

Role of Peroxisome Proliferator-Activated Receptor (PPAR) δ in Embryonic Stem Cell Proliferation

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The peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors that belong to the nuclear receptor family. It is well known that PPARs function as regulators of lipid and lipoprotein metabolism and glucose homeostasis, as well as influence cellular proliferation, differentiation and apoptosis. However, the role of the PPARs with regard to embryonic stem (ES) cells remains unknown. We will review the function of the PPAR δ , one of the three PPAR isoforms, α , δ (also called β/δ), and γ , in ES cells and its role in embryo development. In addition, pluripotent mouse ES cells can be expanded in large numbers *in vitro* due to the process of symmetrical self-renewal. Here we describe how PPAR δ sustains ES cell proliferation.

Keywords: PPAR δ , ES cells, Proliferation, High glucose

Introduction

Peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors that belong to the nuclear receptor superfamily (1). They have been identified in a variety of different species including the xenopus, mouse, rat, and humans (2). After the isolation of PPAR α (NR1C1), as the receptor mediating peroxisome proliferation in rodent hepatocytes in 1990 (3), two related isotypes, PPAR δ (NR1C2; also called PPAR β/δ) and PPAR δ (NR1C3) located on chromosomes 15, 17 and 6 in the mouse and chromosomes 22, 6 and 3 in human, were identified (4, 5). Once activated by their respective ligands, the PPARs control the transcription rate of a large panel of genes implicated in various physiological functions, including adipogenesis, lipid and glucose homeostasis, inflammation, cell proliferation, differentiation,

and carcinogenesis (2, 6-8). PPARs heterodimerize with the retinoid X receptors and modulate gene expression of target genes containing peroxisome proliferators-responsive elements in response to ligand activation (9, 10).

The three isoforms of PPARs display distinct physiological and pharmacological activity that is dependent on their target genes and their tissue distribution (11, 12). PPAR α , activated by polyunsaturated fatty acids and leukotriene B₄, is expressed in tissues with high fatty acid catabolism such as the liver, heart, brown adipose tissues, kidney, and intestine. PPAR γ , mainly expressed in adipocytes, macrophages, placenta, and other tissues, is activated by specific fatty acid metabolites, such as 15-deoxy-prostaglandin J₂ (15d-PGJ₂), and by thiazolidinediones. Both PPAR α and PPAR γ response genes are involved in lipid homeostasis. Therefore, it is not surprising that the main functions of PPAR α and PPAR γ are related to glucose and lipid homeostasis (13-15). On the other hand, the ubiquitous distribution of PPAR δ , (although gut, kidney, and heart express higher levels than other tissues) makes it difficult to associate PPAR δ with a specific biological function (14). Although PPAR δ is the least studied PPAR, it has been reported that PPAR δ is associated with a diverse range of functions. Indeed, PPAR δ partic-

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ipates in many biological processes, including lipid and glucose metabolism (16-19), epidermal maturation and wound healing (20-22), muscle development and function (23-25), tumorigenesis (26-30), inflammation (31, 32), and cytoprotection (33). In addition, combining multiple research approaches, PPAR δ has been implicated in embryo development including stem/progenitor cell proliferation (34, 35), embryo implantation (13, 36, 37), embryo organogenesis, and diabetic embryopathy (38, 39). Therefore, this review will focus on the role of PPAR δ in the ES cells.

Physiological functions of PPAR δ

PPAR δ has been the most elusive among the three PPAR subtypes. Due to its broad tissue distribution, it is difficult to identify a specific function for this receptor. PPAR δ has a broad expression pattern in adult animals, and is detected very early during embryogenesis (40). Several studies have shown that PPAR δ is activated by a large variety of ligands and is implicated in the developmental and metabolic regulation of several tissues. PPAR δ activators include fatty acids (41, 42), triglycerides (43), the cyclooxygenase (COX) product, prostacyclin (42), the COX/prostacyclin synthase derived endocannabinoid metabolites (44), and all transretinoid acid (45). A number of synthetic PPAR δ ligands have been described including GW0742X, GW2433, GW9578, L-783, 483, GW501516, L-796,449, L-165,041, and compound F (46-48). In addition, GW501516 and GW0742 activate PPAR δ at very low concentrations both *in vivo* and *in vitro* with a 1,000-fold selectivity over the other PPAR subtypes (49).

