

Comparison of Inhibitory Effects between Enalapril and Losartan on Adrenal Catecholamine Secretion

Hyo-Jeong Lim, MD¹, Young-Youp Koh, MD², Dong-Yoon Lim, MD³

¹Division of Pulmonary and Critical Care Medicine, Department of Internal Medicine, Veterans Health Service Medical Center, Seoul; Departments of ²Internal Medicine, ³Pharmacology, Chosun University School of Medicine, Gwangju, Korea

ABSTRACT

Background: The present study was attempted to compare enalapril, an angiotensin-converting enzyme inhibitor with losartan an angiotensin II (Ang II) receptor blocker in the inhibitory effects on the secretion of catecholamines (CA) from the perfused model of the rat adrenal gland. **Methods:** The adrenal gland was isolated and perfused with Krebs-bicarbonate. CA was measured directly by using the fluorospectrophotometer. **Results:** Both enalapril and losartan during perfusion into an adrenal vein for 90 minutes inhibited the CA release evoked by acetylcholine (ACh), 1,1-dimethyl-4-phenyl piperazinium (DMPP, a selective N_n agonist), high K⁺ (a direct membrane-depolarizer), 3-(m-chloro-phenyl-carbamoyl-oxy-2-butynyl-trimethyl ammonium (McN-A-343, a selective M₁ agonist), and Ang II in a time-dependent manner. Also, in the presence of enalapril or losartan, the CA release evoked by veratridine (an activator of voltage-dependent Na⁺ channels), 6-dimethyl-3-nitro-4-(2-trifluoromethyl-phenyl)-pyridine-5-carboxylate (BAY-K-8644, an L-type Ca²⁺ channel activator), and cyclopiazonic acid (a cytoplasmic Ca²⁺-ATPase inhibitor) were significantly reduced. Based on the same concentration of enalapril and losartan, for the CA release evoked by ACh, high K⁺, DMPP, McN-A-343, Ang II, veratridine, BAY-K-8644, and cyclopiazonic acid, the following rank order of inhibitory potency was obtained: losartan > enalapril. In the simultaneous presence of enalapril and losartan, ACh-evoked CA secretion was more strongly inhibited compared with that of enalapril- or losartan-treated alone. **Conclusions:** Collectively, these results demonstrate that both enalapril and losartan inhibit the CA secretion evoked by activation of both cholinergic and Ang II type-1 receptors stimulation in the perfused rat adrenal medulla. When these two drugs were used in combination, their effects were enhanced, which may also be of clinical benefit. Based on concentration used in this study, the inhibitory effect of losartan on the CA secretion seems to be more potent than that of enalapril.

(J Korean Soc Hypertens 2014;20(2):51-67)

Key Words: Enalapril; Losartan; Catecholamine secretion; Adrenal medulla; angiotensin II type-1 receptor blocker; Angiotensin-converting enzyme inhibitor

Introduction

Generally, the adrenal medulla releases catecholamines

(CA) chiefly via activation of its nicotinic acetylcholine (ACh) receptors by ACh, which is liberated from the terminal of the splanchnic nerve. Binding of ACh to these nicotinic receptors leads to a depolarization of the cell membrane by an influx of Na⁺ through receptor-gated cation channels, causing an influx of Ca²⁺ through voltage-operative Ca²⁺ channels, which results in CA secre-

Received: 2014.3.14, Revised: 2014.5.13, Accepted: 2014.6.13

Correspondence to: Dong-Yoon Lim, MD

Address: Department of Pharmacology, Chosun University School of Medicine, 309 Pilmun-daero, Dong-gu, Gwangju 501-759, Korea

Tel: +82-62-230-6335, Fax: +82-62-227-4693

E-mail: dylim@chosun.ac.kr

tion by exocytotic mechanism.¹⁻³⁾ Therefore, adrenal chromaffin cells are widely used as a model to study CA secretion in response to stimulation by the sympathetic nervous system.

The renin-angiotensin-aldosterone system (RAAS) is an important mediator in the pathophysiology of hypertension, with excessive activity in the RAAS playing a crucial role in target end-organ damage, such as myocardial infarction, congestive heart failure, coronary artery disease and end-stage renal disease.⁴⁾ The valuable influence of angiotensin-converting enzyme inhibitors (ACEIs) and angiotensin II (Ang II) receptors antagonists (ARBs) may be shared by RAAS, as well as sympathetic overactivity inhibition. Ang II, the main effector of the RAAS, was found to activate sympathetic nerve traffic and its sequelae in numerous tissues.⁵⁾ This facilitatory effect is exerted via Ang II type-1 (AT₁) receptors and opposed by AT₁ receptor antagonism and the reduction of Ang II production. Furthermore, inhibition of ACE affects the bradykinin concentration by peptide degradation to inactive substrates. Therefore, ACEIs are expected to elevate the bradykinin concentration, which has been suggested to affect sympathetic tone. It was found that bradykinin enhanced the epinephrine (EP) release from the adrenal medulla and norepinephrine (NE) from the animal and human heart, indicating the stimulatory action of the peptide on sympathetic excitation.⁶⁻⁸⁾ A counteraction of bradykinin was also described.⁹⁾ Consequently, in respect to some different mechanisms of action of ACEIs and ARBs, it can be presumed that both drug classes may mediate different effects on sympathetic activity. Indeed, data regarding sympathetic inhibition during ACEIs therapy in comparison to ARB are sparse and inconsistent. Several studies have shown a similar contribution of both drug classes to sympathetic inhibition, while others suggested differences existing in this regard

between drugs.¹⁰⁻¹³⁾ Different mechanisms of RAAS inhibition by ACEIs and ARBs mentioned above might at least in part explain existing divergences.

Currently, two classes of drugs that attenuate the action of Ang II and act as antihypertensive agents by different mechanisms are available, which include ACEIs and ARBs. ACEIs act by reducing the biosynthesis of Ang II by blocking the action of ACE on Ang I but do not inhibit the alternative non-ACE Ang II generating pathways whereas ARBs block the AT₁ receptor-mediated actions of Ang II without inhibiting the synthesis of Ang II. Taking into consideration the alternative pathways of Ang II generation, ARBs reduce the activation of AT₁ receptor more effectively than do ACEIs.¹⁴⁾ The current place of ARBs in the treatment of essential hypertension is as an alternative to ACEIs in patients who do not tolerate them due to adverse-effects. But there is constant controversy over the comparative efficacy of ACEIs and ARBs due to the additional advantages of ARBs over ACEIs.¹⁵⁾ Hence, the present study was designed to compare losartan with enalapril in inhibitory effects on the CA secretion from the perfused model of the isolated rat adrenal medulla.

Subjects and methods

1. Experimental procedure

Male Sprague-Dawley rats, weighing 200 to 300 g, were anesthetized with thiopental sodium (50 mg/kg) intraperitoneally. The adrenal gland was isolated by the methods described previously.¹⁶⁾ The abdomen was opened by a midline incision, and the left adrenal gland and surrounding area were exposed by placing three hook retractors. The stomach, intestine and portion of the liver were not removed, but pushed over to the right side and covered by saline-soaked gauze pads and urine in bladder

was removed in order to obtain enough working space for tying blood vessels and cannulations.

A cannula, used for perfusion of the adrenal gland, was inserted into the distal end of the renal vein after all branches of adrenal vein (if any), vena cava and aorta were ligated. Heparin (400 IU/mL) was injected into vena cava to prevent blood coagulation before ligating vessels and cannulations. A small slit was made into the adrenal cortex just opposite entrance of adrenal vein. Perfusion of the gland was started, making sure that no leakage was present, and the perfusion fluid escaped only from the slit made in adrenal cortex. Then the adrenal gland, along with ligated blood vessels and the cannula were carefully removed from the animal and placed on a platform of a leucite chamber. The chamber was continuously circulated with water heated at $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$.

2. Perfusion of adrenal gland

The adrenal glands were perfused by means of ISCO pump (WIZ Co., Lincoln, NE, USA) at a rate of 0.33 mL/min. The perfusion was carried out with Krebs-bicarbonate solution of following composition (mM): NaCl, 118.4; KCl, 4.7; CaCl_2 , 2.5; MgCl_2 , 1.18; NaHCO_3 , 25; KH_2PO_4 , 1.2; glucose, 11.7. The solution was constantly bubbled with 95% O_2 + 5% CO_2 and the final pH of the solution was maintained at 7.4 to 7.5. The solution contained disodium EDTA (10 $\mu\text{g/mL}$) and ascorbic acid (100 $\mu\text{g/mL}$) to prevent oxidation of CA.

3. Drug administration

The perfusions of 1.1-dimethyl-4-phenyl piperazinium (DMPP, 10^{-4} M) and McN-A-343 (10^{-4} M) for 2 minutes and/or a single injection of ACh (5.32×10^{-3} M) and KCl (5.6×10^{-2} M) in a volume of 0.05 mL were made into perfusion stream via a three-way stopcock, respectively. Veratridine (10^{-4} M), BAY-K-8644 (10^{-5} M), and cyclo-

piazonic acid (10^{-5} M) were also perfused for 4 minutes, respectively. In the preliminary experiments, it was found that upon administration of the above drugs, secretory responses to ACh, KCl, 3-(m-chloro-phenyl-carbamoyl-oxy-2-butyryl-trimethyl ammonium (McN-A-343), veratridine, 6-dimethyl-3-nitro-4-(2-trifluoromethyl-phenyl)-pyridine-5-carboxylate (BAY-K-8644), and cyclopiazonic acid returned to preinjection level in about 4 minutes, but the responses to DMPP returned in 8 minutes.

4. Collection of perfusate

As a rule, prior to stimulation with various secretagogues, the perfusate was collected for 4 minutes to determine the spontaneous secretion of CA (background sample). Immediately after the collection of the background sample, collection of the perfusates was continued in another tube as soon as the perfusion medium containing the stimulatory agent reached the adrenal gland. Stimulated sample's perfusate was collected for 4 to 8 minutes. The amounts secreted in the background sample have been subtracted from that secreted from the stimulated sample to obtain the net secretion value of CA, which is shown in all of the figures.

