

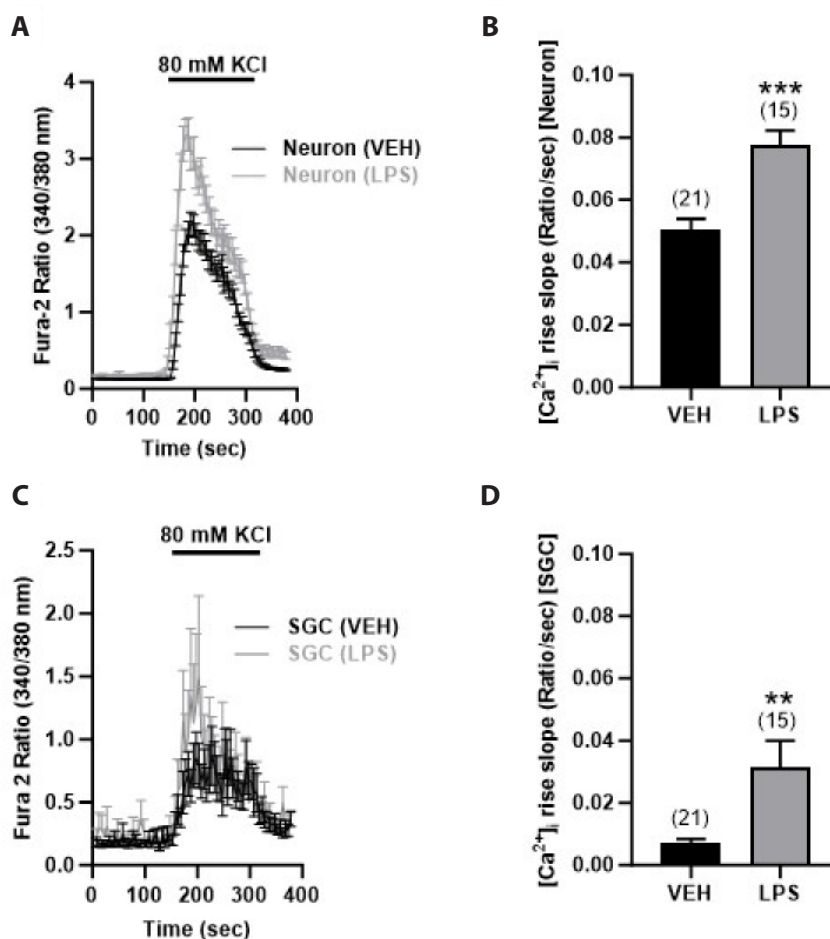
## SUPPLEMENTARY DATA

### Supplementary methods

#### Measurement of intracellular $\text{Ca}^{2+}$

In some experiments (for Supplementary Fig. 1), the initial kinetic rate of calcium influx was calculated as the  $[\text{Ca}^{2+}]_i$  rise slope (ratio/sec) [1,2]. The  $[\text{Ca}^{2+}]_i$  level was calculated as the difference in the peak Fura-2 AM ratio following 80 mM KCl treatment and the basal Fura-2 AM ratio ( $R_{\text{KCl peak}} - R_{\text{basal}}$ ). Then, the initial rate of KCl-induced calcium influx was calculated as the slope representing  $\text{Ca}^{2+}$  influx during the initial 30 sec following 80 mM KCl perfusion using the following formula:

$$[\text{Ca}^{2+}]_i \text{ rise slope (ratio/sec)} = (R_{\text{KCl peak}} - R_{\text{basal}}) / 30 \text{ sec}$$



**Supplementary Fig. 1. The effects of LPS on  $\text{Ca}^{2+}$  influx through VGCCs in SCG neurons.** (A, C) High  $\text{K}^+$  (80 mM)-induced increase in cytosolic  $\text{Ca}^{2+}$  in neurons and SGCs attached to the attendant neurons in culture. The relative concentration of cytosolic  $\text{Ca}^{2+}$  is expressed as a 340 nm/380 nm ratio. Twenty-four hours after LPS exposure (1  $\mu\text{g}/\text{ml}$ ), the high  $\text{K}^+$ -induced increase in cytosolic  $\text{Ca}^{2+}$  was significantly augmented in neurons and SGCs attached to the attendant neurons in culture. (B, D) Summary of the effects of LPS on high  $\text{K}^+$ -induced cytosolic  $\text{Ca}^{2+}$  increase in neurons and SGCs. Comparisons were made for the rising slope of cytosolic  $\text{Ca}^{2+}$  levels over time. The data are presented as means  $\pm$  SEM. The number of the tested cells from three independent experiments is indicated in parentheses. Unpaired Student's t-test, \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . LPS, lipopolysaccharide; VGCCs: voltage-gated  $\text{Ca}^{2+}$  channels; SCG, superior cervical ganglia; SGC, satellite glial cell.

## REFERENCES

1. Park CY, Shcheglovitov A, Dolmetsch R. The CRAC channel activator STIM1 binds and inhibits L-type voltage-gated calcium channels. *Science*. 2010;330:101-105.
2. Ross DG, Smart CE, Azimi I, Roberts-Thomson SJ, Monteith GR. Assessment of ORAI1-mediated basal calcium influx in mammary epithelial cells. *BMC Cell Biol*. 2013;14:57.