



## Anticoccidial effects of *Galla rhois* extract on *Eimeria tenella*-infected chicken

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Anticoccidial effects of *Galla rhois* (GR) extract were evaluated in chickens after oral infection with *Eimeria tenella*. This study was performed using 3-day-old chickens (n=30). The animals were divided into 3 groups as follows: GR 0.5%/infected (n=10), untreated/infected (n=10), and non-infected control (n=10). The chickens were fed a standard diet supplemented with or without GR for 1 week before infection with *E. tenella* (10,000 sporulated oocysts per chicken). The effects of GR on *E. tenella* infection were assessed by 2 parameters, number of fecal oocysts and body weight gain, and the results of the polymerase chain reaction (PCR). The GR-fed chickens produced significantly lower number of fecal oocysts ( $P<0.05$ ) than the *E. tenella*-infected chickens who were fed the standard diet. In addition, GR-based diet improved the loss of body weight caused by *E. tenella* infection. Positive findings of PCR were identified by distinct bands in the samples of *E. tenella*-inoculated chickens. However, PCR analysis revealed no *E. tenella* oocysts in the feces of GR-fed chickens. Our data showed that GR extracts had remarkable anticoccidial activities against *E. tenella*. This finding might have implications for the development of novel anticoccidial drugs.

**Key words:** Anticoccidial activity, coccidiosis, *eimeria*, *eimeria tenella*, *galla rhois*

Received 24 June 2012; Revised version received 5 September 2012; Accepted 8 September 2012

Coccidiosis is induced by infection with organisms belonging to *Eimeria* species, and it is an important parasitic disease of poultry [1]. Losses include mortality, morbidity, and cost of preventative or therapeutic drugs and/or vaccination. In addition, many of the in-feed medications commonly used for prevention of infections with *Eimeria* species have become less effective because some strains of parasites have developed reduced susceptibility to anticoccidials [2]. This finding suggests that coccidiosis is likely to have a greater impact on the profitability of broiler meat production in the future [2]. *Eimeria* infection causes extensive destruction of the intestinal epithelium, which results in reduced feed

efficiency, body weight gain, and a temporary reduction in egg production [3,4]. Although coccidiosis is mainly controlled using chemotherapeutic agents, novel approaches are urgently required because of increasing number of drug-resistant parasite strains in the commercial poultry production settings [5-7].

Our recent *in vitro* and *in vivo* studies showed that the extract of *Galla rhois* (GR) had strong antiviral and antibiotic effects [8-10]. The constituents of GR extract include methyl gallate; 3-galloyl-gallic acid; 4-galloyl-gallic acid isomers; 1,2,3,4,6-penta-*O*-galloyl- $\beta$ -D-glucose; and 2 inactive phenolic compounds, gallic acid methyl ester and gallic acid [11]. Methyl gallate, a major

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component of GR, also exhibits strong antimicrobial activity. GR extract inhibits the growth of *Escherichia coli*, without adversely affecting the growth of lactic acid-producing bacteria, and its activity is more pronounced with the presence of methyl gallate [12].

Although a different types of natural products have been investigated as alternative to control coccidiosis in chickens [3,5], the effects of GR extract on *Eimeria* infection has not been studied. In this work, we aim to evaluate the anticoccidial effects of GR extract on *Eimeria tenella* infection in Chicken

## Materials and Methods

### Experimental materials

To develop a natural herbal antimicrobial compound, over 1,000 kinds of herbal materials were tested by Center for Animal Resources Development, Wonkwang University. The herbal specimens were purchased at the University Oriental Drugstore, Iksan, Korea. Voucher specimens were deposited at the Herbarium of the College of Pharmacy, Wonkwang University. Plant material was extracted with ethanol under ultrasonic conditions for 3 h followed by paper filtration. The filtrates were evaporated in vacuum to yield water-soluble extracts. We employed *in vitro* antibacterial and *in vitro* antiviral screening tests to evaluate the effect of the medicinal herbal extracts on the pathogenic bacteria and viruses. We found that GR extract had very strong antibacterial and antiviral effects *in vitro* and *in vivo* [8-10]. In this study, we administered GR extract to the experimental animals.

