{1*} Min Woo Lee,⁴ Jung Woo Shim,¹ Yu Jin Kim,¹ Sun Young Kyung,¹ Jeong-Woong Park,¹ Sung Hwan Jeong,¹ Joo Hyun Jung⁵

¹Division of Pulmonology and Allergy, Department of Internal Medicine, Gachon University Gil Medical Center, Incheon, Korea

²Department of Radiology, Gachon University Gil Medical Center, Incheon, Korea

³Department of Radiology, Seoul Medical Center, Seoul, Korea

⁴Gachon University Graduate School of Medicine, Incheon, Korea

⁵Department of Otolaryngology-Head and Neck Surgery, Gachon University Gil Medical Center, Incheon, Korea

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Several recent clinical trials reported that intralymphatic immunotherapy (ILIT) for some allergens, such as cat dander and pollen, induce tolerance more rapidly than conventional subcutaneous or sublingual immunotherapy, have a comparable duration of effect after only 3 injections, and do not provoke serious local or systemic reactions. However, the efficacy and safety of ILIT are using *Dermatophagoides farinae* (Df), *Dermatophagoides pteronyssinus* (Dp), and dog, which are indoor allergens that are commonly found globally, need to be evaluated. Furthermore, use of multiple allergens in ILIT should be investigated. We assessed the clinical efficacy and adverse effects of ILIT using aqueous Df, Dp, dog, and cat allergens or mixtures thereof in patients with allergic rhinitis. A total of 11 subjects with AR sensitized to Df, Dp, cat, and/or dog allergens received 3 intralymphatic inguinal injections of sensitized allergen extract (HollisterStier, New Orleans, LA, USA). Clinical parameters were assessed before ILIT, and 4 months and 1 year after the first injection. Rhinitis symptoms were alleviated and quality of life was improved 4 months after ILIT ($P=0.012$ and $P=0.007$, respectively), and these improvements lasted for 1 year after ILIT ($P=0.047$ and $P=0.009$, respectively). However, we observed 2 cases of anaphylaxis, one case of a moderate-to-severe systemic hypersensitivity reaction and the other case of a severe local reaction at the injection site after ILIT. In conclusion, ILIT can rapidly improve allergy symptoms and quality of life, and this effect lasts for 1 year. In hypersensitized patients, however, ILIT can provoke severe systemic and/or local hypersensitivity reactions when performed using aqueous allergen extracts.

Key Words: Allergen-specific immunotherapy; allergic rhinitis; intralymphatic immunotherapy

INTRODUCTION

Intralymphatic immunotherapy (ILIT) was introduced recently as a new modality of allergen-specific immunotherapy (AIT); in this approach, only three intralymphatic injections induce marked clinical improvements as early as 4 months after the day of the first injection, and last for 3 years.¹⁻⁶ Moreover, ILIT causes fewer and milder adverse reactions. However, clinical efficacy has been questioned.⁷ The efficacy and adverse effects of ILIT for *Dermatophagoides farinae* (Df), *Dermatophagoides pteronyssinus* (Dp), and dog allergens, which are indoor allergens common globally, should be investigated. Furthermore, use of multiple allergens in ILIT warrants further investigation.

In this study, we evaluated the clinical efficacy and adverse effects of ILIT using aqueous Df, Dp, dog and cat allergens or mixtures thereof in patients with allergic rhinitis.

MATERIALS AND METHODS

Study population

We enrolled subjects with AR, symptoms of which were provoked by Df, Dp, dog, and/or cat allergens. The subjects met the following enrollment criteria described below. 1) Sensitization proven by skin prick test (SPT) and serum level of allergen-specific IgE measured by ImmunoCAP® (ThermoFisher Scientific, Uppsala, Sweden). Subjects were regarded as being sensi-

