

Comparison of Primers for Oral Mycobiome Study in Intubated Patients

Yoon Hee Choi¹, Soo Hyun Kim², Myoung Soo Kim³, Hee Sam Na^{4*}

¹Department of Internal Medicine, Dongnam Institute of Radiological and Medical Sciences, Busan 46033, Republic of Korea

²Artificial Kidney Room, Busan Medical Center, Busan 47527, Republic of Korea

³Department of Nursing, College of Natural Science, Pukyong National University, Busan 48513, Republic of Korea

⁴Department of Oral Microbiology, BK21 PLUS Project, School of Dentistry, Pusan National University, Yangsan 50612, Republic of Korea

Corresponding

Hee Sam Na, Professor
Department of Oral Microbiology,
School of Dentistry, Pusan National
University, Busandaehak-ro 49,
Mulgeum-eup, Yangsan,
Gyeongsangnam-do, 50612, Republic of
Korea

Phone : +82-51-510-8252

Fax : +82-51-510-8246

E-mail : heesamy@pusan.ac.kr

Received : July 1, 2021

Revised : August 5, 2021

Accepted : August 6, 2021

No potential conflict of interest relevant
to this article was reported.

In intubated patients, oral fungus including *Candida* rapidly can increase. Fungal infections cause a significant proportion of health care associated infections. For efficient prevention and management, characterization of the oral mycobiome is required. In this study, we tested two primers to characterize the oral mycobiome in intubated patients under mechanical ventilation support. Buccal samples from intubated patients were collected using Levine technique. Two primers (ITS3 and ITS7) targeting internal transcribed spacer (ITS)2 region was tested. Next generation sequencing was applied for mycobiome analysis. Alpha diversity, beta diversity and relative abundance was determined to compare the primer performance. The richness and evenness were significantly higher using ITS7 primer. When phylogenetic distance was included, there was no difference in richness. In beta diversity analysis, most of the samples were closely clustered suggesting that similar mycobiome structure was assigned by the tested primers. When relative abundance of the assigned taxa was compared, *Candida* was the most abundant genus. At the species level, similar species was assigned by both primers. However, *C. sake*, *C. intermedia*, *C. parapsilosis* and *Yarrowia porcina* was only assigned by ITS3 primer, while *Alternaria betae-kenyensis* was only assigned by ITS7 primer. In intubated patients, diverse oral mycobiome was detected. Although the tested primers predicted similar mycobiome structure, there were several species that was uniquely assigned by each primer. Selecting optimum primer to characterize the oral mycobiome should provide better understanding the mycobiome community in oral cavity.

Key Words: Oral mycobiome, Intubation, ITS2, *Candida*

INTRODUCTION

Analysis of the mycobiome has provided deep understanding into their role in human health and disease (1). In healthy subjects, the biomass of the oral mycobiome is relatively low compared to the bacterial load (2). Intubation and critical illness can reduce oral immunity and exacerbate poor oral hygiene (3, 4). In intubated patients, fungal load including *Candida* rapidly increases after admission to an intensive care unit (ICU) (5). Fungal infections cause a significant proportion of health care associated infections (HAIs) and *Candida* species have been reported as the most prevalent pathogens causing HAIs (6, 7). Characterizing oral mycobiome is the first step to understand the shift of oral mycobiome and design proper management.

Copyright © 2021 Journal of Bacteriology
and Virology

©This is an Open Access article
distributed under the terms of the Creative
Commons Attribution Non-Commercial
License
([http://creativecommons.org/
license/by-nc/3.0/](http://creativecommons.org/licenses/by-nc/3.0/)).

Over the past few years, internal transcribed spacer (ITS) regions has been considered for mycobiome community survey, and many studies have been used various primer sets to characterize mycobiome structure (8-13). The ITS region is located between the 18S and 26S rRNA genes and is subdivided into the ITS1 region and the ITS2 region. ITS2 is located between the 5.8S and 26S rRNA genes and it has been suggested to be useful for the identification of clinically important fungi (14). ITS3/ITS4 primer and ITS7/ITS4 primer are among the primers targeting ITS2 region to evaluate the mycobiome community (8, 15-17).