To study the effect of enalapril and losartan on the spontaneous and evoked secretion, the adrenal gland was perfused with Krebs solution containing quinine for 20 minutes, and then the perfusate was collected for a certain period (background sample). Then the medium was changed to the one containing the stimulating agent or along with enalapril or losartan, and the perfusates were collected for the same period as that for the background sample. The adrenal gland's perfusate was collected in chilled tubes.

5. Measurement of catecholamines

CA content of perfusate was measured directly using

the fluorometric method of Anton and Sayre¹⁷⁾ without the intermediate purification alumina for the reasons described earlier¹⁶⁾ using fluorospectrophotometer (Kontron Co., Milano, Italy). A volume of 0.2 mL of the perfusate was used for the reaction. The CA content in the perfusate of stimulated glands by secretagogues used in the present work was high enough to obtain readings several folds greater than the reading of control samples (unstimulated). The sample blanks were also lowest for perfusates of stimulated and non-stimulated samples. The content of CA in the perfusate was expressed in terms of NE (base) equivalents.

6. Statistical analysis

The statistical difference between the control and pre-treated groups was determined by the Student t and analysis of variance tests. A p-value of less than 0.05 was considered to represent statistically significant changes unless specifically noted in the text. Values given in the text refer to the means and the standard errors of the mean. The statistical analysis of the experimental results was made using the computer program described by Tallarida and Murray.¹⁸⁾

7. Drugs and their sources

The following drugs were used: losartan potassium (Merk & Co., Inc. Rahway, Nj, USA), enalapril maleate (MSD Korea, Seoul, Korea), DMPP iodide, ACh chloride, NE bitartrate, potassium chloride (KCl), BAY-K-8644, cyclopiazonic acid, veratridine hydrochloride (Sigma Chemical Co., St Louis, MO, USA), and McN-A-343 chloride (RBI, St Louis, MO, USA). Drugs were dissolved in distilled water (stock) and added to the normal Krebs solution as required except BAY-K-8644, which was dissolved in 99.5% ethanol and diluted appropriately with Krebs-bicarbonate solution (final concentration of

alcohol was less than 0.1%). Concentrations of all drugs used are expressed in terms of molar base.

Results

1. Effects of enalapril and losartan on catecholamine secretion evoked by ACh, excess K⁺, DMPP, McN-A-343, and Ang II from the perfused rat adrenal medulla

After the perfusion with oxygenated Krebs-bicarbonate solution for 1 hour, basal CA release from the isolated perfused rat adrenal glands amounted to 21.2 ± 2.1 ng for 2 minutes (n = 6). It was attempted initially to examine the effects of enalapril or losartan itself on CA secretion from the perfused model of the rat adrenal glands. However, in the present study, enalapril (10-100 μ M) or losartan (20-180 μ g/mL) itself did not produce any effect on basal CA output from perfused rat adrenal glands (data not shown). Therefore, it was decided to investigate the effects of enalapril or losartan on CA secretion evoked by several Secretagogues. Secretagogues were given at intervals of 15 or 20 minutes. Enalapril or losartan was perfused for 90 minutes after completion of control responses to various secretagogues. When ACh (5.32×10^{-2} M) in a volume of 0.05 mL was given into the adrenal vein, the amount of CA secreted was $1,261 \pm 32$ ng for 4 minutes. However, the pretreatment with enalapril (50 μ M) or losartan (50 μ M) for 90 minutes inhibited ACh-evoked CA secretion in time-dependent manner. As shown in Fig. 1, in the presence of enalapril or losartan, ACh-evoked CA releasing responses were inhibited maximally by 85% and 68% of the corresponding control release, respectively. Also, it has been found that a depolarizing agent, high K⁺ stimulates CA secretion (608 ± 24 ng for 0 to 4 minutes). However, in the presence of enalapril (50 μ M) or losartan (50 μ M), excess K⁺ (56 mM)-evoked CA secretion was significantly inhibited

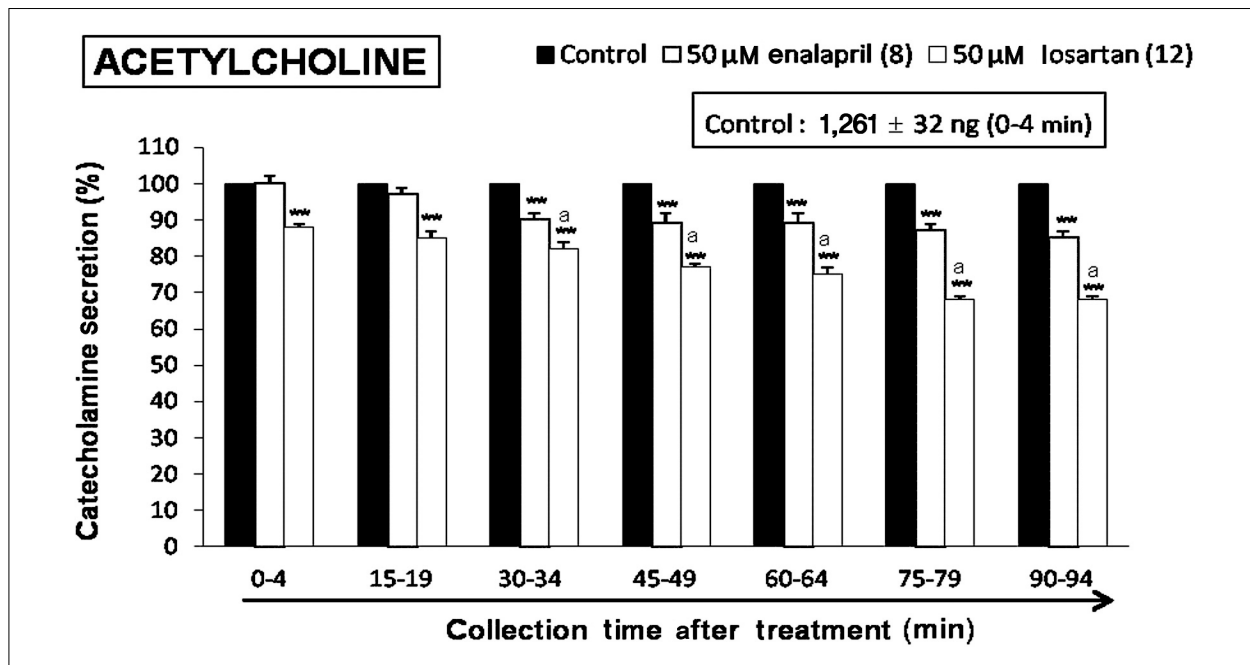


Fig. 1. Time-course effects of losartan or enalapril on the secretory responses of CA evoked by acetylcholine from the isolated perfused rat adrenal glands. The CA secretion by a single injection of (ACh, 5.32×10^{-3} M) in a volume of 0.05 mL was evoked at 15 minutes intervals during loading with losartan (50 μ M) or enalapril (50 μ M) for 90 minutes as indicated by the arrow marks. Columns and vertical lines represent mean \pm standard error. Numbers in the parenthesis indicate number of rat adrenal glands. Ordinate: the amounts of CA secreted from the adrenal gland (% of control). Abscissa: collection time of perfusate (minutes). Statistical difference was obtained by comparing the corresponding control with losartan- or enalapril-treated group (**), and by comparing the losartan-treated group with enalapril-treated group (a). ACh-induced perfusates were collected for 4 minutes. **p < 0.01, ^ap < 0.05. CA, catecholamines; ACh, acetylcholine.

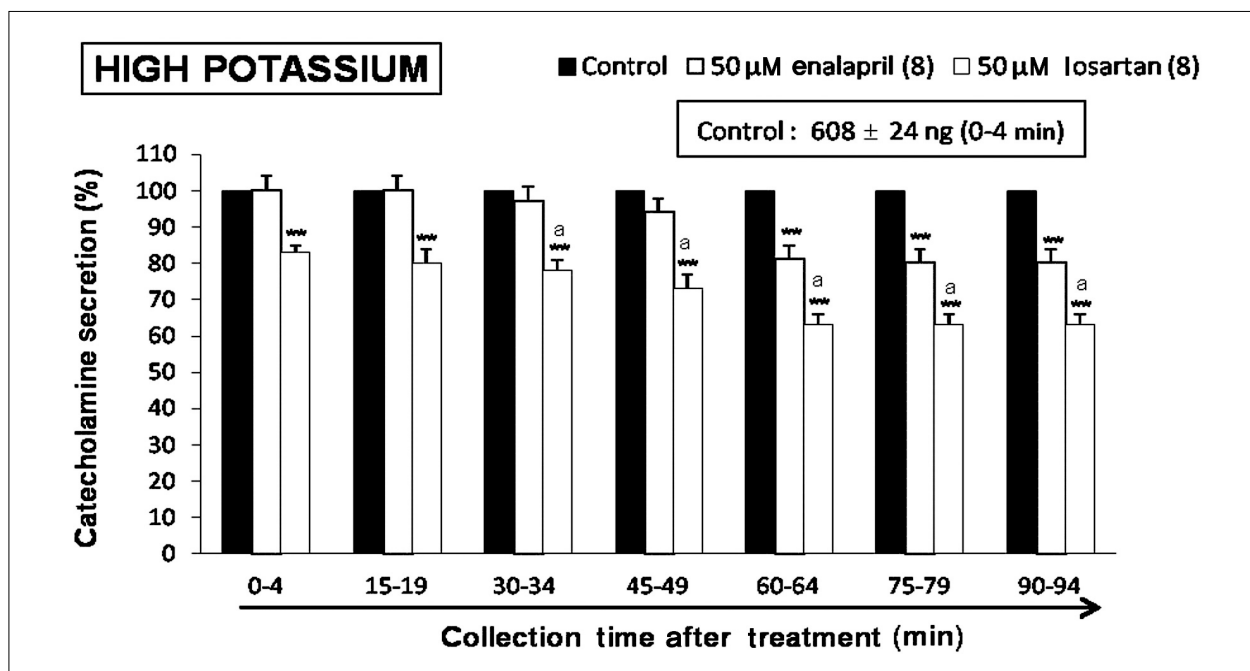


Fig. 2. Time-course effects of losartan or enalapril on high potassium-evoked catecholamines (CA) secretion from the isolated perfused rat adrenal glands. The CA secretion by a single injection of high potassium (56 mM) in a volume of 0.05 mL was evoked at 15 minutes intervals during loading with losartan (50 μ M) or enalapril (50 μ M) for 90 minutes as indicated by the arrow mark. Other legends are the same as in Fig. 1. High potassium-induced perfusates were collected for 4 minutes. **p < 0.01, ^ap < 0.05.