Correspondence to: Sang Min Lee, MD, PhD, Division of Pulmonology and Allergy, Department of Internal Medicine, Gachon University Gil Medical Center, 21 Namdong-daero, 774-beon-gil, Namdong-gu, Incheon 21565, Korea. Tel: +82-32-460-2634; Fax: +82-32-469-4320; E-mail: sangminlee77@naver.com
Received: June 23, 2016; Revised: August 22, 2016; Accepted: September 9, 2016

- Sang Pyo Lee and Seung Joon Choi contributed equally to this work.
- There are no financial or other issues that might lead to conflict of interest.

tized to an allergen if, according to the SPT, the allergen/histamine (A/H) ratio in the wheal was ≥ 1 and serum level of allergen-specific IgE was ≥ 0.35 kU/L. 2) Complaints of AR symptoms during exposure to house dust, dog and/or cat in daily life.

Study design

At the first visit, patient eligibility was determined, information about the study was provided to subjects, written consent was obtained from subjects, and rescue medications including oral antihistamine (cetirizine) and nasal glucocorticosteroid spray (ciclesonide) were prescribed (Supplementary Fig. 1). The patients were also asked to administer oral antihistamines with or without a nasal glucocorticosteroid spray, according to the recommendation of the Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines.⁸

At the second visit, pretreatment status was evaluated using questionnaires addressing allergic symptoms, and SPT, intradermal test (IDT), and nasal allergen provocation test (NAPT) results. Questionnaires used to assess allergic symptoms included the Sinonasal Outcome Test-20 (SNOT-20),⁹ and Rhinoconjunctivitis Quality of life Questionnaire (RQLQ).¹⁰ SPTs and IDTs were performed using serial dilutions of extracts of the sensitized allergen Df, Dp, dog, and/or cat (HollisterStier, New Orleans, LA, USA) according to the manufacturer's instructions. In subjects with Df and/or Dp allergy, NAPTs with the sensitized allergen were performed as previously described.¹¹ In addition to AR symptoms during the NAPT, the mean volume (cm^3) of the nasal cavity in the anterior nasal segment (volume 2-6 cm) was measured before the NAPT (baseline test) and every 15 minutes during NAPT by acoustic rhinometry (SRE 2000 Rhinometer; Rhinometrics, Lyngø, Denmark) according to the guidelines of the Standardization Committee on Acoustic Rhinometry.¹² Subjects were asked to stop oral cetirizine (half-life: 8 h) and nasal ciclesonide spray (half-life: 3.5 h) 3 days before the second visit to ensure the validity of the SPT, IDT, and NAPT results.

At visits 3 to 5, the study subjects received three 0.1 mL injections of their sensitized allergen extract at 4-week intervals. Using ultrasound guidance and a 25-gauge needle, aqueous allergen extracts (HollisterStier, New Orleans, USA) were aseptically injected into the superficial inguinal lymph node in the right-side groin.¹⁻⁷ Before the injections, aspiration was performed to avoid inadvertent intravascular administration. After each injection, subjects were closely monitored for 1 h with checking of vital signs at 5-min intervals, and adverse events, if any, were recorded. Adverse events due to previous injections were also checked before the next injection at visits 4 and 5. Large local reactions (swelling > 10 cm in diameter that persisted for > 24 h) were identified, and systemic hypersensitivity reactions were graded using the Mueller classification.^{1,13} At visit 3, venous puncture was performed to measure the serum levels of total IgE, allergen-specific IgE, and allergen-specific immunoglobu-

lin G4 (IgG4), and subjects were asked to rate the pain provoked by intralymphatic injection and that by venous puncture using the visual analog scale (VAS) ranging from 0 to 100 mm.^{1,3} The initial dose of allergen was a 1,000-fold dilution of the maximal concentration of allergen extract for subcutaneous immunotherapy (SCIT) (initial concentration: 30 AU/mL for Df or Dp; 10 AU/mL for cat hair; and 1:1/10 weight/volume (w/v) for dog hair/dander; HollisterStier) in a volume of 0.1 mL. After the first injection, the allergen concentration was escalated 3-fold on the day of the second injection, and 10-fold on the day of the third injection, if there was no or mild local or systemic hypersensitivity reaction. The allergen concentration did not change on the day of second or third injection if there was a moderate local or systemic reaction. The allergen concentration was decreased by 3- to 1,000-fold from the previous concentration if there was a severe local or systemic reaction. When 2 or more allergens were injected into the inguinal lymph node, the allergen mixture was produced in a volume of 0.1 mL to preserve the target concentration of each allergen.

