

## Original Article



# Associations between Lifestyle Factors and Iron Overload in Korean Adults

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## ABSTRACT

It has been suggested that iron overload, which indicates the accumulation of iron, generates cellular reactive oxygens and causes peroxide damages to the body. Such oxidative stresses, in a broader context, are also caused by lifestyles such as alcohol consumption and smoking. However, there are limited data on the association between these lifestyle factors and internal iron overload. In present study, we evaluated associations between lifestyle factors, such as smoking status, alcohol consumption, and physical activity, and serum markers of iron overload. In a population-based cross-sectional study including 2,347 Korean men and women aged 49–79 years, we assessed serum transferrin saturation (TSAT) levels and defined iron overload as TSAT levels > 50% for men and > 45% for women. After excluding persons with chronic diseases and iron deficiency, multivariate odds ratio (OR) and its 95% confidence interval (CI) were estimated to evaluate associations between lifestyle factors and iron overload in 1,973 participants. In all participants, we examined a significantly positive association between heavy alcohol consumption (> 30 g/day) and iron overload; heavy drinkers showed 1.6-fold higher OR (95% CI, 1.11–2.36) than non-drinkers. Stratified analysis by sex showed that this association was significant only among men. In addition, we observed a potential association between heavy smoking > 10 cigarettes/day and iron overload ( $p = 0.07$ ). In stratified analysis by sex, we examined a significant association between smoking and iron overload only among women; former or current smokers had 1.9-fold higher OR (95% CI, 1.01–3.63) than never-smoker. Our findings suggest that heavy alcohol consumption and smoking may worsen iron accumulation in the body.

**Keywords:** Smoking; Alcohol consumption; Transferrin saturation; Iron overload

## INTRODUCTION

Iron, an essential component in human tissue, plays a crucial role in transporting oxygen in the body as well as in cellular processes including DNA, RNA, and protein synthesis [1-4]. Hemoglobin, an oxygen-carrying protein in red blood cells, accounts for about two-thirds of body's iron supply. About 10% of essential iron contributes to the formation of myoglobin, cytochrome, iron-containing enzymes. The remaining of iron is stored as ferritin and hemosiderin. Iron deficiency has been one of common nutritional problems around the world. Recently, attention has been paid to iron overload, which is the opposite problem of iron deficiency, in developed countries [1,3].

### Conflict of Interest

The authors declare that they have no conflict of interest.

### Author Contributions

Chol Shin and Inkyung Baik had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

Iron overload is characterized by the excessive accumulation of iron in the body, which is known as hemochromatosis. It is defined as serum ferritin levels > 400 ng/mL for men and > 300 ng/mL for women or as transferrin saturation (TSAT) levels > 50% for men and > 45% for women [2]. It has been suggested that excess amount of free iron in the body cause the formation of oxygen radicals leading to peroxidative damages to vital cell structures. In fact, it was reported that iron overload was associated with increased risks of liver disease, diabetes mellitus, cardiovascular disease, and Parkinson's and Alzheimer's diseases [3,4].

Data on the association between lifestyle factors, such as alcohol consumption, smoking, physical activity, and iron overload are still limited and inconsistent. Alcohol consumption tends to increase iron absorption by down-regulating hepcidin, which is a major regulator of iron homeostasis, and may lead to iron accumulation in the body [5,6]. However, epidemiological data on the alcohol consumption and iron overload in general populations are limited [7]. There have been controversial findings on the association between smoking and iron status. One study reported no difference in plasma iron levels between non-smokers and smokers [8]. But, another study examined that pregnant women who had smoked had higher ferritin levels in blood than those who had never smoked [9]. This discrepancy may be due to the use of different iron biomarkers; ferritin and TSAT are better indicators of iron status than iron in serum or plasma. In terms of the link between physical activity and iron status, some studies reported that ferritin and transferrin levels in blood were elevated after treadmill running test [10,11]. Limitations of these previous studies on the association between lifestyle factors and iron overload include a small sample size and a lack of generalizability. In addition, to the best of our knowledge, there is no study to examine joint effects of lifestyle factors on iron overload.

In present study, we investigated on the associations of smoking status, alcohol consumption, and physical activity with iron overload, which was defined by TSAT levels. Furthermore, we conducted joint analysis of smoking status and alcohol consumption in the association with iron overload.