by 80% and 63% of the control release, respectively (Fig. 2). When perfused through the rat adrenal gland, DMPP (10^{-4} M), which is a selective neuronal nicotinic receptor agonist, evoked a sharp and rapid increase in CA secretion ($1,272 \pm 38$ ng for 0 to 8 minutes). However, as shown in Fig. 3, DMPP-evoked CA secretion during treatment with enalapril or losartan was reduced to 88% and 71% of the control release, respectively. McN-A-343 (10^{-4} M), which is a selective muscarinic M_1 -agonist,¹⁹ perfused into an adrenal gland for 4 minutes caused an increased CA secretion (520 ± 27 ng for 0 to 4 minutes). However, McN-A-343-evoked CA secretion in the presence of enalapril or losartan was markedly depressed to 80% and 67% of the corresponding control secretion, respectively (Fig. 4).

Since Hano et al.²⁰ have suggested that Ang II increase EP release from the adrenal medulla via the AT_1 receptors, we examined the effect of enalapril or losartan

on Ang II-evoked CA secretion. Ang II (100 nM) significantly increased the CA secretory response (571 ± 38 ng for 0 to 4 minutes), while in the presence of enalapril (15 μ M) or losartan (15 μ M), Ang II-evoked CA secretion was greatly inhibited to 81% and 46% of the corresponding control secretion, respectively (Fig. 5). Interestingly, as shown in Figs. 1-5, there was significantly statistical difference in inhibitory effect between enalapril and losartan on the CA secretion evoked ACh, high K^+ , DMPP, McN-A-343, and Ang II, indicating that losartan is more potent inhibitory effect on the CA secretion than enalapril.

2. Effects of enalapril and losartan on catecholamine secretion evoked by BAY-K-8644, cyclopiazonic acid, and veratridine from the perfused rat adrenal medulla

It was of interest to determine the effects of enalapril

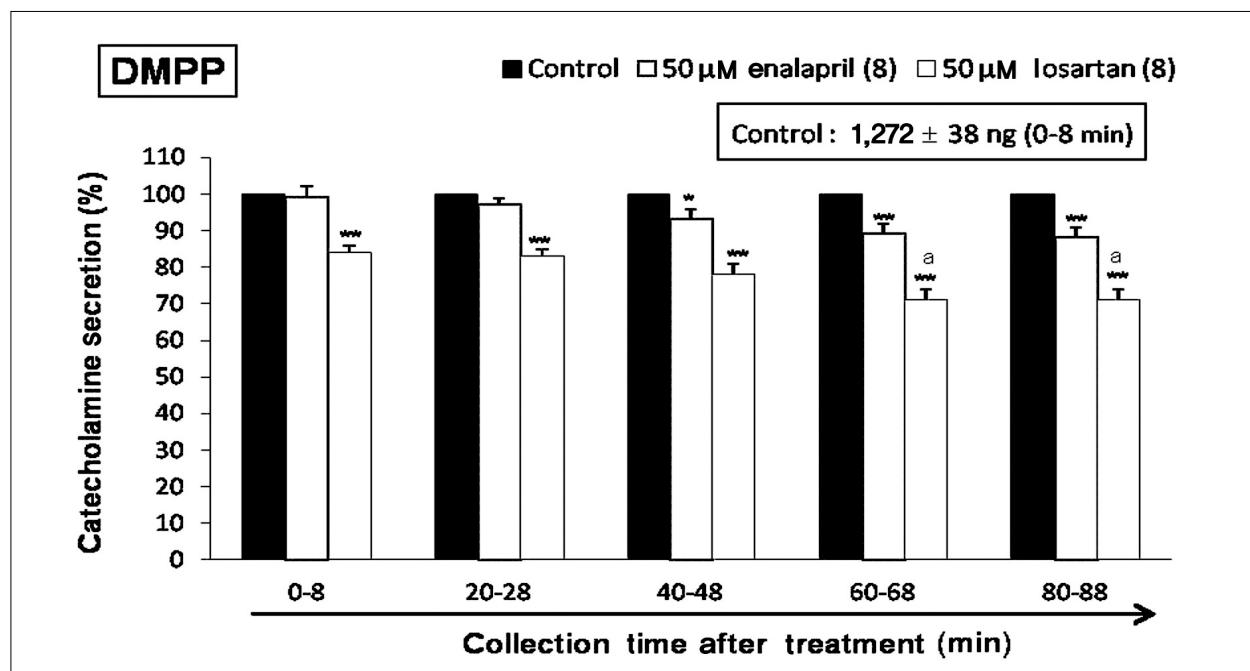


Fig. 3. Time-course effects of losartan or enalapril on 1.1-dimethyl-4-phenyl piperazinium iodide (DMPP)-evoked catecholamines (CA) secretion from the isolated perfused rat adrenal glands. The CA secretion by perfusion of DMPP (10^{-4} M) for 2 minutes was induced at 20 minutes intervals during loading with losartan (50 μ M) or enalapril (50 μ M) for 90 minutes as indicated by the arrow mark. DMPP-induced perfusates were collected for 8 minutes. Other legends are the same as in Fig. 1. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

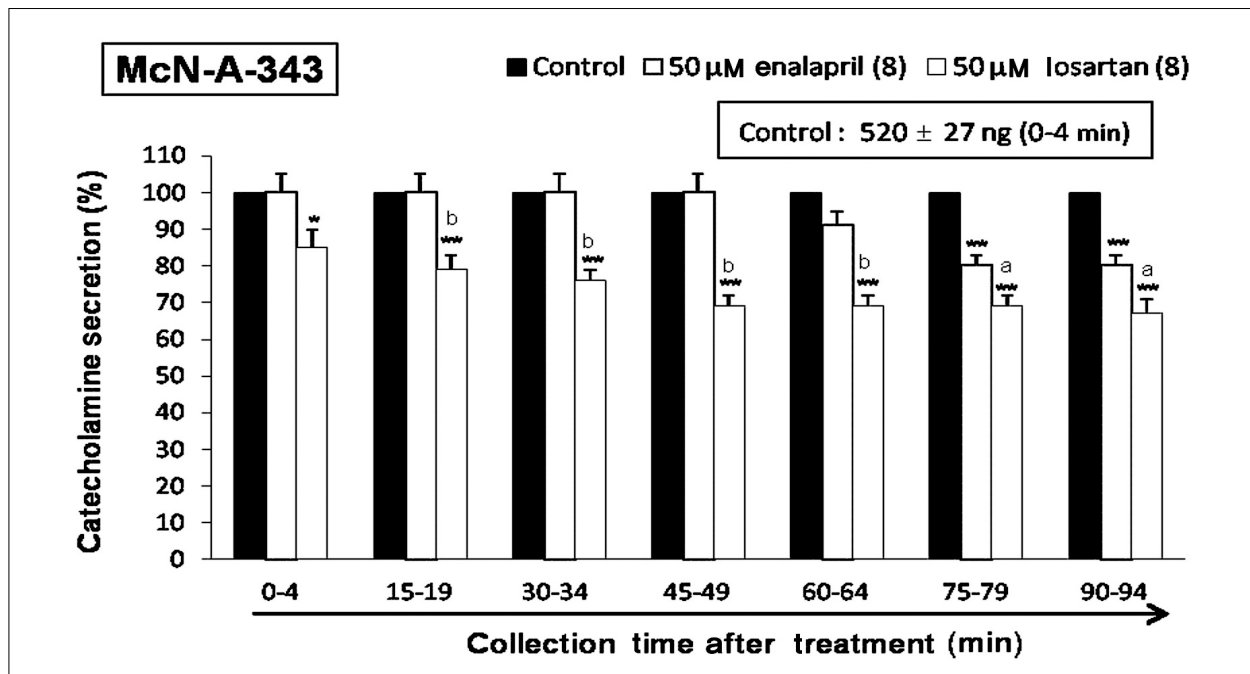


Fig. 4. Time-course effects of losartan or enalapril on McN-A-343-evoked catecholamines (CA) secretion from the isolated perfused rat adrenal glands. The CA secretion by perfusion of 3-(m-chloro-phenyl)-carbamoyl-oxy-2-butynyl-trimethyl ammonium chloride (McN-A-343, 10^{-4} M) for 4 minutes was induced at 15 minutes intervals during loading with losartan (50 μM) or enalapril (50 μM) for 90 minutes as indicated by the arrow mark. McN-A-343-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. * $p < 0.05$, ** $p < 0.01$. ^a $p < 0.05$, ^b $p < 0.01$.