### Experimental animals

The study was performed using 3-day-old chickens (n=30) in the animal facility of Center for Animal Resources Development, Wonkwang University, Korea. Animals were acclimatized and maintained in an animal facility room with regulated temperature (28±2°C), humidity (50±5%) and light/dark cycle (12/12 h). The animals were fed a commercial post-broiler diet without antibiotics and coccidiostat (Hanil Feed Co., Yongin, Korea) and tap water *ad libitum*. The chickens were maintained in wire-floored grower cages during the study period. All studies were performed in accordance with the Guide for Animal Experimentation by Wonkwang University, and the protocol was approved by the Institutional Animal Care and Use Committee of

Wonkwang University (WKU11-007). All efforts were made to minimize pain or discomfort to the animals used.

### Experimental design

Anticoccidial effects of GR extract were evaluated in chickens after infection with *E. tenella* via the oral route. This study was performed using 3-day-old chickens (n=30). The animals were divided into 3 groups: GR 0.5%/infected (n=10), untreated/infected (n=10), and non-infected control (n=10). Chickens were fed a standard diet supplemented with or without GR for 1 week before infection with *E. tenella* (10,000 sporulated oocysts per chicken). We examined the effects of GR on *E. tenella* infection using 2 parameters, number of fecal oocysts and body weight gain, and the findings of polymerase chain reaction (PCR).

### Inoculation of *Eimeria* oocysts

Oocysts of *E. tenella* were cleaned by flotation on 5.25% sodium hypochlorite and washed 3 times using phosphate buffered saline. *E. tenella* was provided kindly by Professor Wongi Min at Gyeongsang National University in Korea. Chickens were treated by oral gavage using a 24-gauge stainless steel animal feeding tube (Popper & Sons, Inc., New York, USA) attached to a 3-mL syringe. The oral infectious dose of *E. tenella* has been approximated to be 10<sup>4</sup> oocysts of *E. tenella* in 1 mL of saline. The control chickens (n=10) received saline through the same route.

### Clinical observation and weight measurements

During the study period, the animals were examined twice daily for morbidity and mortality. Further, we compared clinical signs and changes in body weight of experimental animals. Body weights were individually measured for 2 weeks before infection and for 10 days post-infection.

### Fecal sampling and oocyst counting

Fecal materials were collected from 6 to 10 days post-infection. The fecal samples were analyzed for the presence of coccidial oocysts using a standard fecal flotation technique [13]. Briefly, 1 g from each sample was suspended with 5 mL saline and pelleted by centrifugation at 1500×g for 5 min. The resulting pellet was resuspended in saturated sodium chloride (aqueous) and passed through a sieve with a mesh size of 1 mm to

remove coarse fecal debris. The resulting filtrate was used in a standard gravity vial fecal flotation using 22 mm×22 mm cover slips. After flotation, the cover slip was mounted on a slide and examined in its entirety for the presence of coccidial oocysts. Total number of oocysts was calculated using the following formula: total number of oocysts=oocyst count×dilution factor×(fecal sample volume/counting chamber volume)/number of birds per cage.

### Polymerase chain reaction

For DNA extraction, fecal samples were collected at 10 days after inoculation and cleaned with sodium hypochlorite solution (5%-6% active chlorine) for 10 min at 4°C, washed 3 times with deionized water, and resuspended in extraction buffer (10 mM Tris-Cl, pH 8.0; 50 mM EDTA, pH 8.0). The oocysts and sporocysts were completely broken down by vortexing with half the volume of 425-600 µm acid-washed glass beads (Sigma-Aldrich, St. Louis, Mo., USA). The lysate was centrifuged at 14,000×g for 10 min to eliminate debris and then digested using DNase-free RNase A (20 µg/mL) at 37°C for 1 h. Further digestion was performed using proteinase K (100 µg/mL) and sodium dodecyl sulphate (SDS, 0.5%) at 50°C for 2 h. The DNA was then extracted once with one volume of phenol, phenol/chloroform, and chloroform and precipitated using ethanol and ammonium acetate. The pellet was washed using 70% ethanol. The DNA was eluted in Tris-EDTA buffer (pH 8.0), and an aliquot was used for PCR amplification. The template DNA (50 ng) and 20 pmol of each primer were added to a PCR mixture tube (*AccuPower* PCR PreMix; Bioneer Corp., Korea) containing 2.5 U of Taq DNA polymerase, 250 µM each deoxynucleoside triphosphate, 10 mM Tris-HCl (pH 8.3), 40 mM KCl, 1.5 mM MgCl<sub>2</sub>, and the gel loading dye. The volume was adjusted using distilled water to 20 µL. The genomic DNA was amplified using species-specific forward primer (5'-AATTTAGTCCATCGCAA