## MATERIALS AND METHODS

### Study population

We carried out a cross-sectional study embedded in a population-based cohort study, which is an ongoing longitudinal investigation [12,13]. The study participants were 2,347 Korean aged 49–79 years who completed the follow-up interview of the cohort study between February 2011 and November 2012 and provided blood samples for assays of iron status biomarkers. In the questionnaire-based interview, they provided information on socio-demographics, lifestyle factors, health status, and medical history. They signed an informed consent form approved by the Human Subjects Review Committee at the Korea University Ansan Hospital. After excluding participants who reported diagnosis of cancer, diabetes mellitus, and hypertension or who showed iron deficiency (TSAT levels < 15%) or who had missing values of lifestyle factors were excluded. Finally, 1,973 participants were remained for the analysis.

### Blood collection and assessments

All participants fasted for at least 8 hours before the beginning of blood collection. Serum samples were collected and delivered to the Seoul Clinical Laboratories (Seoul, Korea) for assays of iron, ferritin, and unsaturated iron-binding capacity (UIBC), which was used to calculate total iron-binding capacity (TIBC) and TSAT. TIBC was obtained by adding

serum iron and UIBC concentrations, and TSAT was calculated by multiplying the ratio of serum iron and TIBC by 100. Regarding assay methods, colorimetry for iron and UIBC and immunoassays for ferritin were used. Applicable assay kits were purchased from Siemens (Tarrytown, NY, USA) and Nittobo (Osaka, Japan).

### Lifestyle factors and other confounding variables

Information on lifestyle factors such as smoking status, alcohol consumption, and physical activity levels was collected. Anthropometric measurement was conducted by a trained researcher. According to a standardized protocol, height (cm) and body weight (kg) were measured to the nearest 0.1 cm or 0.1 kg, respectively, without footwear, and body mass index (BMI, kg/m<sup>2</sup>) was calculated. Information on smoking status (never/former smoker/current smoker) and the number of cigarettes smoked was obtained. Information on the average frequency of alcohol drinking occasions, the amount of alcohol consumed on a typical occasion, and the volume of one standard drink for each alcoholic beverage was obtained, and the daily amount of alcohol consumed (g/day) was calculated. Participants were also asked to complete a table that inquired about alcohol drinking amount and pattern in the past 30 days. Thus, we collected information on average frequency of drinking occasion, amount of 6 types of alcoholic beverage (beer, wine, hard liquor, and 3 types of traditional drinks, including soju, chungju, and makgeolli) consumed for a typical occasion, and the volume of 1 standard drink for each type of alcoholic beverage. We calculated beverage-specific alcohol consumption in g/day on the basis of the alcohol content (4.5% for beer, 12% for wine, 40% for hard liquor, 22% or 25% for soju, 16% for chungju, and 6% for makgeolli), the frequency of drinking, and the amount consumed. We calculated total alcohol consumption (g/day) by summing up the beverage-specific amount consumed as well as total liquor consumption (g/day) by summing up the alcohol amount from liquor, including hard liquor, soju, and chungju. Information on physical activity level was obtained using five categories for activity intensity with open-ended questions about the hours spent in a typical day per level of intensity. A total metabolic equivalent score (MET) was calculated by multiplying the hours spent at a particular activity intensity by its MET values (1.0 for sleep or sedentary, 1.5 for very light activity, 2.4 for light activity, 5.0 for moderate activity, and 7.5 for vigorous activity), which were determined based on examples of activities given for each category. Confounding variables such as age, sex, and the presence of hypertension or diabetes mellitus were collected. The presence of hypertension was confirmed by using antihypertensive medications or having systolic blood pressure (BP) of  $\geq 140$  mmHg or diastolic BP of  $\geq 90$  mmHg. BP was measured in a sitting position using a mercury sphygmomanometer after a rest period of at least 5 minutes. Repeated BP measurements were performed at approximately 30-second intervals and recorded to the nearest 2 mmHg. The average of two measurements each in the left and right arms was calculated for systolic and diastolic BP. The presence of diabetes mellitus was confirmed if the following criteria were met: fasting plasma glucose level of  $\geq 126$  mg/dL or postprandial glucose level of  $\geq 200$  mg/dL or use of medications for diabetes mellitus.