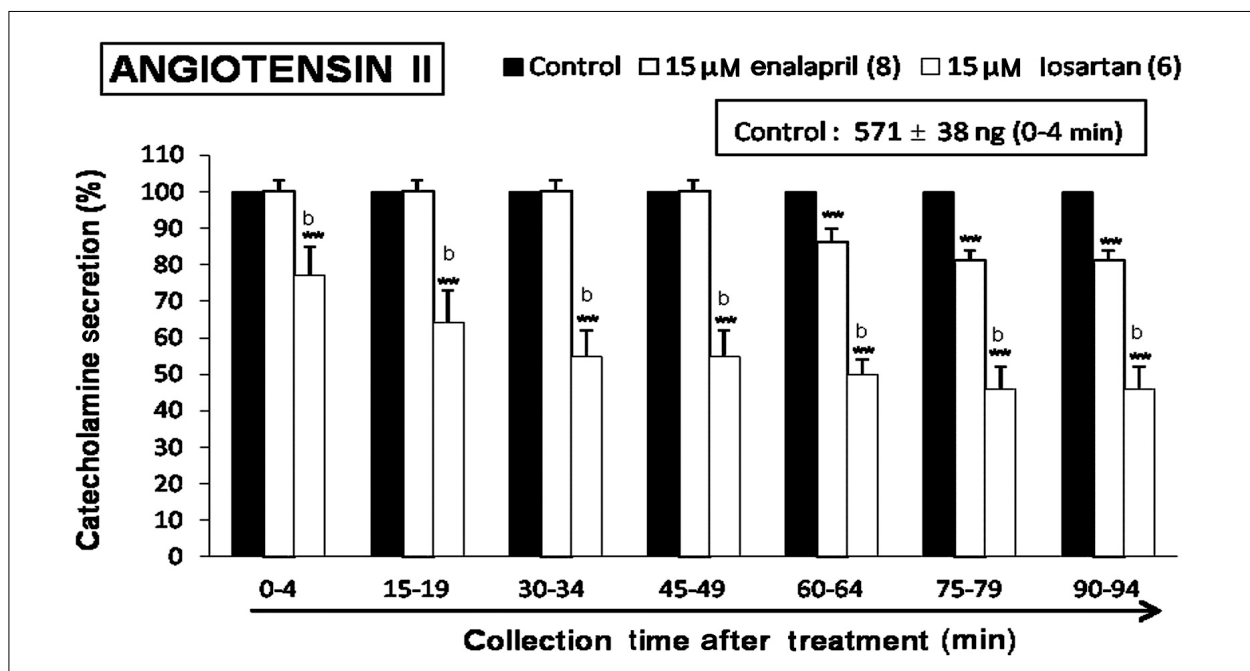


Fig. 5. Time-course effects of losartan or enalapril on angiotensin II-evoked catecholamines (CA) secretion from the perfused rat adrenal glands. The CA secretion by perfusion of angiotensin II (10^{-7} M) for 4 minutes was induced at 15 minutes intervals during loading with losartan (15 μM) or enalapril (15 μM) for 90 minutes as indicated by the arrow mark. Angiotensin II-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. ** $p < 0.01$. ^b $p < 0.01$.

or losartan on BAY-K-8644-stimulated CA secretion from the isolated perfused rat adrenal glands, since BAY-K-8644 is known to be a calcium channel activator, which enhances basal Ca^{2+} uptake²¹⁾ and CA release.²²⁾ BAY-K-8644 (10^{-5} M)-stimulated CA secretion in the presence of enalapril (15 μM) or losartan (15 μM) was inhibited to maximally 82% and 73% of the control as compared to the corresponding control release (619 ± 28 ng for 0 to 4 minutes), respectively (Fig. 6).

Cyclopiazonic acid, a mycotoxin from *Aspergillus* and *Penicillium*, has been described as a highly selective inhibitor of Ca^{2+} -ATPase in skeletal muscle sarcoplasmic reticulum.^{23,24)} The inhibitory action of enalapril or losartan on cyclopiazonic acid-evoked CA secretory response was observed as shown in Fig. 7. In the presence of enalapril (15 μM) or losartan (15 μM) in 8 rat adrenal glands, cyclopiazonic acid (10^{-5} M)-evoked CA secretion was significantly depressed by 85% and 72% of the control secretory response (536 ± 22 ng for 0 to 4 minutes), respectively.

It has also been known that veratridine-induced Na^{+} influx mediated through Na^{+} channels increased Ca^{2+} influx via activation of voltage-dependent Ca^{2+} channels and produced the exocytotic secretion of CA in cultured bovine adrenal medullary cells.²⁵⁾ To characterize the pharmacological action of enalapril and losartan on voltage-dependent Na^{+} channels, the effect of enalapril and losartan on the CA secretion induced by veratridine was examined here. As shown in Fig. 8, veratridine greatly produced CA secretion ($1,244 \pm 24$ ng for 0 to 4 minutes). However, in the presence of enalapril (15 μM) and losartan (15 μM), veratridine (100 μM)-evoked CA secretion was inhibited to 90% and 68% of the corresponding control release in a time-dependent fashion.

Moreover, there was also statistical difference in inhibitory effect between enalapril and losartan on the CA secretion evoked BAY-K-8644, cyclopiazonic acid, and

veratridine, as shown in Figs. 6-8. These results demonstrate that losartan shows more potent inhibitory effect on the CA secretion than enalapril.

3. Effects of enalapril plus losartan on

ACh-evoked catecholamine secretion from the perfused rat adrenal medulla

In the present work, enalapril or losartan caused inhibition in the CA secretion by cholinergic receptor stimulation as well as by Ang II from the perfused rat adrenal gland. Therefore, in order to establish the combined effects of enalapril and losartan on ACh-evoked CA release, we tried to examine enalapril plus losartan-induced inhibitory effects on CA secretion evoked by ACh.

In the simultaneous presence of enalapril (50 μM) and losartan (50 μM) for 90 minutes, Ang II (100 nM)-evoked CA release more slightly inhibited to about 57% of the corresponding control release ($1,261 \pm 32$ ng for 0 to 4 minutes), compared to the inhibitory effect induced by enalapril or losartan alone as shown in Fig. 9. Also, there was statistical difference in inhibitory effect on ACh-evoked CA secretion between enalapril versus losartan as well as enalapril versus losartan + enalapril, as shown in Fig. 9.

Discussion

The rennin-angiotensin system plays a crucial role in regulation of blood pressure (BP). The primary effector peptide Ang II behaves at two receptors, AT_1 and AT_2 . The AT_1 receptor mediates functions like vasoconstriction and aldosterone secretion while the AT_2 receptor mediates vasodilatation and natriuresis. ACEIs inhibit biosynthesis of Ang II whereas ARBs completely block AT_1 receptors, and both are effective antihypertensive agents. But, currently, there is a constant controversy over the comparative efficacy of ACEIs and ARBs due to the possibility of Ang II generation by alternative pathways with

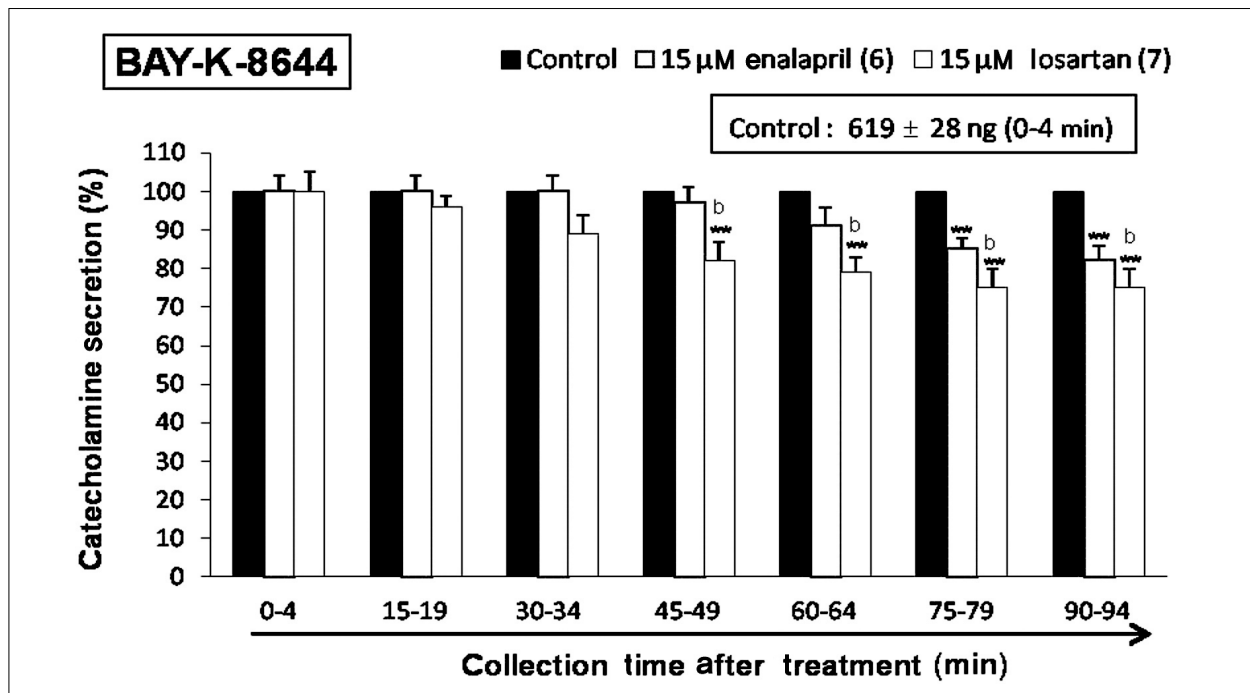


Fig. 6. Time-course effects of losartan or enalapril on 6-dimethyl-3-nitro-4-(2-trifluoromethyl-phenyl)-pyridine-5-carboxylate (BAY-K-8644)-evoked catecholamines (CA) secretion from the perfused rat adrenal glands. The CA secretion by perfusion of BAY-K-8644 (10^{-5} M) for 4 minutes was induced at 15 minutes intervals during loading with losartan (15 μ M) or enalapril (15 μ M) for 90 minutes as indicated by the arrow mark. Bay-K-8644-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. ^{***} p < 0.01, ^b p < 0.01.