At visits 6 and 7, posttreatment status was evaluated in a manner similar to that at the first visit and blood sampling was performed, respectively. Adverse events after the third injections were also checked at visit 6.

The study was approved by our Institutional Review Board and monitored by our Human Research Protection Committee. This study was registered in an open-access trials registry (ClinicalTrials.gov identifier: NCT02301884).

Statistical analysis

Statistical analysis was performed using PASW 20.0 (SPSS Inc. Chicago, IL, USA). Continuous variables were analyzed by paired Man-Whitney U test, whereas categorical variables were analyzed by Fisher's exact test. A *P* value < 0.05 was deemed to indicate statistical significance.

RESULTS

Informed consent was obtained from a total of 24 subjects; however, 4 did not attend a further visit (Supplementary Fig. 2). Therefore, pre-ILIT evaluation of 20 subjects was carried out, but 5 dropped out due to lack of time to participate in this study, and 2 due to occurrence of anaphylaxis during SPT and IDT with Df and Dp allergens, and 2 due to lack of time after the first injection of ILIT. The post-ILIT evaluation was thus performed in 11 subjects. The demographic characteristics of the subjects are shown in Table 1.

Local and systemic adverse effects during ILIT

The pain of intralymphatic injection was comparable to that of venous puncture (Supplementary Fig. 3). Seven subjects complained of mild local or systemic reaction (grade 0-1 in the Mueller classification); however, four experienced large local re-

Table 1. Demographic characteristics

Subject No.	Gender	Age (year)	Duration of AR (year)	Comorbidities	Past medical history	Current treatment	Allergen
1	Female	47	3	None	None	AH, LA, ICS/LABA	Dog
2	Female	35	7.5	Asthma, food allergy, aspirin idiosyncrasy	Otitis media	AH, ICS	Df, Dp, dog, cat
3	Male	42	2	Asthma	None	AH, LA, ICS/LABA	Df, Dp, dog, cat
4	Male	33	5	None	None	AH, NCS, LA	Df
5	Female	22	20	None	Sinusitis, food allergy	AH, NCS	Df, Dp
6	Female	50	2	Asthma	Otitis media	AH, NCS, ICS/LABA	Df, Dp
7	Female	32	18	Asthma, sinusitis	Nasal polyp, urticaria, food allergy	AH, NCS, LA, ICS/LABA	Df, Dp, cat
8	Male	57	3	None	None	AH, NCS, LA	Dp
9	Female	49	2	Asthma, hypertension	None	AH, NCS, ED, ICS/LABA	Df, Dp
10	Male	45	35	Asthma	Otitis media, food allergy	AH, NCS, ED, ICS/LABA	Df, Dp
11	Female	46	2	Urticaria, food allergy	Atopic dermatitis	AH, NCS, ED	Df, Dp, dog

AR, allergic rhinitis; AH, antihistamine; LA, leukotriene antagonist; ICS, inhaled corticosteroid; LABA, long-acting beta2 agonist; NCS, nasal corticosteroid; ED, antihistamine eye drops; Df, *Dermatophagoides farinae*; Dp, *Dermatophagoides pteronyssinus*.