However, no study has tested the performance of these primer sets on oral mycobiome communities. Therefore, we tested two primers (ITS3/ITS4 and ITS7/ITS4) targeting ITS2 to evaluate their performance on oral mycobiome analysis.

MATERIAL AND METHODS

Participants

The inclusion criteria for ICU patients were; 1) > 18 years of age, 2) endotracheal tube use with holding methods, 3) consent for participation from the patient's family. This study was approved by the Institutional Review Board (DAUHIRB-19-072). ICU participants in this study were mostly sedated or at severely poor levels of consciousness; therefore, family members were considered as the legal representatives with the power of attorney based on the International Council for Harmonization (18). During the participant recruitment period, the purpose, voluntary nature of participation, confidentiality of information, and procedures used in the study were explained to family members, and informed consent was obtained from each family.

Oral swab sample preparation

Buccal swap samples were collected using the Levine technique; the swab was rotated over an approximately 1cm² area with sufficient pressure (19). Sterile cotton swabs were used for the collection and were placed in a 15 mL conical tube labelled with the sample ID and stored at -80°C.

Extraction of Genomic DNA and Next Generation Sequencing

Before analysis, samples were suspended in phosphate buffered saline (PBS), vortexed, and then centrifuged. DNA was extracted using a Gram positive DNA purification kit (Lucigen, Biosearch Technology, Novato, CA) following the manufacturer's instructions. The final concentration was measured with a NanoDrop ND-1000 spectrophotometer (Thermo Fisher Scientific Inc, Waltham, MA) and stored at -80°C until use. PCR amplification of the ITS2 was performed. The primer sequences used for amplifications were as follows: ITS3-F: 5'- GCA TCG ATG AAG AAC GCA GC-3', ITS7-F: 5'- GTG ART CAT CGA RTC TTT G-3', ITS4-R: TCC TCC GCT TAT TGA TAT GC-3' (15, 16). Theoretical target size for ITS3/ITS4 and ITS7/ITS4 primer pair were 300-400 bps and 230-330 bps, respectively. Purified amplicons were pooled in equimolar and paired-end sequenced with MiSeq (Illumina, San Diego, CA, USA).

Bioinformatic Analysis, Statistical Analysis, and Visualization

Basic microbiome analyses have been performed using the QIIME2 (version 2020.6) (20) and associated plugins. To measure alpha diversities, Faith PD index, Chao1 index, and Shannon's index method were used. For the beta diversity analysis, the weighted UniFrac was used. The Mann-Whitney U test was used to assess the statistical significances between the groups for alpha diversities. To assign taxonomy to the unique representative sequences, pre-trained Naive Bayes classifier, using UNITE Database (21), was used. Phyloseq package was used to construct phylogenetic tree (22).

RESULTS

Participant characterization and sequence processing result

The 5 enrolled ICU patients (all male, age 68.4 ± 8.5) included a heterogeneous set of underlying diseases and acute indications for mechanical ventilation (Table 1). Four subjects were sampled at multiple time points. A total of 20 samples were tested with both ITS3 and ITS7 primers.

A total of 1,347,141 reads ($67,357 \pm 32,979$) were collected using ITS3 primers and 2,111,074 reads ($91,786 \pm 34,952$) were collected using ITS7 primers. DADA2 was applied and final read count were $62,535 \pm 29,007$ and $82,956 \pm 34,574$ for ITS3 and ITS7, respectively. Total read counts obtained from ITS7 primers was significantly higher than ITS3 primers (Table 2). The minimum read count was 9,011 and 8,697 for ITS3 and ITS7, respectively. When rarefaction curve of observed OTU was plotted, all the samples reached plateau, suggesting sufficient number of read counts were obtained for further analysis (Fig. 1). The paired read length was 359.4 ± 47.3 and 311.5 ± 43.4 for ITS3 and ITS7, respectively. ITS3 primers produced significantly longer read length compared to ITS7 primers.

Diversity of the mycobiome

ITS2 region was targeted to compare the reliability of fungal diversity surveying the oral mycobiome composition. The alpha diversity was estimated by Chao1, Faith PD and Shannon test. When the phylogeny of the taxa was not considered, ITS7 showed higher richness (Chao1, $p=0.022586$) (Fig. 2A) and evenness (Shannon index, $p=0.000018$) (Fig. 2C). However, there was no significant difference in Faith PD, which uses phylogenetic distance to calculate the richness of the diversity (Fig. 2B). To compare the mycobiome structure, beta diversity was determined using weighted UniFrac, which considers the phylogeny of the taxa. The respective oral samples showed nearby distribution suggesting the mycobiome was similar between ITS3 primer and ITS7 primers (Fig. 2D).

