

of rituximab, bendamustine, and ibrutinib in patients with untreated and relapsed/refractory non-Hodgkin lymphoma. *Blood* 2015;125:242-8.

12. Sauter CS, Matasar MJ, Schoder H, et al. A phase 1 study of ibrutinib in combination with R-ICE in patients with relapsed or primary refractory DLBCL. *Blood* 2018;131:1805-8.
13. Sivam V, Cook L, Hughes G, et al. Gemcitabine and vinorelbine chemotherapy for refractory or relapsing aggressive non-Hodgkin lymphoma. *Hematol Oncol* 2012;30:214-5.

Successful recovery of poor graft function by administration of romiplostim in a multiple myeloma case with poor graft function following autologous stem cell transplantation

TO THE EDITOR: Despite advances in treatment options, including the use of novel agents, such as proteasome inhibitors, immunomodulatory drugs, and monoclonal antibodies, autologous hematopoietic stem cell transplantation (auto-HSCT) remains the cornerstone of treatment of multiple myeloma (MM) [1]. Following marrow suppression by high-dose chemotherapy as a prelude to auto-HSCT, most patients achieve hematopoietic recovery after autologous stem cell infusion. However, some patients unexpectedly develop poor graft function (PGF). Although there is no universally accepted definition of PGF after auto-HSCT, the European Society for Blood and Marrow Transplantation (EBMT) guidelines suggest defining PGF as two or three cytopenia lasting for at least 2 weeks, 28 days after hematopoietic stem cell transplantation [2]. In a recent prospective study, PGF was defined as sustained cytopenia, defined as platelet count $\leq 50,000/\mu\text{L}$, and/or hemoglobin $\leq 8 \text{ g/dL}$, and/or absolute neutrophil count (ANC) $\leq 1,000/\mu\text{L}$ [3].

PGF broadly encompasses several rare disease conditions that are characterized by continued requirements for blood transfusions or an increased risk of infection due to neutropenia, such as post-transplantation immune thrombocytopenia purpura (ITP) or bone marrow aplasia [4, 5]. Therefore, persistent PGF can negatively impact the quality of life after auto-HSCT. Moreover, it may limit subsequent treatments, such as maintenance or consolidation therapies, and adversely affect disease control. Despite these potential consequences, no systematic approach for diagnosing and managing PGF is known currently.

Thrombopoietin (TPO) receptor agonists (TPO-RAs) enhance the production of hematopoietic stem cells and megakaryocytes by stimulating the TPO receptor, also known as myeloproliferative leukemia protein. Two TPO-RAs, eltrombopag and romiplostim, have been approved by the Food and Drug Administration to treat cytopenia in con-

ditions, such as ITP and aplastic anemia [6, 7]. Owing to their mechanism of action of stimulating hematopoietic stem cell production, TPO-RAs are currently being explored for the treatment of PGF following auto-HSCT [5]. However, the existing reports have primarily provided descriptive information, often involving small cohorts. Moreover, studies of post-transplantation PGF have predominantly focused on allogeneic hematopoietic stem cell transplantation.

In this report, we present the case of a patient with MM who successfully recovered from PGF following auto-HSCT by treating with romiplostim. This recovery was achieved even after the initial TPO-RA (eltrombopag) was ineffective. This study was approved by the Institutional Review Board of the Catholic University of Korea (KC22 RISI0540) and was conducted in accordance with the Declaration of Helsinki.