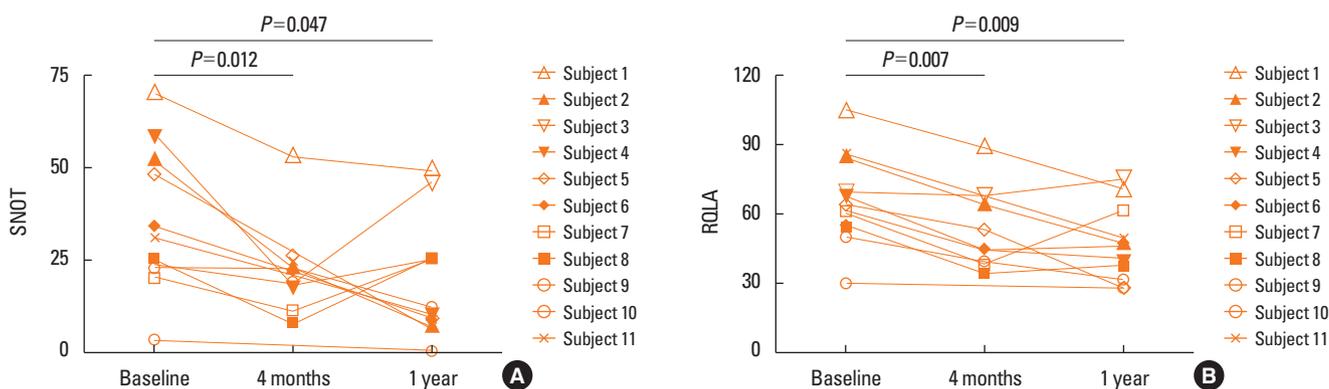


Figure. Subject-reported rhinitis symptoms and quality of life. (A) Sinonasal Outcome Test-20 (SNOT-20) scores. (B) Rhinoconjunctivitis Quality of Life Questionnaire (RQLQ) scores.

actions or moderate-to-severe systemic reactions of grade 2-3 in the Mueller classification (Table 2). Despite those severe reactions, they strongly desired to undergo further ILIT as scheduled, so that additional injections were performed using 3-, 100-, or 1,000-fold dilutions of the concentration previously applied.

AR symptoms, quality of life, and prescription of rescue medication

The SNOT-20 and RQLQ scores were significantly decreased 4 months ($P=0.012$ and $P=0.007$, respectively) and 1 year ($P=0.047$ and $P=0.009$, respectively) after ILIT compared with the baseline levels (Figure).

In general, rescue medications, with the exception of antihistamine eye drops, were prescribed less frequently after ILIT. The frequency of nasal corticosteroid spray prescription was significantly reduced 4 months after the first injection of ILIT ($P=0.04$; data not shown).

Nasal reactivity to house dust mites in NAPT

Nasal symptoms during nasal challenge with house dust mite allergens in NAPT were significantly reduced 1 year after ILIT ($P<0.05$; Supplementary Fig. 4). The decrease in nasal cavity volume during NAPT was also alleviated 1 year after ILIT; however, the magnitude of the decrease was not significant.

Skin reactivity to allergens in the SPT and IDT

Skin reactivity to allergens in the SPT and IDT was generally increased after ILIT, albeit without statistical significance. Skin reactivity to Dp in the SPT was significantly increased 1 year after ILIT ($P<0.05$; data not shown).

Serum total IgE and serum allergen-specific IgE and IgG4

Serum levels of allergen-specific IgE to Df and Dp were significantly increased 4 months after ILIT ($P<0.05$), but they de-