### Statistical analysis

According to the quartiles of TSAT levels, descriptive statistics for demographic and clinical characteristics, information on lifestyle factors, and blood assay results of the study participants were calculated. For continuous data, the ANOVA test was used to assess differences in means across the quartiles of TSAT levels. For categorical data, a chi-square test was used to assess differences in proportions across the quartiles of TSAT levels. We defined iron overload as TSAT levels  $> 50\%$  for men and  $> 45\%$  for women. To analyze associations between iron overload and lifestyle factors, we conducted multivariate logistic regression analysis and obtained odds

ratio (OR) and 95% confidence interval (CI). In the multivariate models, covariates including age (continuous), sex, BMI (continuous), smoking status (categories: nonsmoking, former smoking, current smoking of 1–10 cigarettes/day, or current smoking of > 10 cigarettes/day), alcohol consumption (categories: nondrinking, alcohol consumption of 1–14.9 g/day, 15–30 g/day, or > 30 g/day), MET for physical activity (continuous), the presence of hypertension or diabetes mellitus (binary) were included. We also performed stratified analysis by sex and joint analysis of smoking status and alcohol consumption in the association with iron overload. Statistical significance was set at 0.05. All statistical analyses were performed with the SAS version 9.4 software (SAS Institute, Cary, NC, USA).

## RESULTS

This cross-sectional study included 1,118 (56.7%) men and 855 women (43.3%). Among them, 288 (25.8%) men and 129 (15.1%) women had iron overload. **Table 1** shows descriptive statistics on lifestyle factors, confounding variables, and iron biomarkers according to the quartiles of TSAT levels. Participants with higher TSAT levels were more likely to be younger ( $p < 0.01$ ), men, smokers, and alcohol drinkers ( $p < 0.001$ ) and less likely to have hypertension ( $p < 0.01$ ).

**Table 2** presents results for the associations between iron overload and lifestyle factors in all participants. Heavy alcohol drinkers had 1.6-fold higher OR (95% CI, 1.11–2.36) of iron overload than non-drinkers. Heavy smokers were likely to have a higher OR than never-smokers ( $p = 0.07$ ).

Next, we evaluated associations stratified by sex between lifestyle factors and iron overload (**Table 3**). In men, heavy alcohol drinkers had 1.7-fold higher OR (95% CI, 1.13–2.56) of iron overload than non-drinkers while heavy smoking tended to increase OR ( $p = 0.19$ ). In women, former or current smokers had 1.9-fold higher OR (95% CI, 1.01–3.63) of iron overload than never-smokers while moderate or heavy alcohol consumption tended to increase OR ( $p = 0.62$ ).

**Table 1.** Characteristics of 1,973 participants by transferrin saturation levels (n = 1,973)

Characteristics	Classification	Quartiles of TSAT (range, %)				p value
		1st quartile (15.6–30.6)	2nd quartile (30.6–37.6)	3rd quartile (37.6–46.4)	4th quartile (46.4–98.2)	
No. of subjects	-	490	490	497	496	-
Age, yr	-	58.6 ± 7.0	58.5 ± 7.0	58.7 ± 7.4	57.3 ± 6.4	< 0.01
Men, %	-	42.9	45.7	60.2	77.6	< 0.001
BMI, kg/m <sup>2</sup>	-	24.8 ± 2.9	24.9 ± 3.0	24.5 ± 2.7	24.5 ± 2.7	0.08
Current smoking, %	No	89.6	86.7	87.5	78.8	< 0.001
	Yes	10.4	13.3	12.5	21.2	-
Current alcohol drinking, %	No	59.2	54.5	46.1	35.7	< 0.001
	Yes	40.8	45.5	53.9	64.3	-
Physical activity, MET	-	40.5 ± 5.9	40.9 ± 6.4	40.8 ± 6.4	41.3 ± 6.3	0.28
Presence of hypertension, %	No	59.8	63.7	65.2	70.2	< 0.01
	Yes	40.2	36.3	34.8	29.8	-
Presence of diabetes mellitus, %	No	90.8	90.6	90.1	91.9	0.79
	Yes	9.2	9.4	9.9	8.1	-
Serum iron biomarkers						
Iron, µg/dL	-	79.4 ± 17.6	104.3 ± 16.7	129.1 ± 17.0	175.6 ± 40.0	< 0.001
Ferritin, µg/L	-	92.3 ± 60.5	105.2 ± 80.6	121.7 ± 95.2	152.7 ± 139.2	< 0.001
TIBC, µg/dL	-	316.2 ± 58.3	306.1 ± 46.8	311.8 ± 35.3	306.1 ± 42.0	< 0.001

Values expressed as percentage or mean ± standard deviation (SD).

