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Letter to the Editor: Depending on the Disease Stage and Modifying Factors, mtDNA-Associated Hearing Loss Can Occur With Many mtDNA Mutations

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► See the article “Prevalence and Clinical Characteristics of Mitochondrial DNA Mutations in Korean Patients With Sensorineural Hearing Loss” in volume 38, number 48, e355.

To the Editor:

We read with interest Joo et al.'s article¹ on a prospective, single-centre, study of the frequency of six mitochondrial DNA (mtDNA) variants known to be associated with hearing loss (HL) in a cohort of 1,099 patients from 711 Korean families with pre- or post-lingual HL. It was found that 12 of the 1,099 patients carried either the m.1555A>G variant (n = 10) or the m.7444G>A variant (n = 2).¹ It was concluded that a significant proportion of Korean patients with HL carried mtDNA variants with m.1555A>G being the most common, therefore there is a need for genetic testing in patients with HL.¹ The study is excellent, but some points require discussion.

First, the heteroplasmy rates of the m.1555A>G and m.7444G>A variants were not determined in all patients with them.1555A>G variant.¹ To assess whether there is a causal relationship between the mtDNA variants and HL, it is important to know the individual heteroplasmy rates in different tissues. In addition, mtDNA copy numbers should be specified, as these also have a significant influence on the phenotype. Homoplasmy was reported in eight patients carrying the m.1555A>G variant.¹

Second, carrying an mtDNA variant associated with HL does not necessarily mean that that particular variant is responsible for HL. Several modifying factors can influence phenotypic expression, including heteroplasmy, mtDNA copy numbers, haplotype, and nuclear genes involved in mitochondrial regulation and reproduction. Did the degree of heteroplasmy correlate with the degree of HL?

Third, we disagree that the exact association between visual impairment, hypothyroidism, and diabetes with HL is unknown.¹ Diabetes, hypothyroidism, and visual impairment are common phenotypic features (red flags) of mitochondrial disorders (MIDs) due to mtDNA variants and may also occur in isolation or together with other phenotypic features in multisystem MIDs. The combination of diabetes, hypothyroidism, and visual impairment even suggests that MID could be the underlying cause. Visual impairment in MIDs may be due to cortical/subcortical stroke-like lesions, optic atrophy, retinitis pigmentosa, or diabetic retinopathy.

Fourth, according to **Table 3** of the index article, one patient had a history of exposure to noise (patient 379_21).¹ How did the authors rule out that HL was due to the acoustic trauma and not the MID?

Fifth, there is a discrepancy between the method and the results section in the abstract. According to the method section, 1,099 patients from 711 families were included in the study. According to the results, 711 patients were examined. This discrepancy should be resolved.

Sixth, pre- and postlingual HL is particularly associated with the variants m.1555A>G and m.7444G>A, but can generally be found in any of the mtDNA mutations.² Depending on the disease stage and modifying factors that influence the phenotypic expression of an mtDNA, variant, in principle, all syndromic and non-syndromic MIDs can be associated with HL.

Seventh, we should know why two patients with autosomal recessive inheritance **Table 3** of the index article were included in the analysis. HL in these two patients cannot be due to an mtDNA variant. This discrepancy should be resolved.

We disagree with the statement that the absence of patients with HL who are carriers of the m.3243A>G variant does not necessarily mean that HL in m.3243A>G carriers is extremely rare.¹ On the contrary, HL is a common feature of MELAS.

Overall, the interesting study has limitations that call into question the results and their interpretation. Clarifying these weaknesses would strengthen the conclusions and could add value to the study. HL is more prevalent in MID patients than expected. Factors affecting the phenotypic expression of pathogenic mtDNA variants need to be considered when analysing mtDNA-associated HL.

Authors' Response to the Letter

We acknowledge and value Dr. Josef Finsterer's comprehensive correspondence regarding our article. Here, we aim to discuss the concerns and limitations brought forth by him.

First, we acknowledge that investigating the correlation between the heteroplasmy rate of mtDNA variants and the degree of hearing loss is an intriguing aspect to examine. Nevertheless, within the confines of our study, it is difficult to accurately and precisely determine the heteroplasmy rate or mtDNA copy numbers in patients. We focused primarily on restriction fragment length polymorphism (RFLP) approach to identify pathogenic mtDNA variants. RFLP is a fast and cost-effective screening method, and the homoplasmy of mtDNA variants in eight patients was inferred from the sequence chromatogram of sequenced amplicons. However, RFLP is not sensitive enough to evaluate uneven copies of mtDNA in blood and detect low-level heteroplasmy. This limitation primarily arises from the inherent bias in the amplification bias during PCR, as described in our article.¹ Mutect2, a tool provided by GATK (Genome Analysis Toolkit, <https://gatk.broadinstitute.org/hc/en-us>), is highly effective and sensitive in identifying low-frequency variants such as low-level heteroplasmic mtDNA variants. However, it is important to note that Mutect2 requires whole genome sequencing (WGS) data as input.³ Fortunately, we had WGS data from two out of ten individuals who had the m.1555A>G variant. Among the two patients who had WGS data, YUHL681-21 was detected with 2,022 copies of mitochondrial DNA, all of which included the