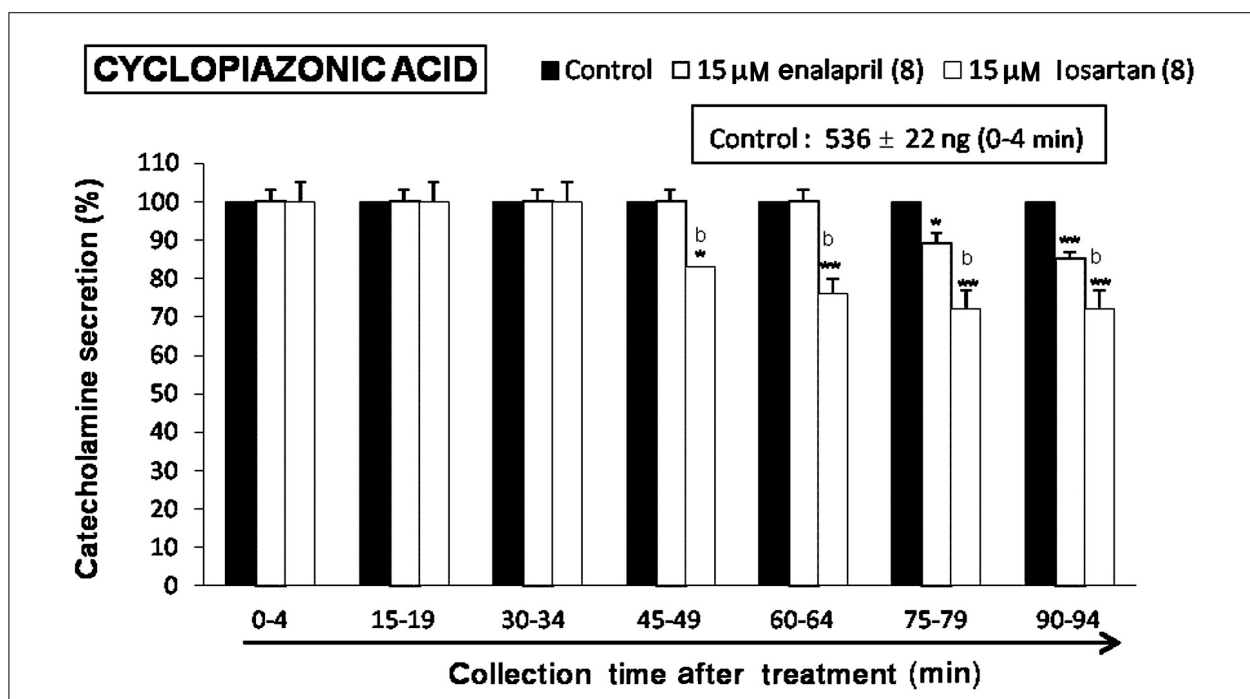


Fig. 7. Time-course effects of losartan or enalapril on cyclopiazonic acid-evoked catecholamines (CA) secretion from the perfused rat adrenal glands. The CA secretion by perfusion of cyclopiazonic acid (10^{-5} M) for 4 minutes was induced at 15 minutes intervals during loading with losartan (15 μ M) or enalapril (15 μ M) for 90 minutes as indicated by the arrow mark. Cyclopiazonic acid-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. ^{*} p < 0.05, ^{**} p < 0.01, ^b p < 0.01.

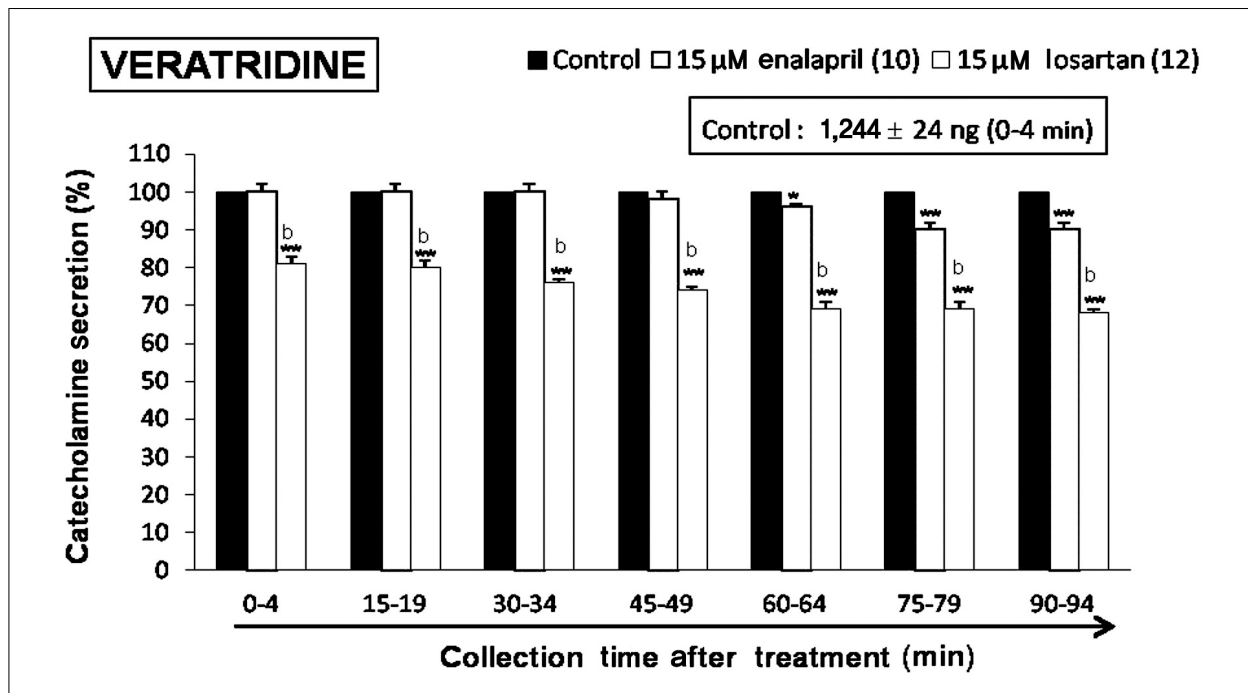


Fig. 8. Time-course effects of losartan or enalapril on veratridine-evoked catecholamines (CA) secretion from the perfused rat adrenal glands. The CA secretion by perfusion of veratridine (10^{-5} M) for 4 minutes was induced at 15 minutes intervals during loading with losartan (15 μ M) or enalapril (15 μ M) for 90 minutes as indicated by the arrow mark. Veratridine-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. * $p < 0.05$, ** $p < 0.01$, ^b $p < 0.01$.

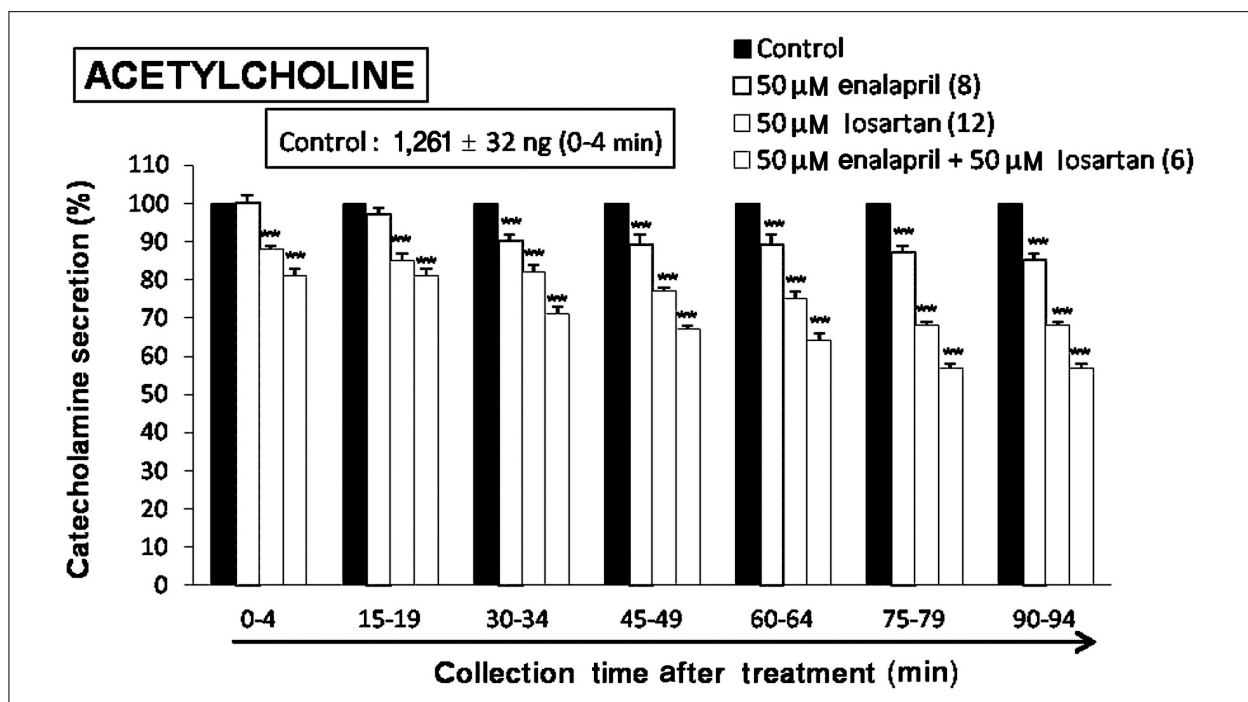


Fig. 9. Comparative time-course effects of losartan, enalapril and losartan + enalapril on acetylcholine-evoked catecholamines (CA) secretion from the perfused rat adrenal glands. The CA secretion by a single injection of acetylcholine chloride (5.32×10^{-3} M) in a volume of 0.05 mL was evoked at 15 minutes intervals during loading with losartan (50 μ M) or enalapril (50 μ M), losartan (50 μ M) + enalapril (50 μ M) for 90 minutes as indicated by the arrow mark. Acetylcholine-induced perfusates were collected for 4 minutes. Other legends are the same as in Fig. 1. ** $p < 0.01$ enalapril-treated group vs. losartan-treated group or enalapril + losartan-treated group.

the use of ACEIs. Hence, ARBs are said to inhibit the activation of AT₁ receptors more effectively than ACEIs. Therefore, the present study was designed to compare the inhibitory effects of ACEIs with ARBs on adrenal CA secretion. As a prototype and widely used ACEI, enalapril was taken for comparison with losartan, which is also prototype ARB and a competitive antagonism at Ang II receptors.