BMI, body mass index; MET, total metabolic equivalent score; TIBC, total Iron binding capacity; TSAT, transferrin saturation.

**Table 2.** Associations between lifestyle factors and iron overload (n = 1,973)

Lifestyle factors	No. of subjects	OR (95% CI)			
		Age-adjusted model	p value	Multivariate model*	p value
Smoking status					
Never smoked	1,105	reference	-	reference	-
Former smoker	585	1.73 (1.35–2.22)	< 0.001	1.24 (0.87–1.75)	0.23
Current smoker, cigarettes/day					
Light smoker (0 to 10)	88	1.94 (1.19–3.17)	< 0.01	1.31 (0.76–2.25)	0.33
Heavy smoker (> 10)	195	2.19 (1.55–3.08)	< 0.001	1.48 (0.97–2.27)	0.07
Alcohol consumption					
Non-drinker	963	reference	-	reference	-
Drinker, g/day					
Light drinker (0 to 14.9)	564	1.33 (1.02–1.73)	< 0.05	1.11 (0.84–1.47)	0.45
Moderate drinker (15 to 30)	216	1.64 (1.16–2.32)	< 0.01	1.24 (0.84–1.81)	0.28
Heavy drinker (> 30)	230	2.08 (1.45–2.89)	< 0.001	1.61 (1.11–2.36)	< 0.05
Physical activity (MET range)					
1st quartile (26 to 36.9)	473	reference	-	reference	-
2nd quartile (37 to 39.9)	548	0.93 (0.68–1.27)	0.64	0.89 (0.65–1.22)	0.46
3rd quartile (40 to 44)	436	1.11 (0.80–1.52)	0.54	1.06 (0.76–1.47)	0.74
4th quartile (> 44)	516	1.14 (0.84–1.56)	0.40	1.03 (0.75–1.40)	0.88

OR, odds ratio; CI, confidence interval; MET, total metabolic equivalent score.

\*Data are adjusted for age, sex, body mass index, smoking status, alcohol consumption, physical activity, and the presence of hypertension or diabetes mellitus.

**Table 3.** Associations between lifestyle factors and iron overload (n = 1,973)

Lifestyle factors	No. of subjects	OR (95% CI)	
		Multivariate model*	p value
Men	1,118	-	-
Smoking status			
Never smoked	266	reference	-
Former smoker	578	1.19 (0.83–1.69)	0.34
Current smoker, cigarettes/day			
Light smoker (0 to 10)	81	0.99 (0.55–1.79)	0.97
Heavy smoker (> 10)	193	1.34 (0.87–2.06)	0.19
Alcohol consumption			
Non-drinker	314	reference	-
Drinker, g/day			
Light drinker (0 to 14.9)	382	1.25 (0.87–1.79)	0.22
Moderate drinker (15 to 30)	199	1.27 (0.84–1.95)	0.26
Heavy drinker (> 30)	223	1.70 (1.13–2.56)	< 0.05
Physical activity (MET range)			
1st quartile (26 to 36.9)	245	reference	-
2nd quartile (37 to 39.9)	293	0.82 (0.55–1.22)	0.33
3rd quartile (40 to 44)	251	0.93 (0.62–1.40)	0.73
4th quartile (> 44)	329	1.02 (0.69–1.49)	0.93
Women	855	-	-
Smoking status			
Never smoked	839	reference	-
Former or current smoker	16	1.91 (1.01–3.63)	< 0.05
Alcohol consumption			
Non-drinker	649	reference	-
Drinker, g/day			
Light drinker (0 to 15)	182	0.92 (0.57–1.50)	0.74
Moderate or heavy drinker (> 15)	24	1.31 (0.45–3.88)	0.62
Physical activity (MET range)			
1st quartile (26 to 36.9)	228	reference	-
2nd quartile (37 to 39.9)	255	1.00 (0.59–1.69)	0.99
3rd quartile (40 to 44)	185	1.36 (0.79–2.35)	0.27
4th quartile (> 44)	187	1.01 (0.56–1.79)	0.98

OR, odds ratio; CI, confidence interval; MET, total metabolic equivalent score.