CASE

A 62-year-old woman was diagnosed with transplant-eligible MM. The patient underwent a predetermined therapeutic approach, which included six cycles of induction chemotherapy followed by auto-HSCT. The induction chemotherapy regimen consisted of bortezomib, thalidomide, and dexamethasone. She achieved complete response at 15 weeks after initiation of the therapeutic approach. Bone marrow biopsy before auto-HSCT revealed stringent CR (Fig. 1D). To prepare auto-HSCT, stem cells were mobilized using $10 \mu\text{g/kg/d}$ of G-CSF for 5 days and of 375 mg/m^2 of etoposide for 1 d. For pretreatment with auto-HSCT, the patient received 3.2 mg/kg/d busulfan for 3 days and $200 \text{ mg/m}^2/\text{d}$ thiotepa for 2 days [8]. Subsequently, $\text{CD}34^+$ cells at a dose of $4.02 \times 10^6/\text{kg}$ were infused for auto-HSCT. No post auto-HSCT maintenance or consolidation treatments were administered. The patient remained free of clinically significant adverse events for 9 months after auto-HSCT.

At 285 days post-auto-HSCT, the patient's platelet count unexpectedly dropped to $5 \times 10^9/\text{L}$, while hemoglobin concentration remained at 11.4 g/dL , and the absolute neutrophil count (ANC) was $1.08 \times 10^9/\text{L}$, which was within a clinically nonsignificant range (Fig. 2). We were unable to identify any potential causes of thrombocytopenia, such as graft failure, MM progression, secondary myeloid neoplasm, infection, or toxic drug effects. The diagnosis that we arrived at was post-auto-HSCT PGF, supported by a bone marrow study that revealed 15% cellularity without cytogenetic abnormalities, malignant cancer cells, fibrosis, or dysplastic changes (Fig. 1G). Considering the prominence of thrombocytopenia within the spectrum of PGF, we initiated treatment with oral dexamethasone (40 mg/d for 4 d) and intravenous immunoglobulin (1 g/kg/d for 2 d) following therapeutic indications for ITP [9]. Regrettably, these interventions resulted in further neutropenia ($0.65 \times 10^9/\text{L}$), and no platelet recovery ($22 \times 10^9/\text{L}$) was observed 189 days after the initiation of high-dose dexamethasone and intravenous immunoglobulin. Consequently, we revised the diag-