The present study has demonstrated that both enalapril and losartan inhibit the CA secretory responses evoked by activation of both cholinergic and Ang II receptors as well as by direct membrane depolarization in the perfused model of the isolated rat adrenal medulla. Based on the same concentration used in this study, it seems that the inhibitory effect of losartan on the CA secretion is more potent than that of enalapril. However, when enalapril and losartan were given in combination, their inhibitory effects on ACh-evoked CA secretion were potentiated, which may also be clinically advantageous in treatment of cardiovascular diseases. In support of this idea, our study showed that both losartan and enalapril retrogradely perfused into the adrenal medulla significantly diminished the CA release evoked by cholinergic stimulation, such as administration of ACh, DMPP (a selective neuronal nicotinic receptor agonist), and McN-A-343 (a selective muscarinic M₁ receptor agonist). However, enalapril showed lower potency in the inhibitory effect on the CA release compared to losartan.

Previously published experimental work where the adrenal gland has been pharmaceutically stimulated has involved the direct administration of Ang II into the gland via the adrenal artery.²⁶⁻²⁹⁾ In the present study, Ang II was also administered via the adrenal vein, which resulted in greatly increases in CA release. In comparison, direct administration of cholinergic secretagogues such as ACh, DMPP, and McN-A-343 into the gland re-

sulted in much greater increases. However, we did not measure the systemic levels of circulating CA, because the isolated perfused model of the rat adrenal medulla was used in this study. Data from Yamaguchi et al.²⁹⁾ have suggested that adrenal blood levels of basal NE release are 10 times and EP release are 100 times greater than their systemic circulation counterparts. Thus, any systemic contribution during direct stimulation of the adrenal medulla would be of little consequence, as adrenal CA release is magnitudes greater than the circulating level.

The main reason for using Ang II in the present study was to demonstrate that the perfusion of losartan was pharmacologically active. However, as enalapril acts via a different pathway, blocking the conversion of Ang, the use of Ang II to determine the activity of the preparation of enalapril was inappropriate to compare with that of losartan. Previously, Cavadas et al.³⁰⁾ have demonstrated that AT₁ stimulation induces CA secretion from human adrenal chromaffin cells probably by raising cytosolic calcium in primary cultures of human adrenal chromaffin cells.

In this study, losartan and enalapril did not increase basal CA release in this study (data not shown). Also, in other studies where direct drug administration into the adrenal gland has been used, no increase in basal CA release has been reported.²⁹⁾ However, it has been found that the systemic administration of candesartan and ramipril increased basal CA release.³¹⁾ But the systemic administration of both candesartan and ramipril resulted in quite significant decreases in blood pressure.³¹⁾ This fall in blood pressure would have been expected to cause some degree of sympathetic response with accompanying release of systemic CAs, predominantly NE from the sympathetic nerve endings. Thus, circulating CAs could have reached levels high enough to influence the adrenal

vein collection.

Losartan may act locally by inhibiting this regulation by blocking the Ang II receptor on the chromaffin cell, while enalapril prevents local conversion of Ang to its active form. Thus, both drugs may act by inhibiting elevation of CA release by inhibiting local RAAS modulation. In the present study, it has been shown that these two drugs also have an additive effect as the reduction in the ACh-evoked CA release following their administration decreased from 15-32% to 57%. This additive effect may be of benefit when treating cardiovascular diseases, such as hypertension or heart failure. Both losartan and enalapril had inhibitory effects on cholinergic stimulation-evoked CA release evoked by ACh, DMPP and McN-A-343. However, Ang converting enzyme inhibitors, such as enalapril, because they share a common pathway, are known to cause bradykinin accumulation, which has the potential to cause cough.³²⁾ Thus, losartan may be advantageous over enalapril because it does not cause cough.

In light of these results, both losartan and enalapril inhibited cholinergic receptor-stimulated CA release and the most likely explanation seems to be blockade of local modulation by RAAS. This effect may prove beneficial when treating hypertension and heart failure. Enalapril is weaker than losartan in inhibiting CA release evoked by stimulation of cholinergic (both nicotinic and muscarinic) receptors as well as Ang II receptors. In previously published studies, it has been shown that telmisartan produces a greater reduction in diastolic BP than enalapril and is free from the adverse-effect of dry cough that is commonly encountered with enalapril in patients of essential hypertension.³³⁾ In hypertensive patients, telmisartan was more effective than ACEIs (enalapril, perindopril, ramipril) in BP control and had fewer adverse events than all the included four ACEIs. The ARB telmi-

sartan is more recommendable than the analyzed ACEIs as far as its lowering BP effect is concerned.³⁴⁾ Despite similar BP control between enalapril and telmisartan in hypertensive patients, telmisartan attenuated autonomic balance more effectively than enalapril. But no differences between both drugs in plasma CA were observed.³⁵⁾ Based on the literature included in review, telmisartan and enalapril produced comparable reductions in BP in a broad range of patients with hypertension. Telmisartan appeared to have a better tolerability profile.³⁶⁾ In terms of these findings, losartan (ARB) also seems to be more powerful in the inhibitory effects on adrenal CA secretion compared to enalapril (ACEI).

In the present study, losartan or enalapril also depressed the CA secretory response evoked by BAY-K-8644, which is known to activate L-type voltage-dependent Ca^{2+} channels, in a time-dependent manner.^{21,37)} This result indicates that losartan or enalapril may inhibit Ca^{2+} influx into the rat adrenomedullary cells. In support of this idea, in cultured bovine adrenal medullary cells, nicotinic (but not muscarinic) receptors mediate the Ca^{2+} -dependent secretion of CA.^{38,39)} It has also been known that the activation of nicotinic receptors stimulates the CA secretion by increasing Ca^{2+} entry through receptor-linked and/or voltage-dependent Ca^{2+} channels in both perfused rat adrenal glands^{40,41)} and isolated bovine adrenal chromaffin cells.⁴²⁻⁴⁴⁾ Wada et al.⁴⁵⁾ have found that the adrenomedullary chromaffin cells have 1) nicotinic receptor-associated ionic channels, responsible for carbachol-induced Na^+ influx, 2) voltage-dependent Na^+ channels, responsible for veratridine-induced Na^+ influx, and 3) voltage-dependent Ca^{2+} channels, suggesting that the influx of Na^+ caused either by carbachol or by veratridine leads to activate voltage-dependent Ca^{2+} channels by altering membrane potentials, whereas high K^+ directly activates voltage-dependent Ca^{2+} channels without increas-

ing Na^+ influx. In the present study, the finding that high K^+ -induced CA secretory response was depressed by pretreatment with losartan or enalapril indicates that this inhibitory effect of losartan or enalapril is exerted by inhibiting Ca^{2+} influx into the adrenomedullary cells through the blockade of voltage-dependent Ca^{2+} channels. Furthermore, slight elevation in the extracellular potassium concentration increases both the frequency of spontaneous action potentials and the secretion of CA,⁴⁶⁾ suggesting that the influx of Ca^{2+} that occurs during action potentials is directly linked to the rate of secretion. These findings that losartan or enalapril inhibited the CA secretion evoked by BAY-K-8644 as well as by high K^+ suggest that losartan or enalapril directly inhibits the voltage-dependent Ca^{2+} channels. But losartan also elicited more potent inhibitory effects on the CA release evoked by BAY-K-8644 and high potassium than that of enalapril. In accordance with these findings, in the bovine chromaffin cells, stimulation of nicotinic, but not muscarinic ACh receptors is known to cause CA secretion by increasing Ca^{2+} influx largely through voltage-dependent Ca^{2+} channels.^{47,48)} Therefore, the finding that losartan or enalapril inhibited the ACh- and DMPP-evoked CA secretion seems to be associated with inhibition of Ca^{2+} influx through voltage-dependent Ca^{2+} channels. Losartan also more potently inhibited the CA secretion evoked by DMPP and ACh compared to that of enalapril.

The present study has also shown that losartan or enalapril also inhibits the CA secretion evoked by cyclopiazonic acid. Cyclopiazonic acid is known to be a highly selective inhibitor of Ca^{2+} -ATPase in skeletal muscle sarcoplasmic reticulum^{23,24)} and a valuable pharmacological tool for investigating intracellular Ca^{2+} mobilization and ionic currents regulated by intracellular Ca^{2+} .⁴⁹⁾ It has been shown that Ca^{2+} -uptake into intracellular storage sites susceptible to caffeine⁵⁰⁾ is almost

completely abolished by treatment with cyclopiazonic acid during the proceeding of Ca^{2+} load.⁴⁹⁾ This is consistent with the findings obtained in exposed smooth muscle fibers of the longitudinal layer of the guinea-pig ileum, where Ca^{2+} -uptake was also inhibited by cyclopiazonic acid.⁵¹⁾ Suzuki et al.⁴⁹⁾ have shown that cyclopiazonic acid easily penetrates into the cytoplasm through the plasma membrane and reduces Ca^{2+} -ATPase activity in sarcoplasmic/endoplasmic reticulum, resulting in increase in the subsequent Ca^{2+} release from those storage sites. Moreover, in bovine adrenal chromaffin cells, stimulation of muscarinic ACh receptors is also proposed to cause activation of phosphoinositide metabolism, resulting in the formation of inositol 1,4,5-trisphosphate, which induces the mobilization of Ca^{2+} from the intracellular pools.^{52,53)} Therefore, in the present work, it can be speculated that the inhibitory effect of losartan or enalapril on CA secretion evoked by McN-A-343 may be associated with the mobilization of intracellular Ca^{2+} from the cytoplasmic calcium store. This indicates that the losartan or enalapril has an inhibitory effect on the release of Ca^{2+} from the intracellular pools induced by stimulation of muscarinic ACh receptors, which is weakly responsible for the secretion of CA. The present results suggest that losartan or enalapril-induced inhibition of the CA secretion evoked by McN-A-343 and cyclopiazonic acid may be due to the inhibition of Ca^{2+} release evoked by stimulation of muscarinic ACh receptors from the intracellular pools. However, in the present study, it is uncertain whether the inhibitory effect of losartan or enalapril on Ca^{2+} movement from intracellular pools is due to its direct effect on the phosphatidyl inositol response or the indirect effects. Further studies are needed to investigate the true nature of these findings.