Table 1. Characterization of patients

Subject	Age	Sex	Underlying disease
1	81	M	COPD, DM, Alcoholic hepatitis
2	65	M	COPD, DM
3	67	M	Alcoholic hepatitis
4	71	M	Lung Cancer
5	58	M	DM, Liver cirrhosis

COPD: Chronic Obstruction Pulmonary Disease; DM: Diabetes mellitus

Table 2. The number of sequence reads during data processing and paired read length

	ITS3	ITS7	<i>p</i> value
Input read count	67357 ± 32979 (range: 9707 - 172172)	91786 ± 34952 (range: 9961 - 140863)	0.023
Filtered read count	64322 ± 31595	87876 ± 33953	0.023
Denoised read count	64273 ± 31582	87802 ± 33935	0.023
Merged read count	63991 ± 31475	87214 ± 33682	0.024
Non-chimeric read count	62535 ± 29007 (range: 9011 - 150044)	82956 ± 34574 (range: 8697 - 133929)	0.041
Paired read length (bp)	359.4 ± 47.3 (range: 272 - 462)	311.5 ± 43.4 (range: 272 - 435)	< 0.001

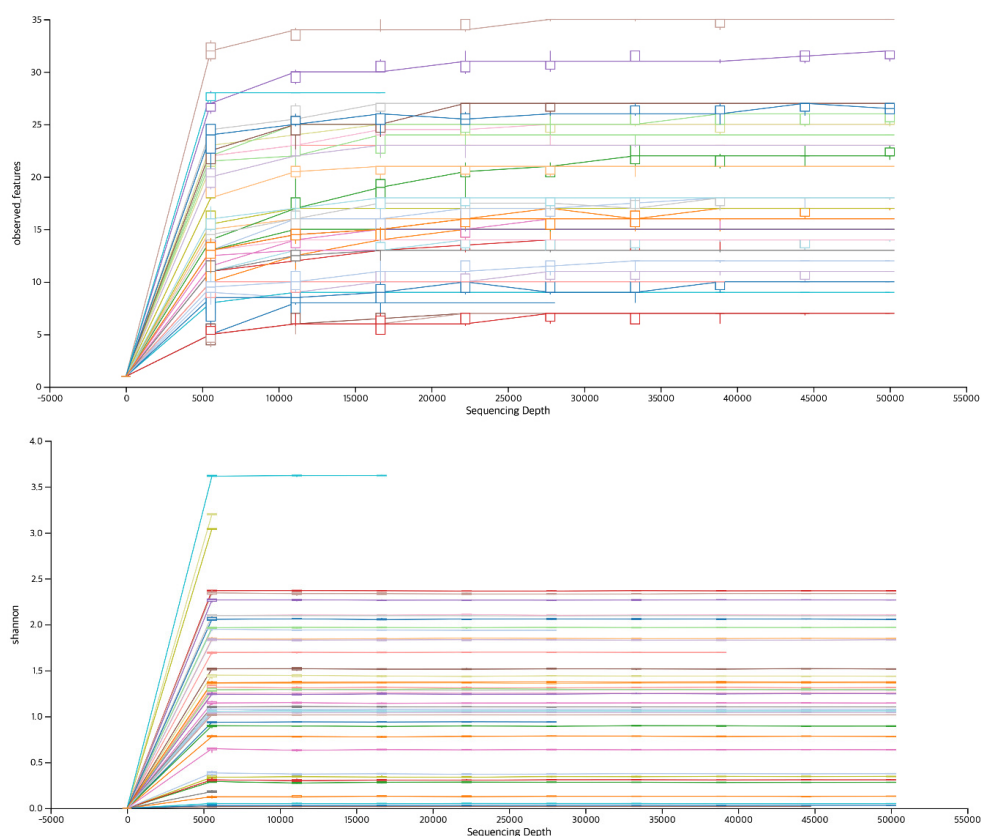


Fig. 1. Rarefaction curve of sequenced oral microbiome samples.