Table 2. Treatment schedule and local/systemic reactions

Subject No.	Injection No.	ILIT dose* (AU/mL or w/v)	Local reaction	Systemic reaction	Mueller classification†
1	1	Dog 1:1/10	None	None	Grade 0
	2	Dog 1:3/10	Heating sensation, edema, itching	None	Grade 0
	3	Dog 1:1	Erythema, itching	None	Grade 0
2	1	Df 30, Dp 30 Dog 1:1/10, Cat 10	None	Generalized itching	Grade 1
	2	Df 100, Dp 100 Dog 1:3/10, Cat 30	Pain, wheal, erythema	None	Grade 0
	3	Df 300, Dp 300 Dog 1:1, Cat 100	Pain, wheal, flare, edema	None	Grade 0
3	1	Df 30, Dp 30 Dog 1:1/10, Cat 10	Edema, itching	Urticaria, generalized itching	Grade 1
	2	Df 30, Dp 30 Dog 1:1/10, Cat 10	Pain, wheal, erythema	Mild headache	Grade 0
	3	Df 100, Dp 100 Dog 1:3/10, Cat 30	Heating sensation, wheal, erythema, edema, itching	None	Grade 0
4	1	Df 30	Pain	None	Grade 0
	2	Df 100	Heating sensation	None	Grade 0
	3	Df 300	Pain	None	Grade 0
5	1	Df 30, Dp 30	None	Anaphylaxis	Grade 3
	2	Df 0.03, Dp 0.03	None	None	Grade 0
	3	Df 0.1, Dp 0.1	None	None	Grade 0
6	1	Df 30, Dp 30	Heating sensation, edema, itching	Delayed-typed dyspnea & wheezing	Grade 0
	2	Df 100, Dp 100	None	Anaphylaxis	Grade 3
	3	Df 1, Dp 1	None	None	Grade 0
7	1	Df 30, Dp 30 Cat 10	Pain	RLQ pain	Grade 0
	2	Df 100, Dp 100 Cat 30	Pain, wheal	RLQ pain	Grade 0
	3	Df 300, Dp 300 Cat 100	Pain	RLQ pain	Grade 0
8	1	Dp 30	None	Diarrhea	Grade 0
	2	Dp 100	None	Fever, chilling, headache	Grade 0
	3	Dp 100	Erythema, itching	None	Grade 0
9	1	Df 30, Dp 30	Myalgia	None	Grade 0
	2	Df 100, Dp 100	Pain	None	Grade 0
	3	Df 300, Dp 300	Pain, erythema	None	Grade 0
10	1	Df 30, Dp 30	None	None	Grade 0
	2	Df 100, Dp 100	Severe edema and erythema with itching and heating sensation	None	Grade 0
	3	Df 30, Dp 30	Severe edema and erythema with itching and heating sensation	None	Grade 0
11	1	Df 30, Dp 30 Dog 1:1/10	None	None	Grade 0
	2	Df 100, Dp 100 Dog 1:3/10	Pain, heating sensation, edema, itching	None	Grade 0
	3	Df 300, Dp 300 Dog 1:1	None	Urticaria, whole body itching, febrile sensation, tightness	Grade 2

*The initial dose of allergen was a 1,000-fold dilution of the maximal concentration of allergen extract for subcutaneous immunotherapy (initial concentration: 30 AU/mL for Df or Dp; 10 AU/mL for cat hair; and 1:1/10 weight/volume (w/v) for dog hair/dander, HollisterStier, New Orleans, USA) in a volume of 0.1 mL. After the first injection, the allergen concentration was escalated 3-fold on the day of the second injection, and 10-fold on the day of the third injection, if there was no or mild local or systemic hypersensitivity reaction. The allergen concentration did not change on the day of second or third injection if there was a moderate local or systemic reaction. The allergen concentration was decreased by 3- to 1,000-fold from the previous concentration if there was a severe local or systemic reaction. †Systemic hypersensitivity reactions were graded using the Mueller classification. ILIT, intralymphatic immunotherapy; AU/mL, allergy units/mL (for Df, Dp, and cat allergens); w/v, weight/volume (for dog allergen); Df, *Dermatophagoides farinae*; Dp, *Dermatophagoides pteronyssinus*; RLQ, right lower quadrant of the abdomen.

creased 1 year after ILIT ($P < 0.05$; Supplementary Fig. 5). The serum level of allergen-specific IgG4 to Df showed a similar

trend; however, it was not significantly different 4 months and 1 year after ILIT compared with baseline. The serum level of al-

lergen specific IgG4 to Dp was significantly increased 1 year after ILIT compared with baseline ($P < 0.05$). Neither the serum level of allergen-specific IgE and IgG4 to dog and cat nor the serum total IgE level changed significantly after ILIT (data not shown).