In the present study, when both two drugs were combined, their inhibitory effect on ACh-evoked CA release

was elevated. In support of this idea, the losartan-enalapril combination is more effective in decreasing blood pressure and increasing plasma active renin than doubling of the enalapril dose in normotensive male volunteers.⁵⁴⁾ Since synergy was observed only after combining low doses of ramipril and candesartan-cilexetil in spontaneously hypertensive rats, prospective clinical trials should be performed on a low-dose combination, revealing the antihypertensive/antiproliferative benefits.⁵⁵⁾ Moreover, some recent studies have reported that the combination of two agents, ACEi and ARB, that inhibit two consecutive steps, promotes a decrease in myocardial fibrosis and left ventricular hypertrophy.⁵⁶⁻⁵⁹⁾ In addition, experimental and clinical studies indicate that the combination of low doses of ACEi and ARB have a synergic and most effectiveness on left ventricular hypertrophy.^{55,60)} Based on these results, the clinically combined use of both losartan and enalapril may contribute greatly to the improvement of cardiovascular diseases such as hypertension, heart failure and angina pectoris.

In conclusion, the results of the present study have demonstrated that both losartan and enalapril inhibits the CA secretion by stimulation of cholinergic nicotinic receptors as well as by membrane depolarization in the perfused rat adrenal medulla. It is thought that this inhibitory effect of losartan or enalapril is exerted by blocking influx of both ions through Ca^{2+} and Na^{+} channels into the adrenomedullary cells as well as by blocking the release of Ca^{2+} from the cytoplasmic calcium store, which are at least partly due to the blockade of local modulation by rennin-Ang system. Based on the same concentration used in the present study, it seems that losartan-induced inhibitory effect on CA release is more potent than enalapril-induced effect. The combined use of both two drugs may contribute greatly to the improvement of cardiovascular diseases such as hypertension,

heart failure and angina pectoris, through inhibition of the CA secretion from adrenomedullary chromaffin cells and consequent reduction of the CA level in the circulation.

Summary

목적: 본 연구는 흰쥐부신의 관류모델에서 카테콜아민(catecholamines, CA) 분비에 대한 안지오텐신 II (angiotensin II, Ang II) 전환효소억제제인 enalapril과 안지오텐신 II 수용체 차단제인 losartan의 억제작용을 비교하는데 목적이 있다.

방법: 부신을 적출하여 Krebs-bicarbonate 액으로 관류하였으며 CA는 형광분석기를 이용하여 직접 측정하였다.

결과: Enalapril과 losartan은 다 같이 부신평맥 내로 90 분간 관류하는 동안 아세틸콜린(acetylcholine, ACh), 선택성 N_n 수용체 작용제(1.1-dimethyl-4-phenyl piperazinium, DMPP), 고칼륨(직접 막탈분극제), 선택성 M_1 수용체 작용제(3-(m-chloro-phenyl-carbamoyl-oxy-2-butynyl-trimethyl ammonium, McN-A-343) 및 Ang II에 의한 CA분비작용을 시간의존적으로 억제하였다. 또한 enalapril이나 losartan 존재상태에서 veratridine(전압의존성 Na 통로 활성화제), L형 칼슘통로활성화제(6-dimethyl-3-nitro-4-(2-trifluoromethyl-phenyl)-pyridine-5-carboxylate, BAY-K-8644), cyclopiazonic acid(세포질내형질세망 Ca^{2+} ATPase 억제제)에 의한 CA분비가 유의하게 억제되었다. Enalapril과 losartan의 동일 농도를 근거로 보면, ACh, 고칼륨, DMPP, McN-A-343, Ang II, veratridine, BAY-K-8644 및 cyclopiazonic acid의 CA분비에 대한 억제작용을 비교하면 losartan > enalapril이다. Enalapril과 losartan 동시 존재상태에서 ACh의 CA분비작용은 enalapril이나 losartan 단독 처치의 경우에 비교하여 현저히 억제되었다.

결론: 위의 결과를 종합하여보면, enalapril과 losartan은 흰쥐관류 부신평맥에서 콜린수용체 및 AngII수용체(Ang II type-1) 흥분에 의한 CA분비를 다 같이 억제하며, 두 약물은 병용시 억제효과가 상승되었으며 이는 임상적으로 유용하다고 생각된다. 본 연구에서 사용한 농도를 근

거로 보면 CA분비에 대한 losartan의 억제작용이 enalapril보다 더욱 강력한 것으로 생각된다.

Conflict of interest

No potential conflict of interest relevant to this article was as reported.

Acknowledgements

This study was supported partly by Academic-Industrial Cooperative Research Fund of Korean Society of Hypertension (2010).

References

- Douglas WW, Poisner AM. Stimulation of uptake of calcium-45 in the adrenal gland by acetylcholine. *Nature*. 1961;192:1299.
- Wilson SP, Kirshner N. The acetylcholine receptor of the adrenal medulla. *J Neurochem*. 1977;28:687-95.
- Holz RW, Senter RA, Frye RA. Relationship between Ca^{2+} uptake and catecholamine secretion in primary dissociated cultures of adrenal medulla. *J Neurochem*. 1982;39:635-46.
- Gavras I, Gavras H. Angiotensin II-possible adverse effects on arteries, heart, brain, and kidney: experimental, clinical, and epidemiological evidence. In: Robertson JI, Nicholls MG, editors. *The renin-angiotensin system*. London: Gower Medical Publishing, 1993: 40.
- Nap A, Balt JC, Mathy MJ, Van Zwieten PA. AT(1)-receptor blockade and sympathetic neurotransmission in cardiovascular disease. *Auton Autacoid Pharmacol*. 2003;23: 285-96.
- Feldberg W, Lewis GP. The action of peptides on the adrenal medulla: release of adrenaline by bradykinin and angiotensin. *J Physiol*. 1964;171:98-108.
- Chulak C, Couture R, Foucart S. Modulatory effect of bradykinin on the release of noradrenaline from rat isolated atria. *Br J Pharmacol*. 1995;115:330-4.
- Rump LC, Berlit T, Schwertfeger E, Beyersdorf F, Schollmeyer P, Bohmann C. Angiotensin converting enzyme inhibition unmasks the sympathofacilitatory effect of bradykinin in human right atrium. *J Hypertens*. 1997;15:1263-70.
- Starke K, Peskar BA, Schumacher KA, Taube HD. Bradykinin and postganglionic sympathetic transmission. *Naunyn Schmiedeberg's Arch Pharmacol*. 1977;299:23-32.
- Franchi F, Lazzeri C, Foschi M, Tosti-Guerra C, Barletta G. Cardiac autonomic tone during trandolapril-irbesartan low-dose combined therapy in hypertension: a pilot project. *J Hum Hypertens*. 2002;16:597-604.
- Karas M, Lacourciere Y, LeBlanc AR, Nadeau R, Dube B, Florescu M, et al. Effect of the renin-angiotensin system or calcium channel blockade on the circadian variation of heart rate variability, blood pressure and circulating catecholamines in hypertensive patients. *J Hypertens*. 2005;23: 1251-60.
- Sakata K, Yoshida H, Obayashi K, Ishikawa J, Tamekiyo H, Nawada R, et al. Effects of losartan and its combination with quinapril on the cardiac sympathetic nervous system and neurohormonal status in essential hypertension. *J Hypertens*. 2002;20:103-10.
- Balt JC, Mathy MJ, Pfaffendorf M, van Zwieten PA. Inhibition of angiotensin II-induced facilitation of sympathetic neurotransmission in the pithed rat: a comparison between losartan, irbesartan, telmisartan, and captopril. *J Hypertens*. 2001;19:465-73.
- Randa HD. Renin and angiotensin. In: Brunton LL, Bruce AC, Bjorn CK, editors. *Goodman and Gilman's the pharmacological basis of therapeutics*. 12th ed. New York: McGraw-Hill, Health Publishing Division, 2011: 721-44.
- Ball SG, White WB. Debate: angiotensin-converting enzyme inhibitors versus angiotensin II receptor blockers--a gap in evidence-based medicine. *Am J Cardiol*. 2003;91(10A): 15G-21G.
- Wakade AR. Studies on secretion of catecholamines evoked by acetylcholine or transmural stimulation of the rat adrenal gland. *J Physiol*. 1981;313:463-80.
- Anton AH, Sayre DF. A study of the factors affecting the aluminum oxide-trihydroxyindole procedure for the analysis of catecholamines. *J Pharmacol Exp Ther*. 1962;138:360-75.
- Tallarida RJ, Murray RB. *Manual of pharmacologic calculation with computer programs*. 2nd ed. New York: Springer-Verlag, 1987: 110-20, 131-6.
- Hammer R, Giachetti A. Muscarinic receptor subtypes: M1 and M2 biochemical and functional characterization. *Life Sci*. 1982;31:2991-8.