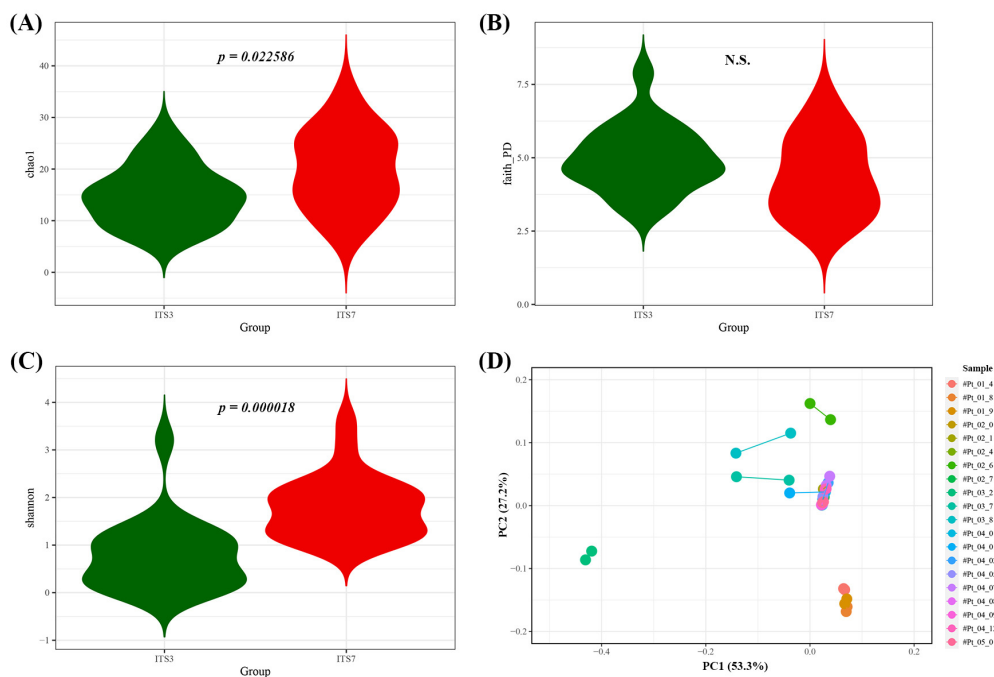


Fig. 2. Mycobiome community comparisons. Alpha diversity comparison between ITS3 primer and ITS7 primer. Chao1 alpha diversity (A), Faith's PD alpha diversity (B), and Shannon alpha diversity (C). (D) Beta diversity of each sample connected with line. Principal coordinate analysis (PCoA) of the Weighted UniFrac was performed to determine the mycobiome structure.

Species comparison

Next, phylogenetic tree was constructed to compare the species assigned by ITS3 primer and ITS7 primer. *Candida* formed the major tree which included *C. albicans*, *C. tropicalis*, and *C. parapsilosis*. Second tree included *Wickerhamomyces anomalus*, *Meyerozyma carbbica*, *Debaryomyces hansenii*, *C. glabrata* and *C. sake*. Another tree included *Clavispora lusitaniae* (*C. lusitaniae*), *C. intermedia* and *Yarrowia porcina* (*Y. porcina*). There were three more trees that were unidentified at the species level. Interestingly, there were several species that was only assigned by ITS3 primer which include *C. sake*, *C. intermedia*, *C. parapsilosis* and *Y. porcina* (Fig. 3).

Finally, relative abundance of each sample was plotted to observe the fungal abundance (Fig. 4). The most abundant species was *C. albicans* and the relative abundance showed similar proportion in most samples. However, the abundance of *C. lusitaniae* was assigned more abundant (9.98% vs 6.29%) using ITS3 primer in several samples ($p=0.047$). Also, *C. sake* (5/20), *C. intermedia* (8/20), *C. parapsilosis* (15/20) and *Y. porcina* (2/20) was only assigned by ITS3 primer. Although both primers assigned *Alternaria* at the genus level, *A. betae-kenyesis* was only assigned by ITS7 primer (10/20). Although not significant, the abundance of *Malassezia* species including *M. arunalokei* and *M. restricta* was assigned more abundant using ITS7 primer ($p=0.057$).