identical m.1555A>G variant at the site, implicating homoplasmic m.1555A>G variant in this patient **Fig. 1.**¹ Furthermore, YUHL847-21 was found to have a total of 1,347 mitochondria, with only one of them being a different mitochondrial allele from m.1555A>G. This suggests that the m.1555A>G variant is homoplasmic, as an alternate allele frequency above 95% from WGS data is classified as homoplasmic according to gnomAD (v4.0.0) **Fig. 1.**^{1,4} Due to the limited number of available patients (YUHL 681-21 and 847-21), it is currently challenging to examine the precise association between the heteroplasmy rates of affected individuals and the degree of hearing loss. Furthermore, assessing the heteroplasmy rates in various tissues was hard to perform because obtaining biopsied tissues, such as muscle, from patients with isolated hearing loss is uncommon and completely unnecessary under standard clinical care.

Second, as Dr. Finsterer stated, there are other modifying factors that affect the penetrance and expressivity of hearing loss caused by mtDNA variants. Therefore, we assessed additional variables that could potentially influence the outcome, such as the exposure to aminoglycoside, the mtDNA haplotype, the presence of mutations in modifier genes such as *TRMU*, and any other pathogenic or likely pathogenic mutations in hearing loss-associated nuclear genes for patients whose whole exome or genome sequencing were available. Detailed results of this evaluation are already available in our article.¹

Third, Dr. Finsterer pointed out that diabetes, hypothyroidism, and visual impairment are common phenotypes of syndromic MID, alongside hearing loss. As stated in our article,¹ we acknowledge that the involvements of multiple mtDNA variants in conditions such as diabetes like maternally-inherited diabetes and deafness (MIDD), visual impairment like Leber's hereditary optic neuropathy, and hypothyroidism. However, there is a paucity of data supporting the notion that m.1555A>G or m.7444G>A is a sole cause of those phenotypes. In fact, there is no documented association between m.1555A>G or m.7444G>A variant and diabetes, visual impairment, or hypothyroidism according to OMIM (<https://omim.org/>) and Clinvar (<https://www.ncbi.nlm.nih.gov/clinvar/>) databases. In addition, these phenotypes were not co-segregated within affected family members. Thus, while it is not entirely incorrect to hypothesize that the observed syndromic phenotypes may be caused by the identified mtDNA variants combined with unknown modifying factors, adhering to the ACMG standards

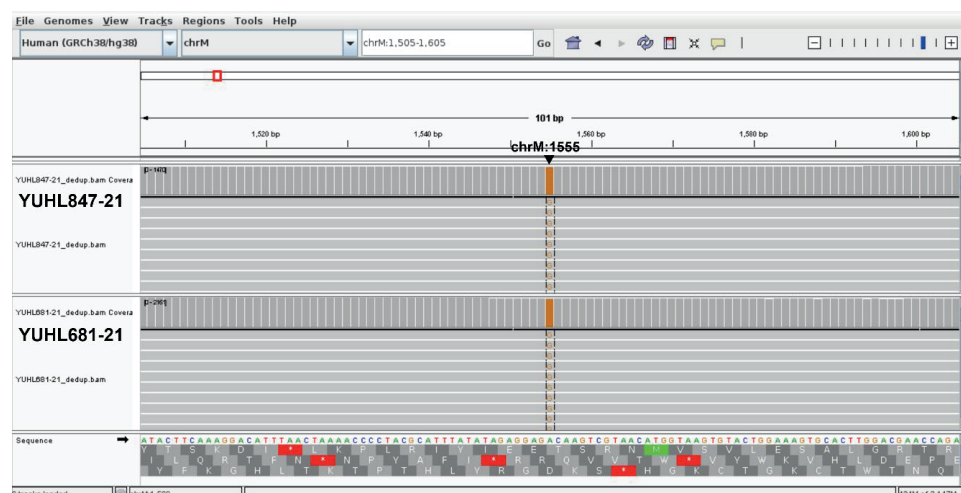


Fig. 1. Visualization of genomic locus 'chrM:1555' using the Integrative Genomics Viewer (IGV). The input files were BAM (Binary Alignment Map) files of YUHL847-21 and 681-21. Both individuals had m.1555A>G variant at a homoplasmy level.

and guidelines for the interpretation of sequence variants,⁵ we cannot ascertain that the m.1555A>G or m.7444G>A variant is responsible for the observed phenotypes, such as diabetes, visual impairment, and hypothyroidism.

Fourth, YUHL379-21 had a history of noise exposure in his twenties. However, this exposure alone was inadequate in explaining his profound hearing loss because the onset of his hearing loss occurred before the noise exposure, and the intensity and duration of the noise exposure were insufficient to cause a permanent threshold shift. Hence, it is reasonable to conclude that the m.7444G>A mtDNA variant detected in this patient is more likely a factor contributing to hearing loss.