20. Hano T, Mizukoshi M, Baba A, Nakamura N, Nishio I. Angiotensin II subtype 1 receptor modulates epinephrine release from isolated rat adrenal gland. *Blood Press Suppl.* 1994;5:105-8.
21. Garcia AG, Sala F, Reig JA, Viniegra S, Frias J, Fonteriz R, et al. Dihydropyridine BAY-K-8644 activates chromaffin cell calcium channels. *Nature.* 1984;309:69-71.
22. Lim DY, Kim CD, Ahn KW. Influence of TMB-8 on secretion of catecholamines from the perfused rat adrenal glands. *Arch Pharm Res.* 1992;15:115-25.
23. Goeger DE, Riley RT. Interaction of cyclopiazonic acid with rat skeletal muscle sarcoplasmic reticulum vesicles. Effect on Ca^{2+} binding and Ca^{2+} permeability. *Biochem Pharmacol.* 1989;38:3995-4003.
24. Seidler NW, Jona I, Vegh M, Martonosi A. Cyclopiazonic acid is a specific inhibitor of the Ca^{2+} -ATPase of sarcoplasmic reticulum. *J Biol Chem.* 1989;264:17816-23.
25. Wada Y, Satoh K, Taira N. Cardiovascular profile of Bay K 8644, a presumed calcium channel activator, in the dog. *Naunyn Schmiedeberg's Arch Pharmacol.* 1985;328:382-7.
26. Martineau D, Yamaguchi N, Briand R. Inhibition by BMS 186295, a selective nonpeptide AT_1 antagonist, of adrenal catecholamine release induced by angiotensin II in the dog in vivo. *Can J Physiol Pharmacol.* 1995;73:459-64.
27. Martineau D, Briand R, Yamaguchi N. Functional evidence for L-type Ca^{2+} channels controlling ANG II-induced adrenal catecholamine release in vivo. *Am J Physiol.* 1996;271(6 Pt 2):R1713-9.
28. Martineau D, Lamouche S, Briand R, Yamaguchi N. Functional involvement of angiotensin AT_2 receptor in adrenal catecholamine secretion in vivo. *Can J Physiol Pharmacol.* 1999;77:367-74.
29. Yamaguchi N, Martineau D, Lamouche S, Briand R. Functional role of local angiotensin-converting enzyme (ACE) in adrenal catecholamine secretion in vivo. *Can J Physiol Pharmacol.* 1999;77:878-85.
30. Cavadas C, Grand D, Mosimann F, Cotrim MD, Fontes Ribeiro CA, Brunner HR, et al. Angiotensin II mediates catecholamine and neuropeptide Y secretion in human adrenal chromaffin cells through the AT_1 receptor. *Regul Pept.* 2003;111:61-5.
31. Critchley L, Ding B, Fok B, Wang D, Tomlinson B, James A, et al. The effects of candesartan and ramipril on adrenal catecholamine release in anaesthetized dogs. *Eur J Pharmacol.* 2004;489:67-75.
32. Koji T, Onishi K, Dohi K, Okamoto R, Tanabe M, Kitamura T, et al. Addition of angiotensin II receptor antagonist to an ACE inhibitor in heart failure improves cardiovascular function by a bradykinin-mediated mechanism. *J Cardiovasc Pharmacol.* 2003;41:632-9.
33. Akat PB, Bapat TR, Murthy MB, Karande VB, Burute SR. Comparison of the efficacy and tolerability of telmisartan and enalapril in patients of mild to moderate essential hypertension. *Indian J Pharmacol.* 2010;42:153-6.
34. Zou Z, Xi GL, Yuan HB, Zhu QF, Shi XY. Telmisartan versus angiotensin-converting enzyme inhibitors in the treatment of hypertension: a meta-analysis of randomized controlled trials. *J Hum Hypertens.* 2009;23:339-49.
35. Lewandowski J, Abramczyk P, Dobosiewicz A, Bidiuk J, Sinski M, Gaciong Z. The effect of enalapril and telmisartan on clinical and biochemical indices of sympathetic activity in hypertensive patients. *Clin Exp Hypertens.* 2008;30:423-32.
36. Smith DH. Treatment of hypertension with an angiotensin II-receptor antagonist compared with an angiotensin-converting enzyme inhibitor: a review of clinical studies of telmisartan and enalapril. *Clin Ther.* 2002;24:1484-501.
37. Schramm M, Thomas G, Towart R, Franckowiak G. Novel dihydropyridines with positive inotropic action through activation of Ca^{2+} channels. *Nature.* 1983;303:535-7.
38. Fisher SK, Holz RW, Agranoff BW. Muscarinic receptors in chromaffin cell cultures mediate enhanced phospholipid labeling but not catecholamine secretion. *J Neurochem.* 1981;37:491-7.
39. Yanagihara N, Isosaki M, Ohuchi T, Oka M. Muscarinic receptor-mediated increase in cyclic GMP level in isolated bovine adrenal medullary cells. *FEBS Lett.* 1979;105:296-8.
40. Wakade AR, Wakade TD. Contribution of nicotinic and muscarinic receptors in the secretion of catecholamines evoked by endogenous and exogenous acetylcholine. *Neuroscience.* 1983;10:973-8.
41. Lim DY, Hwang DH. Studies on secretion of catecholamines evoked by DMPP and McN-A-343 in the rat adrenal gland. *Korean J Pharmacol.* 1991;27:53-67.
42. Kilpatrick DL, Slepatis RJ, Corcoran JJ, Kirshner N. Calcium uptake and catecholamine secretion by cultured bovine adrenal medulla cells. *J Neurochem.* 1982;38:427-35.

43. Kilpatrick DL, Slepetis RJ, Corcoran JJ, Kirshner N. Calcium uptake and catecholamine secretion by cultured bovine adrenal medulla cells. *J Neurochem.* 1982;38:427-35.
44. Knight DE, Kesteven NT. Evoked transient intracellular free Ca^{2+} changes and secretion in isolated bovine adrenal medullary cells. *Proc R Soc Lond B Biol Sci.* 1983;218:177-99.
45. Wada A, Takara H, Izumi F, Kobayashi H, Yanagihara N. Influx of ^{22}Na through acetylcholine receptor-associated Na channels: relationship between ^{22}Na influx, ^{45}Ca influx and secretion of catecholamines in cultured bovine adrenal medulla cells. *Neuroscience.* 1985;15:283-92.
46. Kidokoro Y, Ritchie AK. Chromaffin cell action potentials and their possible role in adrenaline secretion from rat adrenal medulla. *J Physiol.* 1980;307:199-216.
47. Burgoyne RD. Mechanisms of secretion from adrenal chromaffin cells. *Biochim Biophys Acta.* 1984;779:201-16.
48. Oka M, Isosaki M, Yanagihara N. Isolated bovine adrenal medullary cells: studies on regulation of catecholamine synthesis and release. In: Usdin E, Kopin IJ, editors. *Catecholamines: basic and clinical frontiers.* Oxford: Pergamon Press, 1979:70-2.
49. Suzuki M, Muraki K, Imaizumi Y, Watanabe M. Cyclopiazonic acid, an inhibitor of the sarcoplasmic reticulum Ca^{2+} -pump, reduces Ca^{2+} -dependent K^{+} currents in guinea-pig smooth muscle cells. *Br J Pharmacol.* 1992;107:134-40.
50. Iino M. Calcium-induced calcium release mechanism in guinea pig taenia caeci. *J Gen Physiol.* 1989;94:363-83.
51. Uyama Y, Imaizumi Y, Watanabe M. Effects of cyclopiazonic acid, a novel Ca^{2+} -ATPase inhibitor, on contractile responses in skinned ileal smooth muscle. *Br J Pharmacol.* 1992;106:208-14.
52. Cheek TR, O'Sullivan AJ, Moreton RB, Berridge MJ, Burgoyne RD. Spatial localization of the stimulus-induced rise in cytosolic Ca^{2+} in bovine adrenal chromaffin cells: distinct nicotinic and muscarinic patterns. *FEBS Lett.* 1989;247:429-34.
53. Challis RA, Jones JA, Owen PJ, Boarder MR. Changes in inositol 1,4,5-trisphosphate and inositol 1,3,4,5-tetrakisphosphate mass accumulations in cultured adrenal chromaffin cells in response to bradykinin and histamine. *J Neurochem.* 1991;56:1083-6.
54. Azizi M, Guyene TT, Chatellier G, Wargon M, Menard J. Additive effects of losartan and enalapril on blood pressure and plasma active renin. *Hypertension.* 1997;29:634-40.
55. Raasch W, Jöhren O, Schwartz S, Gieselberg A, Dominiak P. Combined blockade of AT_1 -receptors and ACE synergistically potentiates antihypertensive effects in SHR. *J Hypertens.* 2004;22:611-8.
56. Klingbeil AU, Schneider M, Martus P, Messerli FH, Schmieder RE. A meta-analysis of the effects of treatment on left ventricular mass in essential hypertension. *Am J Med.* 2003;115:41-6.
57. Brilla CG, Funck RC, Rupp H. Lisinopril-mediated regression of myocardial fibrosis in patients with hypertensive heart disease. *Circulation.* 2000;102:1388-93.
58. Diez J, Querejeta R, Lopez B, Gonzalez A, Larman M, Martinez Ubago JL. Losartan-dependent regression of myocardial fibrosis is associated with reduction of left ventricular chamber stiffness in hypertensive patients. *Circulation.* 2002;105:2512-7.
59. Devereux RB, Dahlof B, Gerds E, Boman K, Nieminen MS, Papademetriou V, et al. Regression of hypertensive left ventricular hypertrophy by losartan compared with atenolol: the Losartan Intervention for Endpoint Reduction in Hypertension (LIFE) trial. *Circulation.* 2004;110:1456-62.
60. Suzuki H, Kanno Y, Kaneko K, Kaneko M, Kotaki S, Mimura T, et al. Comparison of the effects of angiotensin receptor antagonist, angiotensin converting enzyme inhibitor, and their combination on regression of left ventricular hypertrophy of diabetes type 2 patients on recent onset hemodialysis therapy. *Ther Apher Dial.* 2004;8:320-7.