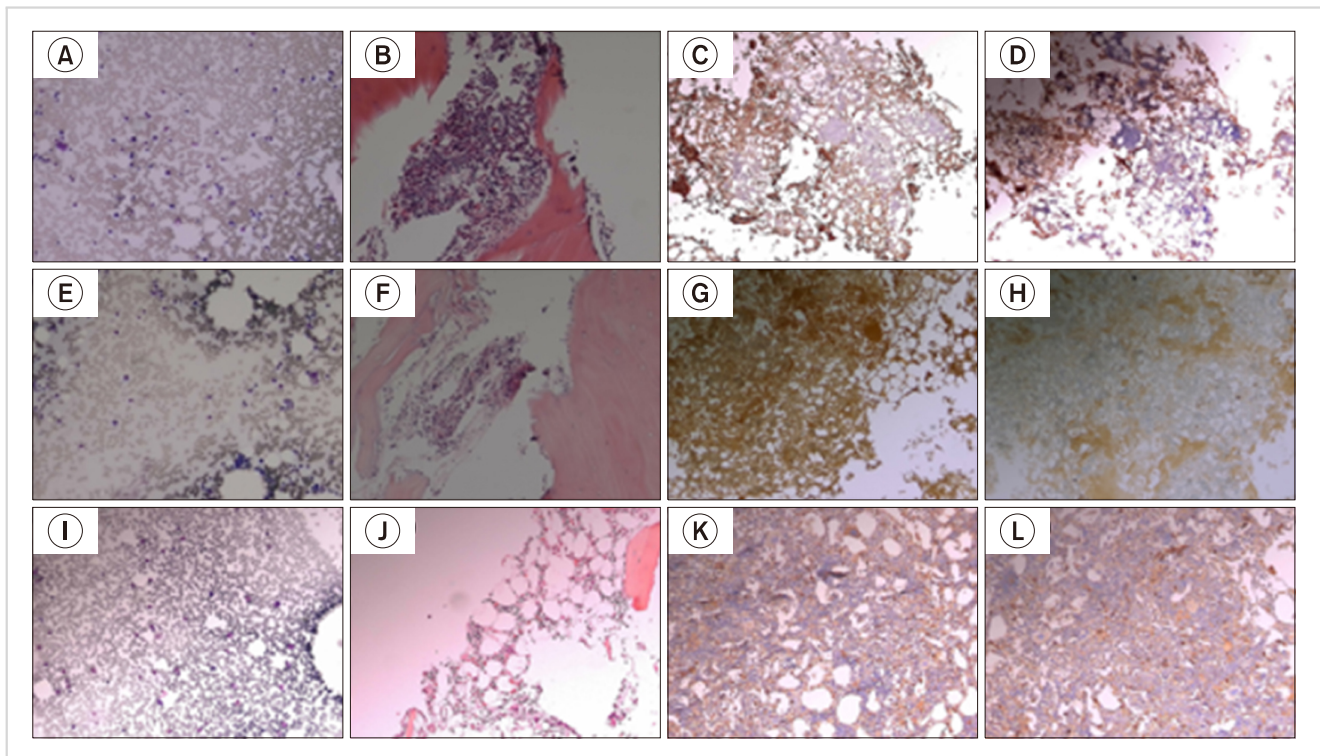


Fig. 1. Images of bone marrow at different time points: before autologous stem cell transplantation (ASCT, upper row: **A–D**); at 282 days after ASCT, when the patient was diagnosed with poor graft function (middle row: **E–H**); and at 138 weeks after initiation of romiplostim, demonstrating stringent complete response of multiple myeloma with no evidence of fibrosis or dyspoiesis (lower row: **I–L**). The first column (**A**, **E**, and **I**) presents bone marrow aspiration, magnified $\times 100$; the second column (**B**, **F**, and **J**) illustrates bone marrow biopsy, magnified $\times 100$; the third column (**C**, **G**, and **K**) shows kappa staining, magnified as $\times 40$ to $\times 100$; the fourth column (**D**, **H**, and **L**) shows lambda staining magnified $\times 40$ to $\times 100$.

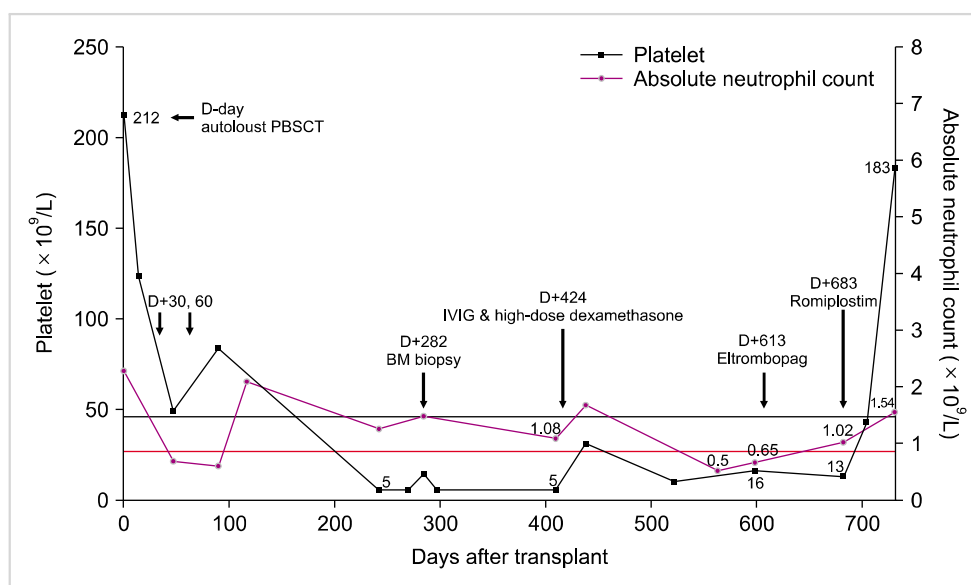


Fig. 2. Dynamics of platelet count (unit, $\times 10^9/L$) and absolute neutrophil count (unit, $\times 10^9/L$) following autologous hematopoietic stem cell transplant and the response to romiplostim.

nosis to post-HSCT PGF rather than ITP, and initiated eltrombopag at a dose of 25 mg/d. However, even with escalated doses of up to 75 mg/day, the platelet count did not recover beyond $22 \times 10^9/L$. Subsequently, the patient expressed a preference for romiplostim instead of increasing

the off-label eltrombopag dose (eltrombopag, indicated for ITP, is approved for use in South Korea at a maximum dose of 50 mg).