Fig. 3. Phylogenetic relationships among mycobiome sequenced with ITS3 and ITS7 primers. Primers are shown in shape and patient samples are shown in color. The size of the shapes indicates the relative abundance of the taxon in the sample.

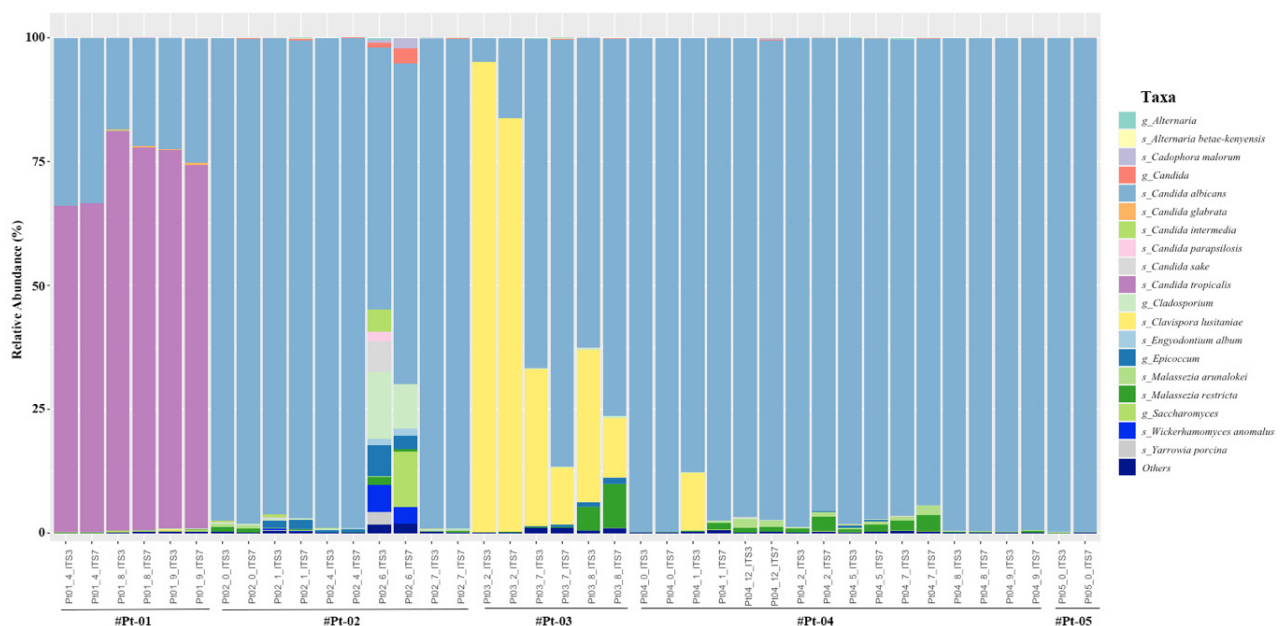


Fig. 4. Comparison of the relative abundance of the species produced by ITS3 primer and ITS7 primer in each sample.

DISCUSSION

While the biomass of the oral mycobiome is low in healthy subjects (2), mycobiome including *Candida* rapidly increases in intubated patients with reduced oral immunity and poor oral hygiene (5). To determine the mycobiome community, ITS region, which is located between the 18S and 26S rRNA genes, is one of the most frequently targeted for clinically important fungi (14). In this study, we tested two primer sets (ITS3/ITS4 and ITS7/ITS4) targeting ITS2 to evaluate their performance on oral mycobiome analysis.

Since same reverse primer was used, the read length was dependent on the forward primer and ITS3 primers (359.4 ± 47.3) produced significantly longer read length compared to ITS7 primers (311.5 ± 43.4). To determine the richness and evenness of the mycobiome, alpha diversity was analyzed. The richness and evenness of ITS7 primer were significantly higher at the OTU (operational taxonomy unit) level compared to ITS3 primers (Figs. 2B and 2C). However, there was no significant difference in Faith PD, which considers phylogenetic distance. To compare the mycobiome structure, beta diversity was determined using weighted UniFrac, which takes into account the phylogeny of the taxa. The respective oral samples showed nearby distribution suggesting the mycobiome structure was similar between ITS3 primer and ITS7 primer (Fig. 2D). Taken together, although ITS7 produced more OTUs, the overall performance in determining mycobiome structure was similar between ITS3 primer and ITS7 primer.