\*Data are adjusted for age, body mass index, smoking status, alcohol consumption, physical activity, and the presence of hypertension or diabetes mellitus.

**Table 4.** Odds ratios of iron overload in joint analysis of smoking status and alcohol consumption in all participants (n = 1,973)

Smoking status	Alcohol consumption	No. of subjects	OR (95% CI)			
			Age-adjusted model	p value	Multivariate model*	p value
Non-smoker	No	748	reference	-	reference	-
Former smoker, cigarettes/day	No	150	1.76 (1.15–2.71)	< 0.01	1.30 (0.77–2.20)	0.33
0 to 10		18	2.76 (1.01–7.52)	< 0.05	1.88 (0.66–5.36)	0.24
> 10		47	1.58 (0.78–3.20)	0.21	1.09 (0.51–2.36)	0.82
Non-smoker	0 to 14.9 g/day	283	1.26 (0.88–1.81)	0.21	1.12 (0.77–1.63)	0.56
Former smoker, cigarettes/day	0 to 14.9 g/day	202	1.83 (1.25–2.67)	< 0.01	1.41 (0.87–2.30)	0.17
0 to 10		35	1.89 (0.86–4.14)	0.11	1.32 (0.57–3.04)	0.52
> 10		44	1.99 (0.99–4.00)	0.05	1.44 (0.67–3.11)	0.35
Non-smoker	15 to 30 g/day	45	1.96 (0.98–3.92)	0.06	1.66 (0.80–3.46)	0.17
Former smoker, cigarettes/day	15 to 30 g/day	115	1.62 (1.01–2.63)	< 0.05	1.26 (0.71–2.22)	0.43
0 to 10		18	1.08 (0.31–3.79)	0.91	0.78 (0.21–2.87)	0.71
> 10		38	3.35 (1.69–6.64)	< 0.001	2.63 (1.23–5.61)	< 0.05
Non-smoker	> 30 g/day	29	0.83 (0.28–2.45)	0.74	0.81 (0.27–2.47)	0.71
Former smoker, cigarettes/day	> 30 g/day	118	2.49 (1.60–3.86)	< 0.001	2.05 (1.19–3.52)	< 0.01
0 to 10		17	3.57 (1.33–9.60)	< 0.05	3.04 (1.07–8.68)	< 0.05
> 10		66	2.94 (1.71–5.07)	< 0.001	2.38 (1.26–4.48)	< 0.01

OR, odds ratio; CI, confidence interval.

\*Data are adjusted for age, sex, body mass index, smoking status, alcohol consumption, physical activity, and the presence of hypertension or diabetes mellitus.

**Table 4** presents results for the association iron overload in joint analysis of smoking status and alcohol consumption. Compared with non-smokers who abstained from alcohol consumption, smokers with > 10 cigarettes/day who consumed alcohol > 15 g/day had a 2.6-fold higher OR (95% CI, 1.23–5.61) and any smokers with heavy alcohol consumption > 30 g/day had higher ORs between 2 and 3 for iron overload.

## DISCUSSION

In a population-based cross-sectional study, we observed positive associations of alcohol consumption and smoking status with iron overload. In particular, the association for alcohol consumption was significant only among men while that for smoking was only among women. In addition, heavy smokers were more likely to have iron overload even if they had moderate alcohol consumption with 15 to 30 g/day. Heavy alcohol drinkers with > 30 g/day were more likely to have iron overload even if they were former smokers or moderate smokers with < 10 cigarettes/day.

Serum TSAT levels indicates the proportion of iron-binding sites of transferrin loaded with iron and has been used as a sensitive biomarker of iron status than serum iron or TIBC [14]. Its level is determined by sex, race, iron consumption, and the presence of infection or metabolic diseases as well as by genetic factors. Its normal values range between 15% and 40% approximately and thus TSAT levels > 50% for men or > 45% for women indicate iron overload or hemochromatosis. Iron overload with TSAT levels > 50% may lead to the accumulation of non-transferrin-bound iron or labile iron, which is involved in the production of reactive oxygen species (ROS) via the Fenton reaction or the Haber-Weiss reaction [3,4]. ROS have potency to damage cells and DNA.