A mouse knockout model, although difficult to generate due to highly penetrant lethality (21, 50), indicated a role for PPAR δ during embryo implantation, as well as in myelination of the brain, lipid metabolism and adiposity, and epidermal cell proliferation (37, 50, 51). In addition, PPAR δ has been linked to cell differentiation, inflammation, cell motility, and cell growth (9, 52). Recent studies suggest that PPAR δ plays a role in cell growth. For example, the PPAR δ expression is increased in colorectal cancer cells compared with normal colon epithelial cells (30, 53). Treatment of GW501516, a PPAR δ ligand, increased the number and size of intestinal polyps (29). In addition, PPAR δ has been implicated in the growth of several other cell types including vascular smooth muscle cells (54), preadipocytes (17, 55) and epithelial cells (9). It has recently become clear that PPAR δ has a function in epithelial tissues; however, this role continues to be debated

in reports with inconsistent findings. Indeed, some reports suggest that ligand activation of PPAR δ potentiates cell growth (56), whereas other reports suggest that ligand activation of PPAR δ attenuates cell growth (57). Activation of PPAR δ by its agonist increases COX2 expression in human hepatocellular carcinoma cells (58). In addition, PPAR δ has been shown to mediate prostaglandin E₂ (PGE₂)-induced cell growth (59). More importantly, several reports have suggested that PPAR δ -mediated cell growth is induced by COX/prostaglandin (PG) signal pathways. Consequently, the interplay between the PPAR δ and cytosolic phospholipase A₂ (cPLA₂)/COX2/PGE₂ signaling pathways acts as a positive regulator in cell growth.

Apart from cell growth, the fact that PPAR δ plays an important role in embryonic development is supported by the observation that most PPAR δ -deficient mice die early during embryonic development due to a placental defect. A recent study showed that the genetic loss of PPAR δ signaling does not influence ovulation, fertilization or preimplantation (60). Mouse embryos express PPAR δ detectable at the two-cell stage (61) or eight-cell stage (36), and throughout the preimplantation period. Mouse blastocysts also express PPAR δ in the inner cell mass and the trophoblast. On the embryonic cell level, prostacyclin or the PPAR δ agonist increased the embryonic cell mass, indicating that PPAR δ is essential for embryo development, blastocyst hatching and implantation (36).

PPAR δ and embryonic stem (ES) cells

The elucidation of PPAR δ modulation of ES cells will provide new insights into embryonic development. In addition, research on how normal embryonic development is regulated will provide new clues as to how to maintain stem cells in culture. The description of the interaction among signal molecules is a key to our fundamental understanding of stem cell proliferation and its translation into therapeutic strategies. However, information regarding the potential role of PPAR δ in physiological and/or developmental processes is very limited, although PPAR δ is widely expressed in embryonic tissue. There is evidence that PPAR δ can modulate stem and progenitor cell expansion and a differentiated phenotype. In neural stem cells, PPAR δ contributes to the maintenance of the undifferentiated and proliferative status, by regulating both the genes involved in cell cycle control, as observed in other cell types (54, 62, 63), and inhibiting the activity of the other PPARs, which may be involved in cellular differentiation (64-66). In addition, the PPAR δ agonist GW

501516 has been shown to be a promoter involved in the development of adenocarcinomas with high expression of stem cell markers CK19 and Notch1, as well as Proliferin, a growth factor that mediates many of the effects of the stem cell markers such as Musashi1, in mammary cells (67). PPAR δ is expressed in the crypt cells of the small intestine and negatively regulates Hedgehog signaling to block differentiation (68), a process that would be expected to promote transformation. The association of Wnt activation with stem cell expansion, activa-

tion of β -catenin/T-cell factor (TCF) signaling by 3-phosphoinositide-dependent protein kinase 1 (PDK1) and the identification of PPAR δ responsive genes suggest a common mechanism for the tumor promoting action of PPAR δ agonists that may involve stem and progenitor cell proliferation (35).

For successful implantation and pregnancy, recent evidence suggests that the implantation timing of PG signaling resulting from cPLA₂, COX2 or lysophosphatidic acid receptor 3 plays an important role in the subsequent developmental processes (69-72). However, the underlying mechanism and the molecular link between the critical steps are still unclear. A previous report provided evidence that PPAR δ serves as a molecular link that coordinates multiple signaling pathways in mouse ES cell proliferation allowing their self-renewal (Fig. 1) (34). In that study, high glucose (25 mM) increased PPAR δ gene expression rather than PPAR α or PPAR γ in the ES cells. In addition, the PPAR δ agonist, L-165,041 increased ES cell proliferation, but the PPAR δ antagonist, GW9662 or the PPAR δ specific small interfering RNAs inhibited the effects of high glucose. Moreover, high glucose increased COX2 and PGE₂ synthesis activating PPAR δ , which increases cell cycle regulatory protein expression such as cyclins and cyclin dependent kinases (CDKs). It has been consistently shown that COX2-derived PGE₂ and PGI₂ mediate their function via PPAR δ receptors during the early steps of decidualization in mice (73). Although the exact mechanisms involved in COX2-derived PG activation of PPAR δ has not been completely elucidated, previous reports have suggested that the activity of PGE₂ is mediated by possible activation of the EP₂ receptor, which increases the cAMP levels (74-76) and PPAR δ receptors. Previous work has shown that embryos from streptozotocin (STZ)-induced diabetic rats have diminished PGE₂ content, although they can produce PGE₂ in large amounts (77). One can speculate that arachidonic acid (AA) might be depleted if PGE₂ generation and release is increased in the diabetic embryo in order to maintain the intracellular PGE₂ levels (39). In addition, it has been suggested that ligand activation of PPAR δ induces the expression of COX2 (56), which could theoretically promote cell growth and inhibit apoptosis through mechanisms that involve the production of prostaglandins. These data raise the possibility that impaired activation of PPAR δ may alter the lipid signaling required for normal self-renewal of ES cells, which raises that possibility that PPAR δ might be a putative target for the maintenance of ES cell characteristics. The results of a previous study suggested that the loss of PPAR δ leads to reduction of the phosphorylation