Finally, relative abundance was compared at the species level. In both primer set, the most abundant species was *C. albicans* and the relative abundance showed similar proportion in most samples. An increase in oral *C. albicans* has been shown to increase the risk of candidemia (23). Although *C. albicans* is the predominant cause of candidiasis, a shift toward non-albicans *Candida* species has been reported (24). Non-albicans *Candida* species have reduced susceptibility to antifungal drugs. Resistance to the azole group was more common in non-albicans *Candida* species compared to *C. albicans* (25). Thus, discriminating *C. albicans* from non-albicans *Candida* species is important for efficient management. Interestingly, there were several species that were only assigned by ITS3 primer including *C. sake*, *C. intermedia*, *C. parapsilosis* and *Y. porcina*. Moreover, the abundance of *C. lusitanae* was higher (9.98% vs 6.29%) using ITS3 primer. *C. lusitanae* can be isolated from environmental samples and it can rapidly develop resistance to multiple antifungal drugs

during treatment (26, 27). Taken together, ITS3 primer showed better performance in characterizing *Candida* species in oral cavity at the species level.

In this study, we compared the performance between two primers in predicting oral mycobiome in intubated patients. Other primers should also be performed for optimum prediction. Selecting optimum primers should provide better understanding of the mycobiome community in oral cavity.

ACKNOWLEDGEMENTS

This work was supported by a 2-year research grant of Pusan National University.

REFERENCES

- 1) Tiew PY, Mac Aogain M, Ali NABM, Thng KX, Goh K, Lau KJX, et al. The Mycobiome in Health and Disease: Emerging Concepts, Methodologies and Challenges. *Mycopathologia* 2020;185:207-31.
- 2) Diaz PI, Dongari-Bagtzoglou A. Critically Appraising the Significance of the Oral Mycobiome. *J Dent Res* 2021;100:133-140.
- 3) Labeau SO, Van de Vyver K, Brusselaers N, Vogelaers D, Blot SI. Prevention of ventilator-associated pneumonia with oral antiseptics: a systematic review and meta-analysis. *Lancet Infect Dis* 2011;11:845-54.
- 4) Alhazzani W, Smith O, Muscedere J, Medd J, Cook D. Toothbrushing for critically ill mechanically ventilated patients: a systematic review and meta-analysis of randomized trials evaluating ventilator-associated pneumonia. *Crit Care Med* 2013;41:646-55.
- 5) Krause R, Halwachs B, Thallinger GG, Klymiuk I, Gorkiewicz G, Hoenigl M, et al. Characterisation of *Candida* within the Mycobiome/Microbiome of the Lower Respiratory Tract of ICU Patients. *PLoS one* 2016;11:e0155033.
- 6) Strollo S, Lionakis MS, Adjemian J, Steiner CA, Prevots DR. Epidemiology of Hospitalizations Associated with Invasive Candidiasis, United States, 2002-2012. *Emerg Infect Dis.* 2016;23:7-13.
- 7) Mencarini J, Mantengoli E, Tofani L, Riccobono E, Fornaini R, Bartalesi F, et al. Evaluation of candidemia and antifungal consumption in a large tertiary care Italian hospital over a 12-year period. *Infection* 2018;46:469-76.
- 8) Baraniya D, Chen T, Nahar A, Alakwaa F, Hill J, Tellez M, et al. Supragingival mycobiome and inter-kingdom interactions in dental caries. *J Oral Microbiol* 2020;12:1729305.
- 9) Ghannoum MA, Jurevic RJ, Mukherjee PK, Cui F, Sikaroodi M, Naqvi A, et al. Characterization of the oral fungal microbiome (mycobiome) in healthy individuals. *PLoS Pathog* 2010;6:e1000713.
- 10) Dupuy AK, David MS, Li L, Heider TN, Peterson JD, Montano EA, et al. Redefining the human oral mycobiome with improved practices in amplicon-based taxonomy: discovery of *Malassezia* as a prominent commensal. *PLoS One* 2014;9:e90899.
- 11) Findley K, Oh J, Yang J, Conlan S, Deming C, Meyer JA, et al. Topographic diversity of fungal and bacterial communities in human skin. *Nature* 2013;498:367-70.
- 12) Imabayashi Y, Moriyama M, Takeshita T, Ieda S, Hayashida JN, Tanaka A, et al. Molecular analysis of fungal populations in patients with oral candidiasis using next-generation sequencing. *Sci Rep* 2016;6:28110.
- 13) Cui L, Lucht L, Tipton L, Rogers MB, Fitch A, Kessinger C, et al. Topographic diversity of the respiratory tract mycobiome and alteration in HIV and lung disease. *Am J Respir Crit Care Med* 2015;191:932-42.