Given the failure of eltrombopag and uncontrolled PGF with ANC $1.02 \times 10^9/L$ and a platelet count of $13 \times 10^9/L$, we

switched from the eltrombopag to romiplostim, initiating romiplostim at a dose of 1 µg/kg per week. Six weeks after initiation, despite using a low dose of romiplostim (1 µg/kg) without dose escalation, PGF was completely resolved, with an ANC of $1.54 \times 10^9/L$, a platelet count of $183 \times 10^9/L$, and hemoglobin concentration of 12.5 g/dL (Fig. 2). At 138 weeks after the initiation of romiplostim treatment, restored blood counts were adequately maintained. The bone marrow study showed no minimal residual disease based on 8-color flowcytometry nor other dyspoietic findings, as observed in the morphology and cytogenetic analyses (Fig. 1H-K).

DISCUSSION

Understanding PGF has been challenging owing to varying definitions across publications, and reports of PGF after auto-HSCT are rare. PGF occurs in 5–27% of patients after allogeneic hematopoietic stem cell transplantation and in approximately 2% of patients following auto-HCT, representing a clinically significant unmet need [3]. EBMT guidelines define PGF as “Two or three cytopenia >2 weeks after 28 days in the presence of donor chimerism >5%”, regardless of whether the donor cells are autologous or allogeneic [2]. In the context of auto-HSCT, donor chimerism should theoretically be calculated as 100%. However, specific cutoff levels for each cytopenia are not specified, and establishing cutoff levels for cytopenia after auto-HSCT poses a challenge due to the limited literature. To the best of our knowledge, there is only one large-scale retrospective study by Lutfi *et al.* [10] and one prospective study by McGuirk *et al.* [3], both defining PGF as platelet count $\leq 50,000/\mu L$, and/or Hb ≤ 8 g/dL, and/or ANC $\leq 1,000/\mu L$ persisting after auto-HSCT. In our case, PGF, which is distinct from primary graft failure involving failure of the initial engraftment, was ruled out, as the initial engraftment was successful. The lowest ANC in our case was 0.65×10^9 , surpassing the threshold of 0.5×10^9 for secondary graft failure, thereby further excluding this diagnosis [2]. After excluding other potential causes of cytopenia, such as disease relapse, fibrotic/dysplastic changes, and toxic drug effects, we ultimately diagnosed the patient with PGF. The proposed factors contributing to PGF after auto-HSCT include CMV status, weight, and pre-transplant hemoglobin/hematocrit [10].

Although various treatment options such as CD34+ stem cell infusion, TPO-RAs, and mesenchymal stem cell infusion [11], there is no established standard treatment for PGF. Guidelines for treatment strategies, especially after the failure of one TPO-RA, are lacking. Nevertheless, switching to a different TPO-RA, as demonstrated in our case, is supported by previous reports involving ITP or AA, where effective responses were observed when eltrombopag was replaced with romiplostim and vice versa [12, 13]. The mechanism underlying the improved response with different TPO-RAs is not fully understood, but may involve differences in dosage, pharmacokinetics, and pharmacodynamics. For example, the Asian population typically has lower el-

trombopag clearance and is often treated at a maximum dose of 75 mg/d than Caucasians at 150 mg/d [14]. Another explanation could be bone marrow fibrosis induced by eltrombopag, which may have been reversed after the discontinuation or late benefits of eltrombopag treatment.