DISCUSSION

All previous studies of ILIT have reported that ILIT causes only mild adverse effects.¹⁻⁷ However, we observed that ILIT could cause severe adverse reactions even at very low concentrations that were not expected to cause serious reactions in SCIT. We therefore propose that AIT of the lymph nodes is not entirely safe, as they are connected to the systemic circulation through the thoracic duct, and ILIT can cause severe adverse reactions even when very low doses of allergen are used.

In subjects who showed moderate-to-severe systemic reactions (subjects 5, 6, and 11), ILIT using Df and Dp at concentrations that, according to SPTs, led to A/H ratios in wheals of more than 1 caused systemic reactions (Supplementary Table 1). We thus suggest that the allergen concentration be reduced in hypersensitized patients, as recommended by the manufacturer of SCIT. In detail, we propose that SPTs be performed with serial dilutions of allergens, that the initial dose of allergen in ILIT not exceed the maximal concentration leading to an A/H ratio in wheals of less than 1, and that we carefully monitor patients undergoing ILIT with allergens at doses exceeding this concentration.

No severe reaction occurred in 2 patients (subject numbers 1 and 7 in Supplementary Table 1), although the allergen dose used in ILIT exceeded the above-mentioned concentration. Regarding severe local and systemic reactions to ILIT, we must also consider other factors such as the type and preparation of allergen and patient clinical characteristics other than hypersensitization.

Like most previous studies of ILIT, the symptoms of AR and quality of life in this study were improved as early as 4 months after the first injection of ILIT, and lasted for 1 year.¹⁻⁶ In NAPT, nasal reactivity to HDM allergens was decreased after ILIT as previously described.^{1-3,5,6} Furthermore, the serum levels of allergen-specific IgE and IgG₄ to Df and Dp were increased 4 months after ILIT, being consistent with the results of previous studies.^{2,3,14} However, these levels were again decreased 1 year after ILIT. Previous studies have reported decreased allergen-specific IgE levels 3 years after ILIT¹ and decreased allergen-specific IgG₄ levels 1 year after ILIT.¹⁴ Regarding dog and cat allergens, we failed to observe any significant change in the level of allergen-specific IgE or IgG₄, due to an inadequate number of subjects. Unlike previous reports,^{1,2,6,7} skin reactivity to allergens in SPT and IDT generally increased after ILIT in this study.

This study has several limitations. First, it is not placebo-controlled. Therefore, the effects of other factors—including phar-

macotherapy, allergen avoidance or other lifestyle modifications, natural course, and subject expectations—should be considered. Additionally, application of ILIT using multiple allergens might have hampered interpretation of the results of this study.

Despite these limitations, our study provides useful information on ILIT. First, the findings suggest that ILIT can provoke serious local or systemic reactions and that a reduced allergen dose, especially of aqueous allergen extracts, should be applied in hypersensitized patients. Secondly, this study is the first of ILIT to evaluate Df, Dp, and dog allergens, which are prevalent globally. Thirdly, this is to our knowledge the first study to use multiple allergens in ILIT.

In conclusion, ILIT can rapidly improve AR symptoms and reduce the frequency of prescription of rescue medication, and this effect lasts for 1 year. However, in hypersensitized patient, ILIT can also cause severe systemic and/or local hypersensitivity reactions when performed using aqueous allergen extract.

ACKNOWLEDGMENTS

We thank Seon Tae Kim for introducing and recommending ILIT to us, and we are especially grateful to Mi Young Myung, a clinical research coordinator for her devotion to our study. This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT and Future Planning (NRF-2015R1D1A1A02061943) and the Gachon University Gil Medical Center (Grant number: 2013-11). We thank ThermoFisher Scientific Korea for support in measuring serum total and allergen-specific IgE/IgG₄ (ImmunoCAP®). For providing allergen extracts for NAPT, we thank the Research Center for Standardization of Allergic Diseases (RCSAD) of Yonsei University supported by a grant from the Korea Healthcare Technology R & D Project, Ministry of Health, Welfare and Family Affairs, Republic of Korea (A092076).