- 14) Turenne CY, Sanche SE, Hoban DJ, Karlowsky JA, Kabani AM. Rapid identification of fungi by using the ITS2 genetic region and an automated fluorescent capillary electrophoresis system. *J Clin Microbiol* 1999;37:1846-51.
- 15) Fujita SI, Senda Y, Nakaguchi S, Hashimoto T. Multiplex PCR using internal transcribed spacer 1 and 2 regions for rapid detection and identification of yeast strains. *J Clin Microbiol* 2001;39:3617-22.
- 16) Ihrmark K, Bodeker IT, Cruz-Martinez K, Friberg H, Kubartova A, Schenck J, et al. New primers to amplify the fungal ITS2 region--evaluation by 454-sequencing of artificial and natural communities. *FEMS Microbiol Ecol* 2012;82:666-77.
- 17) Shelburne SA, Ajami NJ, Chibucos MC, Beird HC, Tarrand J, Galloway-Peña J, et al. Implementation of a Pan-Genomic Approach to Investigate Holobiont-Infecting Microbe Interaction: A Case Report of a Leukemic Patient with Invasive Mucormycosis. *PLoS One* 2015;10:e0139851.
- 18) Harmonization ICf. The international council fo harmonization of technical requirements for phamaceuticals for human use (ver.3) 2017. Available from: [http://www.ich.org/fileadmin/Public_Web_Site/ABOUT_ICH/Available from: Articles_Procedures/ICH_EWG_IWG_SOP_v3.0_final_22Jun2017-.pdf](http://www.ich.org/fileadmin/Public_Web_Site/ABOUT_ICH/Available%20from%20Articles_Procedures/ICH_EWG_IWG_SOP_v3.0_final_22Jun2017-.pdf).
- 19) Bonham PA. Swab cultures for diagnosing wound infections: a literature review and clinical guideline. *J Wound Ostomy Continence Nurs* 2009;36:389-95.
- 20) Hall M, Beiko RG. 16S rRNA Gene Analysis with QIIME2. *Methods Mol Biol* 2018;1849:113-29.
- 21) Nilsson RH, Larsson KH, Taylor AFS, Bengtsson-Palme J, Jeppesen TS, Schigel D, et al. The UNITE database for molecular identification of fungi: handling dark taxa and parallel taxonomic classifications. *Nucleic Acids Res* 2019;47:D259-64.
- 22) McMurdie PJ, Holmes S. phyloseq: an R package for reproducible interactive analysis and graphics of microbiome census data. *PLoS One* 2013;8:e61217.
- 23) Batista GC, Krebs VL, Ruiz LS, Auler ME, Hahn RC, Paula CR. Oral colonization: a possible source for candidemia in low-weight neonates. *J Mycol Med* 2014;24:81-6.
- 24) Sardi JCO, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJS. Candida species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J Med Microbiol* 2013;62:10-24.
- 25) Deorukhkar SC, Saini S, Mathew S. Non-albicans Candida Infection: An Emerging Threat. *Interdiscip Perspect Infect Dis* 2014;2014:615958.
- 26) Atkinson BJ, Lewis RE, Kontoyiannis DP. Candida lusitaniae fungemia in cancer patients: risk factors for amphotericin B failure and outcome. *Med Mycol* 2008;46:541-6.
- 27) Asner SA, Giulieri S, Diezi M, Marchetti O, Sanglard D. Acquired Multidrug Antifungal Resistance in Candida lusitaniae during Therapy. *Antimicrob Agents Chemother* 2015;59:7715-22.