Alcohol-induced disorders with abnormal iron metabolism have been investigated in animal models as well as in clinical and epidemiological studies. It was reported that patients with alcoholic liver disease are likely to have increased iron stores, as reflected by elevated serum



iron biomarkers such as TSAT or ferritin. Alcohol consumption worsens the severity of disease in patients with genetic hemochromatosis [15]. A potential biological mechanism underlying these observations is that alcohol may be implicated in the regulation of hepcidin expression in the liver. Hepcidin is a key regulator of iron status by controlling iron absorption in the intestine. Reduced hepcidin concentrations by down-regulation of hepcidin expression allow intestinal absorption of iron leading to the elevation of circulating iron as well as iron storage. It was reported that alcohol consumption down-regulates hepcidin expression in a vitro study with hepatoma cells as well as in a vivo study with mice exposed to alcohol [16]. Hepcidin concentrations decreased after drinking alcohol and then iron absorption increased with promoting ferroportin as a corridor in the gastrointestinal cells and membrane of macrophage [17,18].

Prior studies have examined elevated iron concentrations in smokers. As a potential biological mechanism, it has been suggested that smoking may stimulate erythropoiesis in a dose-related manner due to hypoxia, which is induced by smoking. In addition, hypoxia is likely to suppress hepcidin expression [8,9,19-22].

Physical activity, especially intense physical activity, may cause a number of unfavorable changes, including an increase in the level of pro-inflammatory cytokines and decreased iron absorption [10]. One study showed that a high-intensity exercise test caused significant changes in hepcidin levels, interleukin-6, and iron metabolism during the recovery period [11]. Another study observed that serum iron levels decreased significantly during the recovery period after exercise [23].

The strengths of our study include a large sample size, investigation in a general population, consideration of a variety of lifestyle factors, and presentation of joint effects of lifestyle factors in the association with iron overload. Study limitations should be taken into account for the interpretation of our findings. Because the study design was cross-sectional, a causal relationship remains unclear. Because the findings may be generalizable to middle-aged and older Asians, further studies for other age groups or other ethnicities are warranted.

In this cross-sectional study, we observed that heavy alcohol drinking and smoking are positively associated with iron overload. On the basis of the findings, heavy alcohol drinking and smoking may need to be avoided to prevent iron accumulation in the body.

## REFERENCES

1. Lieu PT, Heiskala M, Peterson PA, Yang Y. The roles of iron in health and disease. *Mol Aspects Med* 2001;22:1-87.  
[PUBMED](#) | [CROSSREF](#)
2. Zijp IM, Korver O, Tijburg LB. Effect of tea and other dietary factors on iron absorption. *Crit Rev Food Sci Nutr* 2000;40:371-98.  
[PUBMED](#) | [CROSSREF](#)
3. Papanikolaou G, Pantopoulos K. Iron metabolism and toxicity. *Toxicol Appl Pharmacol* 2005;202:199-211.  
[PUBMED](#) | [CROSSREF](#)
4. Casu C, Rivella S. Iron age: novel targets for iron overload. *Hematology Am Soc Hematol Educ Program* 2014;2014:216-21.  
[PUBMED](#) | [CROSSREF](#)