Fifth, we apologize for the confusion that we made in the methods and results sections. In the results section, the term “patients” refers to “probands.” In order to avoid increasing the detection rate in our cohort, we selected one affected proband from each family, regardless of whether the family had multiple affected or unaffected members. There is no change in the description that we performed screening for 1,099 individuals from 711 families, including 711 probands. To enhance clarity, we would like to revise the wording in **Fig. 1** as follows: “10 unrelated hearing loss probands with the m.1555A>G variant and 2 probands with the m.7444G>A variant.”

Sixth, Dr. Finsterer inquired about the inclusion of two probands (YUHL58-21 and 82-21) who exhibited autosomal recessive inheritance in our study. As evidenced by the segregation analysis presented in our article (Supplementary Fig. 1),¹ the m.7444G>A variant of YUHL 58-21 is inherited maternally. This is supported by the fact that both the patient’s mother and brother also carried the same mtDNA mutation. The observed phenotype in the family may be explained by incomplete penetrance. Unfortunately, we were unable to obtain samples from other family members of YUHL82-21 in order to determine whether the detected mtDNA variant occurred sporadically or was inherited maternally from his mother with incomplete penetrance. Like these instances, incomplete penetrance of hearing loss resulting from mtDNA variants is not uncommon. This might lead to misinterpretation of the inheritance pattern based solely on the observed phenotype. Hence, we assessed the mtDNA variants linked to hearing loss in all 711 families, irrespective of the inheritance pattern. Moreover, there is a growing body of research indicating that de novo mtDNA mutations are responsible for MID.⁶⁻¹⁰ Therefore, comprehensive screening of mtDNA variants in a large hearing loss cohort with various modes of inheritance and varying degrees of hearing loss is a more appropriate approach to accurately determine the prevalence of these variants.

Seventh, Dr. Finsterer expressed his disagreement with the assertion that hearing loss is exceptionally uncommon in individuals with the m.3243A>G variant. We did not dispute the rarity of hearing loss in individuals with m.3243A>G variant. Among Korean patients with hearing loss, the occurrence of m.3243A>G variant is very rare, however, it does not mean that m.3243A>G does not cause hearing loss.

Finally, we express our gratitude to Dr. Finsterer for the critical reading and insightful comments about our article. Based on this response, we believe we have thoroughly clarified the issues he raised.

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REFERENCES

1. Joo SY, Jang SH, Kim JA, Kim SJ, Kim B, Kim HY, et al. Prevalence and clinical characteristics of mitochondrial DNA mutations in Korean patients with sensorineural hearing loss. *J Korean Med Sci* 2023;38(48):e355. [PUBMED](#) | [CROSSREF](#)
2. Fancello V, Fancello G, Palma S, Monzani D, Genovese E, Bianchini C, et al. The role of primary mitochondrial disorders in hearing impairment: an overview. *Medicina (Kaunas)* 2023;59(3):608. [PUBMED](#) | [CROSSREF](#)
3. Laricchia KM, Lake NJ, Watts NA, Shand M, Haessly A, Gauthier L, et al. Mitochondrial DNA variation across 56,434 individuals in gnomAD. *Genome Res* 2022;32(3):569-82. [PUBMED](#) | [CROSSREF](#)
4. Chen S, Francioli LC, Goodrich JK, Collins RL, Kanai M, Wang Q, et al. A genomic mutational constraint map using variation in 76,156 human genomes. *Nature* 2024;625(7993):92-100. [PUBMED](#) | [CROSSREF](#)
5. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015;17(5):405-24. [PUBMED](#) | [CROSSREF](#)
6. Sallevelt SC, de Die-Smulders CE, Hendrickx AT, Hellebrekers DM, de Coo IF, Alston CL, et al. De novo mtDNA point mutations are common and have a low recurrence risk. *J Med Genet* 2017;54(2):73-83. [PUBMED](#) | [CROSSREF](#)
7. Lou X, Zhou Y, Liu Z, Xie Y, Zhang L, Zhao S, et al. De novo frameshift variant in MT-ND1 causes a mitochondrial complex I deficiency associated with MELAS syndrome. *Gene* 2023;860:147229. [PUBMED](#) | [CROSSREF](#)
8. O'Donnell L, Blakely EL, Baty K, Alexander M, Bogdanova-Mihaylova P, Craig J, et al. Chronic Progressive External Ophthalmoplegia due to a Rare de novo m.12334G>A MT-TL2 Mitochondrial DNA Variant1. *J Neuromuscul Dis* 2020;7(3):355-60. [PUBMED](#) | [CROSSREF](#)
9. Uittenbogaard M, Brantner CA, Fang Z, Wong LC, Gropman A, Chiaramello A. Novel insights into the functional metabolic impact of an apparent de novo m.8993T>G variant in the MT-ATP6 gene associated with maternally inherited form of Leigh Syndrome. *Mol Genet Metab* 2018;124(1):71-81. [PUBMED](#) | [CROSSREF](#)
10. Burrage LC, Tang S, Wang J, Donti TR, Walkiewicz M, Luchak JM, et al. Mitochondrial myopathy, lactic acidosis, and sideroblastic anemia (MLASA) plus associated with a novel de novo mutation (m.8969G>A) in the mitochondrial encoded ATP6 gene. *Mol Genet Metab* 2014;113(3):207-12. [PUBMED](#) | [CROSSREF](#)