CCCTTG-3') and reverse primer (5'-CGAGCGCTCTG CATACGACA-3') for the amplification of the 278-bp segment of *E. tenella* [13]. The reaction mixture was subjected to denaturation at 96°C for 5 min followed by 35 cycles of denaturation at 95°C for 30 s, annealing at 58°C or 65°C for 30 s, and extension at 72°C for 1 min, and a final extension step of 72°C for 3 min; the samples were maintained at 4°C until analysis [13]. Reactions were performed using My Genie 32 Thermal Block PCR (Bioneer, Korea). Each sample (8 µL) was mixed with 2 µL loading buffer and was analyzed by electrophoresis in 2% agarose gels stained with 0.5 µg/mL ethidium bromide.

### Statistical analysis

Differences in the mean oocyst production and mean weight gain between the 4 groups were tested using one-way analysis of variance (ANOVA; GraphPad InStat; GraphPad Software Inc., San Diego, CA) and considered significant at  $P<0.05$ .

## Results

The effects of GR on *E. tenella* infection were assessed using 2 parameters, number of fecal oocysts and body weight gain. The GR-fed chickens produced significantly lower number of fecal oocysts ( $P<0.05$ ) than *E. tenella*-infected chickens fed a standard diet. In addition, GR-based diet improved body weight loss caused by *E. tenella* infection. Our results showed that compared to untreated controls, chickens treated with GR extracts had significantly lower levels of fecal oocyst shedding and higher anticoccidial activities ( $P<0.05$ ).

Fecal oocyst shedding was significantly higher in inoculated chickens than in control chickens ( $P<0.05$ ). The number of fecal oocysts was the highest on day 7 post-inoculation (Table 1). Moreover, body weight gain was lesser in the animals in the inoculated group than in the control group (Table 2). Fecal oocyst shedding was

**Table 1.** Results of number of oocysts in the feces of chickens with or without *Galla rhois* extracts

Group	Number of oocysts (×10 <sup>6</sup> ) per gram of feces /Days post infection				
	6	7	8	9	10
Control	0±0*	0±0*	0±0*	0±0*	0±0*
Infected control <sup>a</sup>	12.9±1.5	58.6±2.3	22.4±1.9	6.9±1.6	1.8±0.6
GR+ <i>Eimeria</i> <sup>b</sup>	8.2±1.1*	22.4±2.2*	11.8±1.6*	1.2±1.2*	0±0*

<sup>a</sup>The chickens inoculated with *Eimeria tenella* oocysts.

<sup>b</sup>The chickens inoculated with *Eimeria* live oocysts and treated with 0.5% *Galla rhois* extract (GR).

\*Significantly different than that of control chickens not infected with *Eimeria* ( $P<0.05$ )

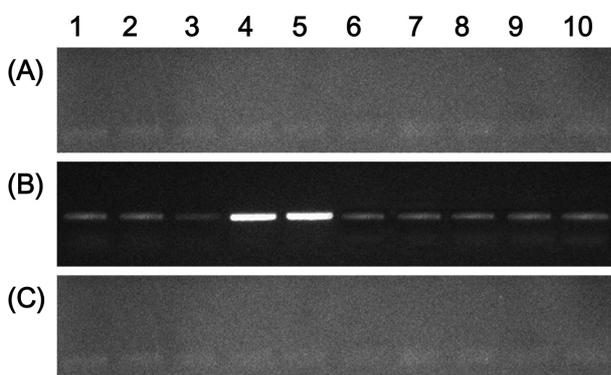
**Table 2.** Results of body weight changes of chickens with or without *Galla rhois* extracts