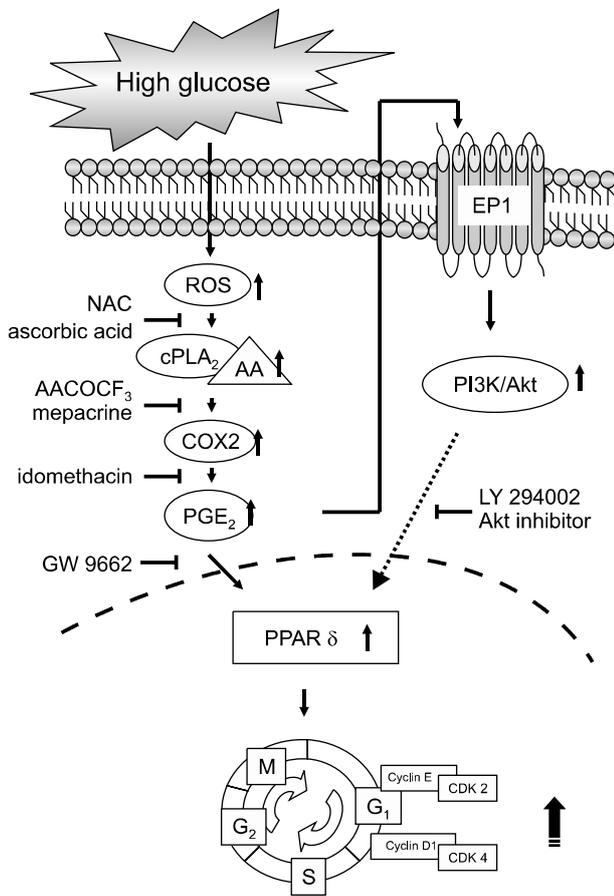


Fig. 1. The hypothesized model for the signal pathways involved in high glucose-induced mouse ES cell proliferation. High glucose increased PGE₂ synthesis, which is controlled by the coupled activation of cPLA₂/AA and COX2 via ROS. After PGE₂ is released into the extracellular space, it binds to the membrane coupled EP receptors. This effect is mediated, at least in part, by the activation of PI3K/Akt. Finally, these molecules may induce PPAR δ , which increases cell cycle regulatory protein expression levels. Abbreviations: AA, arachidonic acid; CDK, cyclin-dependent kinase; COX2, cyclooxygenase-2; cPLA₂, cytosolic phospholipase A₂; NAC, N-acetylcysteine; PGE₂, prostaglandin E₂; PI3K, phosphoinositide 3-kinase; PPAR, peroxisome proliferator activated receptor; ROS, reactive oxygen species (34).

status of Akt and STAT3 in the trophoblast (60). Akt is a down-stream pathway of PPAR δ signaling that is active during cell proliferation and survival (60); it was observed that PPAR δ null mice were not able to progress through the normal developmental steps. The PI3K/Akt signaling pathway also has been implicated in ES cell self-renewal in studies of ES cells without PTEN (78). Thus, this observation supports the participation of PPAR δ in ES cell proliferation and maintenance of self-renewal. In addition, several groups have shown that STAT3 is an important signal transducer and activator in the maintenance of pluripotency and the propagation of mouse ES cells (79-81). Although STAT3 phosphorylation with LIF was not influenced by the PPAR δ $-/-$ trophoblast, the STAT3 was not phosphorylated by the PPAR δ agonist. This suggests that PPAR δ activity not only plays a role in normal development but also is involved in ES cell proliferation and self-renewal. It has been shown that STAT3 could direct the expression of key regulators of the mitotic cycle in ES cells and stimulates their entry into the S phase (79). Thus, it is possible that PPAR δ plays a role in inducing cell cycle molecules that are involved in ES cell proliferation. Gene expression profiling experiments will help gain insights into the mechanisms involved in PPAR δ activity during the process of self-renewal.

Conclusions

Over the past few years, knowledge of the physiological activity of PPAR δ has expanded. The study of PPAR δ characteristics has added to improve understanding of cell physiology. PPAR δ has been implicated in many cell processes, from the embryo to adult cells, and from cell proliferation to cell differentiation; it has been shown to be crucial for energy homeostasis. Until recently, because the function of PPAR δ remained elusive, the therapeutic potential of PPAR δ agonists for lipid and glucose metabolism, embryo development and wound healing has been tested in mice only. Our current understanding of PPAR δ has demonstrated that PPAR δ also plays a critical role in ES cell proliferation. Future studies will likely clarify the physiological role of PPAR δ in ES cell growth and differentiation.

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Potential Conflict of Interest

The authors have no conflicting financial interests.

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