5. Lemos Ados R, Ismael LA, Boato CC, Borges MT, Rondó PH. Hepcidin as a biochemical parameter for the assessment of iron deficiency anemia. *Rev Assoc Med Bras* (1992) 2010;56:596-9.  
[PUBMED](#) | [CROSSREF](#)
6. Ganz T. Hepcidin and its role in regulating systemic iron metabolism. *Hematology Am Soc Hematol Educ Program* 2006;29-35, 507.  
[PUBMED](#) | [CROSSREF](#)
7. Lieb M, Palm U, Hock B, Schwarz M, Domke I, Soyka M. Effects of alcohol consumption on iron metabolism. *Am J Drug Alcohol Abuse* 2011;37:68-73.  
[PUBMED](#) | [CROSSREF](#)
8. Kocyigit A, Erel O, Gur S. Effects of tobacco smoking on plasma selenium, zinc, copper and iron concentrations and related antioxidative enzyme activities. *Clin Biochem* 2001;34:629-33.  
[PUBMED](#) | [CROSSREF](#)
9. Chelchowska M, Ambroszkiewicz J, Gajewska J, Jabłońska-Głąb E, Maciejewski TM, Oltarzewski M. Hepcidin and iron metabolism in pregnancy: correlation with smoking and birth weight and length. *Biol Trace Elem Res* 2016;173:14-20.  
[PUBMED](#) | [CROSSREF](#)
10. Sim M, Dawson B, Landers G, Trinder D, Peeling P. Iron regulation in athletes: exploring the menstrual cycle and effects of different exercise modalities on hepcidin production. *Int J Sport Nutr Exerc Metab* 2014;24:177-87.  
[PUBMED](#) | [CROSSREF](#)
11. Schumacher YO, Schmid A, König D, Berg A. Effects of exercise on soluble transferrin receptor and other variables of the iron status. *Br J Sports Med* 2002;36:195-9.  
[PUBMED](#) | [CROSSREF](#)
12. Baik I, Seo HS, Yoon D, Kim SH, Shin C. Associations of sleep apnea, NRG1 polymorphisms, alcohol consumption, and cerebral white matter hyperintensities: analysis with genome-wide association data. *Sleep* 2015;38:1137-43.  
[PUBMED](#) | [CROSSREF](#)
13. Baik I, Shin C. Interactions between the FTO rs9939609 polymorphism, body mass index, and lifestyle-related factors on metabolic syndrome risk. *Nutr Res Pract* 2012;6:78-85.  
[PUBMED](#) | [CROSSREF](#)
14. Wish JB. Assessing iron status: beyond serum ferritin and transferrin saturation. *Clin J Am Soc Nephrol* 2006;1 Suppl 1:S4-8.  
[PUBMED](#) | [CROSSREF](#)
15. Whitfield JB, Zhu G, Heath AC, Powell LW, Martin NG. Effects of alcohol consumption on indices of iron stores and of iron stores on alcohol intake markers. *Alcohol Clin Exp Res* 2001;25:1037-45.  
[PUBMED](#) | [CROSSREF](#)
16. Harrison-Findik DD. Role of alcohol in the regulation of iron metabolism. *World J Gastroenterol* 2007;13:4925-30.  
[PUBMED](#) | [CROSSREF](#)
17. Ioannou GN, Dominitz JA, Weiss NS, Heagerty PJ, Kowdley KV. The effect of alcohol consumption on the prevalence of iron overload, iron deficiency, and iron deficiency anemia. *Gastroenterology* 2004;126:1293-301.  
[PUBMED](#) | [CROSSREF](#)
18. Fletcher LM, Halliday JW, Powell LW. Interrelationships of alcohol and iron in liver disease with particular reference to the iron-binding proteins, ferritin and transferrin. *J Gastroenterol Hepatol* 1999;14:202-14.  
[PUBMED](#) | [CROSSREF](#)
19. Shibata Y, Inoue S, Igarashi A, Yamauchi K, Abe S, Aida Y, Nunomiya K, Sato M, Nakano H, Sato K, Watanabe T, Konta T, Ueno Y, Kato T, Kayama T, Kubota I. Elevated serum iron is a potent biomarker for spirometric resistance to cigarette smoke among Japanese males: the Takahata study. *PLoS One* 2013;8:e74020.  
[PUBMED](#) | [CROSSREF](#)
20. Zeng Q, Shen LJ, Li S, Chen L, Guo X, Qian CN, Wu PH. The effects of hemoglobin levels and their interactions with cigarette smoking on survival in nasopharyngeal carcinoma patients. *Cancer Med* 2016;5:816-26.  
[PUBMED](#) | [CROSSREF](#)
21. Pateva IB, Kerling EH, Reddy M, Chen D, Carlson SE, Tancabelic J. Effect of maternal cigarette smoking on newborn iron stores. *Clin Res Trials* 2015;1:4-7.  
[PUBMED](#)



22. Ghio AJ, Hilborn ED, Stonehuerner JG, Dailey LA, Carter JD, Richards JH, Crissman KM, Foronjy RF, Uyeminami DL, Pinkerton KE. Particulate matter in cigarette smoke alters iron homeostasis to produce a biological effect. *Am J Respir Crit Care Med* 2008;178:1130-8.  
[PUBMED](#) | [CROSSREF](#)
23. Skarpańska-Stejnborn A, Basta P, Trzeciak J, Szcześniak-Pilaczyńska Ł. Effect of intense physical exercise on hepcidin levels and selected parameters of iron metabolism in rowing athletes. *Eur J Appl Physiol* 2015;115:345-51.  
[PUBMED](#) | [CROSSREF](#)