In conclusion, TPO-RAs can be considered for the treatment of PGF following auto-HSCT, and switching to another TPO-RA may be a viable strategy even if one TPO-RA fails.

Jeongmin Yim¹, Sung-Soo Park¹, Jong-Mi Lee²,
Jae-Ho Yoon¹, Hee-Je Kim¹, Chang-Ki Min¹

Departments of ¹Hematology, ²Laboratory Medicine, Seoul
St. Mary's Hospital, College of Medicine, The Catholic
University of Korea, Seoul, Korea

Correspondence to: Sung-Soo Park

Department of Hematology, Seoul St. Mary's Hospital,
College of Medicine, The Catholic University of Korea, 222
Banpo-daero, Seocho-gu, Seoul 06591, Korea
E-mail: sspark@catholic.ac.kr

Received on Sep. 29, 2023; Revised on Nov. 12, 2023; Accepted on Nov. 24, 2023

<https://doi.org/10.5045/br.2023.2023185>

Acknowledgments

J.Y. reviewed and wrote the manuscript. J.Y. and S.S.P. conceived of and designed the study. J.M.L. reviewed the bone marrow images. All the authors reviewed the drafted manuscript, namely J.Y., S.S.P., J.M.L., J.H.Y., H.J.K., and C.K.M. The authors acknowledge all members of the Catholic Hematology Hospital, particularly the house staff, for their excellent patient care.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

REFERENCES

1. Jung SH, Jo JC, Song GY, et al. Frontline therapy for newly diagnosed patients with multiple myeloma. *Blood Res* 2020;55(S1): S37-42.
2. Valcárcel D, Sureda A. Graft failure. In: Carreras E, Dufour C, Mohty M, Kröger N, eds. *The EBMT handbook: hematopoietic stem cell transplantation and cellular therapies*. 7th ed. Cham: Springer, 2019:307-13.
3. McGuirk JP, Metheny L 3rd, Pineiro L, et al. Placental expanded mesenchymal-like cells (PLX-R18) for poor graft function after hematopoietic cell transplantation: a phase I study. *Bone Marrow Transplant* 2023;58:1189-96.
4. Hequet O, Salles G, Ketterer N, et al. Autoimmune thrombocytopenic purpura after autologous stem cell transplantation. *Bone Marrow Transplant* 2003;32:89-95.
5. Bortolotti M, Pettine L, Zaninoni A, Croci GA, Barcellini W, Fattizzo B. Efficacy and immunomodulating properties of el-

trombopag in aplastic anemia following autologous stem cell transplant: case report and review of the literature. *Pharmaceuticals (Basel)* 2022;15:419.

6. Young NS. Aplastic anemia. *N Engl J Med* 2018;379:1643-56.
7. Jang JH, Tomiyama Y, Miyazaki K, et al. Efficacy and safety of romiplostim in refractory aplastic anaemia: a phase II/III, multi-centre, open-label study. *Br J Haematol* 2021;192:190-9.
8. Song GY, Jung SH, Kim JS, et al. Busulfan and thiotepea as a conditioning regimen for autologous stem cell transplantation in patients with multiple myeloma: a study of the Korean Multiple Myeloma Working Party (KMMWP-1801 study). *Front Oncol* 2022;12:959949.
9. Park YH, Kim DY, Kim S, et al. Management of immune thrombocytopenia: 2022 update of Korean experts recommendations. *Blood Res* 2022;57:20-8.
10. Lutfi F, Skelton WP, Rosenau E, et al. Poor graft function following autologous stem cell transplant (ASCT): a retrospective analysis over two decades at a transplant center. *Blood* 2017;130 Suppl 1:4529.
11. Prabahan A, Koldej R, Chee L, Ritchie D. Clinical features, pathophysiology, and therapy of poor graft function post-allogeneic stem cell transplantation. *Blood Adv* 2022;6:1947-59.
12. González-Porras JR, Mingot-Castellano ME, Andrade MM, et al. Use of eltrombopag after romiplostim in primary immune thrombocytopenia. *Br J Haematol* 2015;169:111-6.
13. Ise M, Iizuka H, Kamoda Y, Hirao M, Kida M, Usuki K. Romiplostim is effective for eltrombopag-refractory aplastic anemia: results of a retrospective study. *Int J Hematol* 2020;112:787-94.
14. Ahn HJ, Byun JM, Kim I, et al. Eltrombopag for post-transplant poor graft function in East Asian patients. *J Korean Med Sci* 2022;37:e48.