REFERENCES

1. Senti G, Prinz Vavricka BM, Erdmann I, Diaz MI, Markus R, McCormack SJ, et al. Intralymphatic allergen administration renders specific immunotherapy faster and safer: a randomized controlled trial. *Proc Natl Acad Sci U S A* 2008;105:17908-12.
2. Senti G, Cramer R, Kuster D, Johansen P, Martinez-Gomez JM, Graf N, et al. Intralymphatic immunotherapy for cat allergy induces tolerance after only 3 injections. *J Allergy Clin Immunol* 2012; 129:1290-6.
3. Hylander T, Latif L, Petersson-Westin U, Cardell LO. Intralymphatic allergen-specific immunotherapy: an effective and safe alternative treatment route for pollen-induced allergic rhinitis. *J Allergy Clin Immunol* 2013;131:412-20.
4. Patterson AM, Bonny AE, Shiels WE 2nd, Erwin EA. Three-injection intralymphatic immunotherapy in adolescents and young adults with grass pollen rhinoconjunctivitis. *Ann Allergy Asthma*

- Immunol 2016;116:168-70.
5. Hylander T, Larsson O, Petersson-Westin U, Eriksson M, Kumlien Georén S, Winqvist O, et al. Intralymphatic immunotherapy of pollen-induced rhinoconjunctivitis: a double-blind placebo-controlled trial. *Respir Res* 2016;17:10.
 6. Schmid JM, Nezam H, Madsen HH, Schmitz A, Hoffmann HJ. Intralymphatic immunotherapy induces allergen specific plasma-blasts and increases tolerance to skin prick testing in a pilot study. *Clin Transl Allergy* 2016;6:19.
 7. Witten M, Malling HJ, Blom L, Poulsen BC, Poulsen LK. Is intralymphatic immunotherapy ready for clinical use in patients with grass pollen allergy? *J Allergy Clin Immunol* 2013;132:1248-1252.e5.
 8. Brozek JL, Bousquet J, Baena-Cagnani CE, Bonini S, Canonica GW, Casale TB, et al. Allergic rhinitis and its impact on asthma (ARIA) guidelines: 2010 revision. *J Allergy Clin Immunol* 2010;126:466-76.
 9. Shin KS, Cho SH, Ahn TH, Jeong JH, Kim KR, Jung SA, et al. Sino-nasal and psychological symptoms in rhinologic patients (SNOT-20 & SCL-90-R). *Korean J Otorhinolaryngol-Head Neck Surg* 2009; 52:816-21.
 10. Park KH, Cho JS, Lee KH, Shin SY, Moon JH, Cha CI. Rhinoconjunctivitis quality of life questionnaire (RQLQ) as an evaluator of perennial allergic rhinitis patients-the first report. *Korean J Otolaryngol-Head Neck Surg* 2002;45:254-62.
 11. Rondón C, Romero JJ, López S, Antúnez C, Martín-Casañez E, Torres MJ, et al. Local IgE production and positive nasal provocation test in patients with persistent nonallergic rhinitis. *J Allergy Clin Immunol* 2007;119:899-905.
 12. Hilberg O, Pedersen OF. Acoustic rhinometry: recommendations for technical specifications and standard operating procedures. *Rhinol Suppl* 2000;16:3-17.
 13. Nittner-Marszalska M, Cichocka-Jarosz E. Insect sting allergy in adults: key messages for clinicians. *Pol Arch Med Wewn* 2015; 125:929-37.
 14. Freiburger SN, Zehnder M, Gafvelin G, Grönlund H, Kündig TM, Johansen P. IgG4 but no IgG1 antibody production after intralymphatic immunotherapy with recombinant MAT-Feld1 in human. *Allergy* 2016;71:1366-70.