Group	Body weight (g)/Days post infection				
	1	3	5	7	10
Control	117.1±2.9	135.3±3.4*	169.4±2.0*	215.5±2.4*	248.7±3.2*
Infected control <sup>a</sup>	114.3±1.8	125.5±2.6	149.8±3.0	176.5±3.0	216.5±3.6
GR+ <i>Eimeria</i> <sup>b</sup>	115.6±1.8	132.8±2.7*	165.9±2.1*	212.8±2.6*	242.6±3.2*

<sup>a</sup>The chickens inoculated with *Eimeria tenella* oocysts.

<sup>b</sup>The chickens inoculated with *Eimeria* live oocysts and treated with 0.5% *Galla rhois* (GR).

\*Significantly different than that in control chickens infected with *Eimeria* ( $P < 0.05$ )



**Figure 1.** Amplification of the sample DNAs by species-specific polymerase chain reaction (PCR) for *Eimeria tenella* identified on a 2% agarose gel electrophoresis. 1~10: Animal numbers in the Group A, B, C. (A) DNAs extracted from the fecal samples of control chickens. (B) DNAs extracted from the fecal samples of *E. tenella*-infected chickens. (C) The DNAs extracted from the fecal samples of *E. tenella*-infected chickens treated with 0.5% *Galla rhois*.

observed in the inoculated animals. PCR assay, based on the amplification of internal transcribed spacer 1 (ITS1) regions of ribosomal DNA, was used to detect the chicken coccidian species *E. tenella*. The positive reactions were identified in the samples of *E. tenella* inoculated chickens. Although the strengths of PCR signals in the samples of *E. tenella* infected group were various, those of animal number 4 and 5 revealed distinct bands. Those results were suggested that the amounts of fecal shedding oocysts were various in the infected chickens. However, PCR analysis revealed no *E. tenella* oocysts in the feces of GR-fed chickens (Figure 1).

## Discussion

Coccidiosis of domestic fowl is a worldwide disease caused by obligate intracellular protozoan of the genus *Eimeria*. Coccidiosis results in economic losses in poultry production. *E. tenella* is an important pathogen causing avian coccidiosis in the poultry industry [1,14].

This disease is characterized by enteric lesions of variable extent and severity, which reduce the absorptive function of the intestinal mucosa and lead to weight loss, diarrhea, poorer feed conversion, and a higher mortality in the affected flocks [15].

Prophylactic medication has been used to control and prevent coccidiosis in commercially grown chickens. However, organisms belonging to *Eimeria* rapidly develop resistance to drugs, and an increasing concern of consumers about the drugs used to control has prompted development of alternative control strategies against avian coccidiosis. The new approaches include the use of natural products, probiotics, improved farm management practices, and modulation of the immune system of chicken [3,5].

To develop a natural herbal antimicrobial additive, we performed a screening study of over 1,000 kinds of herbal materials. Previously, we performed antibacterial and antiviral studies to evaluate the effect of the medicinal herbal extracts on pathogenic bacteria and viruses. We found that GR, a herbal antimicrobial compound, had very strong antibacterial and antiviral effects [8-10]. Assessment of host disease susceptibility to avian coccidiosis has been evaluated by counting the number of fecal oocysts and body weight changes after challenge infection with live coccidia parasites [16,17]. In this study, anticoccidial effects of the GR extract were evaluated in chickens after oral infection with *E. tenella*. Our results showed that GR-fed chickens produced significantly lower number of fecal oocysts ( $P < 0.05$ ) than the *E. tenella*-infected chicken fed the standard diet. In addition, GR-based diet improved body weight loss caused by *E. tenella* infection. Furthermore, PCR analysis revealed no *E. tenella* oocysts in the feces of GR-fed chickens.

Our data showed that GR extracts had remarkable anticoccidial activities against *E. tenella*. This finding might have implications for development of a new

anticoccidial drug. To our knowledge, this is the first study that showed the anticoccidial effect of GR on *Eimeria* parasites.

## Acknowledgments

This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (2010-0021940).

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