Clinico-pathological features and treatment outcomes of high-grade B cell lymphoma—a tertiary cancer center experience

TO THE EDITOR: High-grade B-cell lymphoma (HG-BCL), an uncommon condition, displays a blastoid morphology and an aggressive disease course, but lacks the features of Burkitt's lymphoma or diffuse large B-cell lymphoma [1, 2]. Previously termed "B cell lymphoma having features intermediate between DLBCL and Burkitt's lymphoma" [3], its rarity has primarily resulted in descriptions of isolated case series.

We conducted a retrospective study on 14 patients diagnosed with HG-BCL who were registered in our department between January 1, 2020 and April 30, 2022. Their clinicopathological variables and outcomes at the end of the follow-up period are outlined in the Results.

RESULTS

Epidemiology

The median age was 55.5 (range, 29–7) yr. Most patients were men (N=10, 71.4%). The common comorbidities were diabetes (N=3, 21.4%) and hypertension (N=2, 14.3%). The median symptom duration before diagnosis was eight (range, 1–20) wk. Seven (50%) patients did not have any B symptoms, whereas weight loss (N=3, 21.4%) and fatigue (N=4, 28.6%) were observed in the remaining seven (50%).

Disease workup

Bone marrow biopsy showed disease involvement in six (42.9%) patients. Central nervous system (CNS) involvement was confirmed in 1 out of 14 (7.1%) by a positive cerebrospinal fluid cytology. Four patients (28.6%) showed involvement of the spleen. Extra nodal site involvement was categorized as 1 site (N=7, 50%), ≥2 sites (N=4, 28.6%), and none (N=3, 21.4%). The median IPI and CNS IPI score was 3. High IPI (score of 4–5) and High CNS IPI (score of 4–6) were observed in three (21.4%) patients each.

The common stages observed were stages IV (N=8, 57.1%) and II (N=3, 21.4%). On IHC, five (35.6%) patients were found to be double-expressors of the markers C-MYC, BCL-2, and BCL-6. On analysis by FISH, C-MYC rearrangement was found in two (15.4%) patients with none having concurrent rearrangement of BCL-2 or BCL-6, as shown in [Table 1](#).

Therapy

Nine (64.2%) patients received R-CHOP and five (35.7%) received R-DA-EPOCH as first-line therapy. CNS prophylaxis with intrathecal-methotrexate (IT-MTx) was administered concurrently with the R-DA-EPOCH regimen in all five patients, whereas five out of nine patients treated with R-CHOP received the same. High-dose MTx was administered to the remaining four patients on R-CHOP. Three (21.4%) patients relapsed and received salvage chemotherapy with R-GDP (N=2) and R-DHAP (N=1) regimens.

Treatment outcome

The median follow-up duration was 10.5 (1.13–19.3) mo. Response rates after first-line therapy were: complete response (CR), 71.4% (N=10); partial response (PR), 14.2% (N=2); stable disease (SD), none, and progressive disease (PD), 14.2% (N=2), with overall response rates (ORR) of 85.7%. One patient progressed rapidly on first line and died soon after starting therapy. Of the three (21.4%) patients who relapsed, two (14.2%) died from disease progression. The third death was due to COVID-19 pneumonia ([Table 2](#)).

Survival

The mean progression-free survival (PFS) and overall survival (OS) were 14.86 (median, not reached) and 15.58 (median, not reached) mo, respectively. One-year PFS and OS were both 76.2%.